

## Research Based Affirmation of Medicinal Plants Possessing Analgesic Activity

ASSISTANT PROFESSOR, PAL RUCHI  
MAA BHAGWATI COLLEGE OF PHARMACY,LUCKNOW

Date of Submission: 27-05-2023

Date of Acceptance: 08-06-2023

### ABSTRACT:

The current communication provides a more comprehensive overview of herbs that have analgesic and anti-inflammatory properties, with a focus on plants found in various parts of the globe. As per the World Health Organization, nearly 80% of the population in underdeveloped nations cannot afford synthetic pharmaceuticals and must rely on traditional medicines, mostly of plant origin, to meet their basic health care requirements. Plants have been used in various disorders such as gastrointestinal disorders, genitourinary problems, pain discomforts, and psychological and respiratory problems since the dawn of time, and people in Western countries are now returning to herbal

medicines due to their extensive biological and medicinal activities, higher safety, and lower cost. Many plants have been used in the treatment of pain for centuries, and their anti-inflammatory properties have been scientifically proven. The value of medicinal plants in the management of pain is clearly demonstrated in the current review. This article will benefit both the general public and researchers in their efforts to isolate and characterize the active chemical ingredients responsible for analgesic as well as anti-inflammatory activity.

**Keywords:** Analgesic, Herbal plants, Folk Medicine.



### I. INTRODUCTION:

Natural compounds produced from plants have played an important role in medicine development and will continue to do so. Herbs are gaining in popularity and re-establishing themselves as medicines all over the world. In comparison to synthetic components, which are considered harmful to humans, herbal components currently represent protection. Although herbs have been admired for their medicinal properties for decades, synthetic drugs of the modern era have overshadowed their importance. Human beings are switching to natural components in search of protection and security, while chemical components are being questioned [1].

As per the World Health Organization, about 80% of current medicines are derived from herbs used in herbal medicine. The majority of them

are chemical analogs created as proto-type items. In the coming days, the vital use of medicinal plants in therapeutics will proceed. The increasing awareness and use of biologically active items derived from plants in the pharmaceutical industry, and also raising public costs in the regular maintenance of a person's health or well, has indeed been regarded [2].

Plants were the source of remedy and prophylaxis in ancient times. Even so, the decreasing effectiveness of synthetic drugs, as well as the growing number of contraindications to their use, has reintroduced the use of conventional medicine [2, 3].

On the basis of our interest in analgesic drugs, a screening program of some Iranian medicinal plants for analgesic activity has been started. This evaluation was based on the traditional

use in Iranian medicine and elsewhere, according to other similar selections of plants for research. Natural products are believed to be an important source of new chemical substance with potential therapeutic applicability. Several plant species are traditionally used as analgesics [4].

#### PATHOGENESIS OF ANALGESIC:

Arachidonic acid is derived from phospholipids (lipids of a membrane). Metabolism of arachidonic acid is done by prostaglandin synthases (PGA<sub>2</sub>), which, through its cyclooxygenase (COX) and endoperoxides actions, outcomes in the process and production of PGG<sub>2</sub> to PGH<sub>2</sub>. Prostacyclins (PGI<sub>2</sub>), PGD<sub>2</sub>, PGE<sub>2</sub>, PGF<sub>2</sub>, and TXA<sub>2</sub> are derived from PGH<sub>2</sub> by prostaglandin synthases.

PGE<sub>2</sub> is chiefly present in the Brain, Kidney, Vascular smooth muscle, and Platelets. PGE<sub>2</sub> is proceeding as a vasodilator by reacting on vascular smooth muscle to cause blood vessel dilation. This result is increased blood circulation. This includes high blood pressure and headache pain [5, 6, 7, 8].

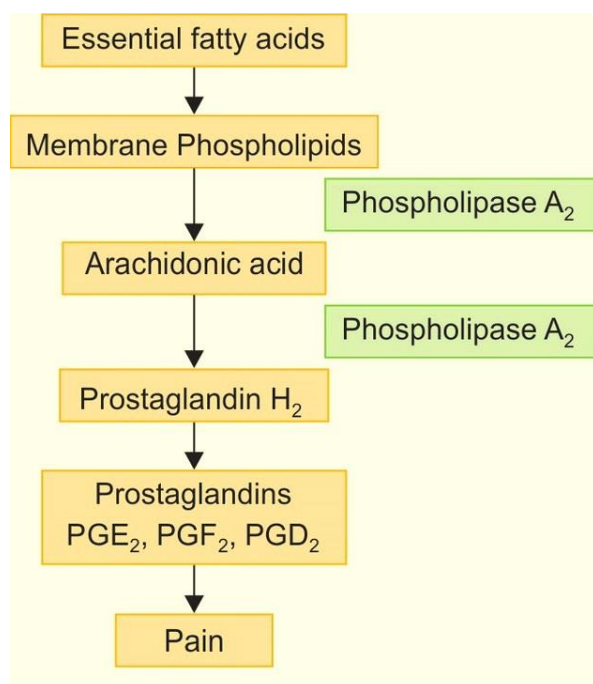


Fig. 1 - Pathogenesis of Analgesic [5, 6, 7, 8].

#### 1) Devil's Claw (*Harpogophytum procumbens* DeCandolle, Pealiaceae) :

*Harpogophytum procumbens* DeCandolle (Pealiaceae) has brilliant pinkish-red blooms with

many tiny hooks. It's also known as 'Devil's claw,' 'Windhoek's root,' wood spider,' hook plant,' and 'grapple plant' in the area. It is regarded as a useful anti-inflammatory and analgesic medicine in Europe and abroad for the treatment of low back pain. Doctors in Europe use Devil's claw extract injections to treat illnesses like joint pain, arthritis, low back pain, and knee discomfort. The principal folkloric applications and relevant sources on Devil's claw's therapeutic benefits are presented in a tabular format for convenient reference [9].

Fever, diabetic, and blood sickness are some of the ethno - medicinal usage of *H. procumbens*; however there are few recorded historical records. Extracts from the secondary tubers of *H. procumbens* have been shown to be useful in the treatment of degenerative rheumatoid arthritis, osteoarthritis, tendonitis, renal inflammation, dyspepsia, and lack of appetite in recent scientific investigations. [10].The swelling of arthritic joints in rats with chemically arthritis was reduced significantly by *H. procumbens*. *H. procumbens* was shown to have a powerful anti-inflammatory or anti-rheumatic chemical, and further experiments were conducted. The results were encouraging, but the entire plant extract performed better [11].

The ability of *H. procumbens* to inhibit the synthesis of inflammatory mediators like PGE<sub>2</sub> is thought to explain its efficacy in lowering pain and inflammation associated with rheumatoid arthritis and osteoarthritis [12].

The antagonistic activity of an ethanol extract of *H. procumbens* tubers and its primary major compounds, harpagide, harpagoside, 8-p-coumaroylharpagide, and acteoside, on COX-2 expression, was found [13]. In rats, the aqueous secondary tuber extract of *H. procumbens* (50–800 mg/kg) provided strong analgesic effects against nociceptive pain stimuli elicited thermally (hot-plate) and chemically (acetic acid) [14].

Mice were given 400mg/kg aqueous extract of *H. procumbens* single dose, which dramatically decreased the number of writhing responses [15]. In the formalin test in mice, *H. procumbens* extract produced substantial antinociceptive activities [16].

## 2) Winter Melon (*Benincasa hispida*, *Cucurbitaceae*):

Winter melon, winter gourd, white pumpkin, ash gourd, ash guard, and wax gourd are all names for *Benincasa hispida* (*Benincasa cerifera*). The cucurbitaceae family includes gourd melon, white gourd, tallow gourd, and Chinese watermelon [17]. Volatile oils, flavonoids, glycosides, saccharides, proteins, minerals,  $\beta$ -sitosterin, carotenes, vitamins, and uronic acid were the main contents of *Benincasa hispida* fruits [18, 19].

The anti-inflammatory effects of *Benincasa hispida* aqueous extract were discovered during early research. In a rat model, petroleum ether and methanolic extract of *Benincasa hispida* fruit at 300 mg/kg inhibited carrageenan-induced paw edema, histamine-induced paw edema, and cotton pellet-induced granuloma in a dose-dependent and substantial manner [20].

In rats, the methanolic extract of *Benincasa hispida* at doses of 250 and 500 mg/kg substantially ( $P < 0.05$ ) improved the antinociceptive efficacy (as measured by an analgesiometer that applies force at a steady pace on the rat paw) in a dosage-dependent way. Similarly, the extract substantially ( $P < 0.05$ ) reduced Brewer's yeast-induced pyrexia in rats at dosages of 250 and 500 mg/kg [21].

## 3) Nata Karanja (*Caesalpinia bonducella*, *Caesalpinaceae*):

Seeds of *Caesalpinia bonducella* have been used in traditional medicine for a long time. It's a large straggling, thorny shrub with hooks and straight, firm yellow prickles on the branches. The leaves have a complex structure. Flowers are pale yellow and grow in supra-axillary racemes at the top of the plant. Nata Karanja (*Caesalpinia bonducella*), a prickly shrub widespread across India's hotter regions belongs to the *Caesalpinaceae* family [22].

Based on the findings of this investigation, it can be determined that the oil of *C. bonducella* seeds has potential anti-acute and anti-chronic effects in a dose-dependent approach (anti-inflammatory, antipyretic, and analgesic activity) As a result, our research provides a fundamental perspective on *C. bonducella* and its health advantages. The edema paw volume was significantly decreased after using the oil. There was a dose-related suppression of hind paw edema between 2 and 4 hours. The reference medicine phenylbutazone (100 mg/kg, orally) had a

strong inhibitory effect equivalent to the *C. bonducella* seed oil examined [23].

Induced writhing in rats by acetic acid When compared to aspirin, 100 mg/kg (66.5%) and control groups, the oil of *C. bonducella* seeds at dosages of 100, 200, and 400 mg/kg demonstrated substantial ( $p < 0.05$ ) suppression of the control writhes at rates of 16.7%, 27.9%, and 48.6%, accordingly. When compared to the control on a hot plate response time in rats, the oil of *C. bonducella* seeds demonstrated substantial ( $p < 0.01$ ) analgesic efficacy at all dosages tested. Furthermore, when oil was combined with conventional medicine (morphine 5 mg/kg) at different dosages (100, 200, 400 mg/kg), the analgesic activity (response time) was potentiated to 21.37, 23.68, and 27.53 minutes, accordingly, compared to 19.17 minutes with morphine (5 mg/kg). [23, 24].

## 4) Wild Jujube (*Zizyphus lotus*, *Rhamnaceae*):

The majority of plant-based medications that have become essential in contemporary medicine have a folkloric basis and are used in traditional medical systems. *Zizyphus* spp. fruit trees are examples of multifunctional plants with a lot of promise for ethnomedicinal usage all around the world. Seven cyclopeptide alkaloids [25, 26] and four dammarane saponins were identified from *Z. lotus* root barks in phytochemical research. The anti-inflammatory and analgesic effects of several root bark extracts are described in this study [27].

When compared to NSAIDs (aspirin (ASA) and piroxicam), the aqueous and methanolic extracts of *Z. lotus* root barks had a substantial anti-inflammatory impact in the acute phase of the inflammation process. When mice were given acetic acid to cause writhing, aqueous extract of root barks (50, 100, and 200 mg/kg) was found to have a substantial analgesic effect when compared to the control group. In addition, the effects of methanolic, ethyl acetate, and chloroformic extracts of *Z. lotus* root barks were studied. The methanolic extract exhibited much higher analgesic effectiveness than chloroform and ethyl acetate extracts, according to the findings [28].

**5) Jerusalem Sage (*Phlomis umbrosa Turcz, Labiatae*) :**

*Phlomis umbrosa Turcz* (Caosu) is a medicinal grass (Labiatae) that grows to be between 40 and 100 cm tall. Antinociceptive and anti-inflammatory properties have been identified in several plants related to *P. umbrosa*. The iridoid glycosides extract of *Lamiophlomis rotata* (400 mg/kg, i.v.) and the ethanol extracts of *Phlomis younghusbandii* (200 mg/kg, i.p.) reduced the number of writhings in mice caused by acetic acid and inhibited the inflammatory production caused by certain reagents (pb<0.01). [29].

The aqueous extract of *P. umbrosa* exhibited strong antinociceptive and anti-inflammatory effects, and indirectly confirmed the local folklore practitioners' traditional usage of *P. umbrosa* in certain inflammatory and pain illnesses. The Aqueous extract of *P. umbrosa* (25, 50, and 100 mg/kg) inhibited the writhing activity elicited by intraperitoneal administration of 0.7 percent acetic acid in animals, according to the findings. Our findings suggested that AEP might increase the animal's reaction speed while also acting as an analgesic against the hot plate [29, 30].

**6) Gular (*Ficus racemosa Linn, Moraceae*) :**

*Ficus racemosa Linn.* (Moraceae) is a well-known plant species in India that has long been utilized in Ayurvedic medicine, India's traditional medical system, to treat a variety of ailments and problems [31]. It is generally known as 'Jagyadumur' (Bengali), 'Gular' (Hindi), and 'Udumbara' (Sanskrit) [32].

The analgesic efficacy of an ethanolic extract of barks and fruits has been documented [33]. The focus of this study was to assess the analgesic efficacy of *F. racemosa* fruits and *S. dulcis* entire herb in mice utilizing two different pain models [34].

When compared to the reference medication diclofenac sodium, the crude extracts of both plants exhibited considerable analgesic effect, although *F. racemosa* was shown to have better analgesic activity than *S. dulcis* against acetic acid produced pain in mice at two dosage levels, 100 and 200 mg/kg [35].

In the examined mice, the crude extracts of both plants had considerable (p0.001) analgesic efficacy at oral doses of 100 and 200 mg/kg. In the hot plate test, *S. dulcis* had a longer latency time than *F. racemosa*, but in the acetic acid-induced writhing test, *F. racemosa* had a lower number of writhes than *S. dulcis* at two dosage levels that were statistically significant (p0.001) when compared to the control [32, 36].

Up to 5000 mg/kg, the extract did not cause death. Ethanol extract at the maximum dose (500 mg/kg) showed comparatively significant (p 0.05) activity in the tail-flick method, significant inhibition of writhes in the writhing test, more significant (p 0.05) response at 90, 120, and 180 minutes in the hot plate method, and comparatively significant (p 0.01) down regulation of paw volume in carrageenan and egg albumin induced paw edema method to that of standard diclofenac [35].

**7) Resam (*Dicranopteris linearis, Gleicheniaceae*) :**

An aqueous extract of *Dicranopteris linearis* leaves was tested in experimental animals to see if it has antinociceptive, anti-inflammatory, and antipyretic qualities. The abdominal constriction, hot plate, and formalin tests were used to assess antinociceptive effect. The carrageenan-induced paw edema and brewer's yeast-induced pyrexia tests were used to determine the anti-inflammatory and antipyretic properties, respectively. The extract was shown to exhibit considerable (P 0.05) concentration-independent antinociceptive, anti-inflammatory, and anti-pyretic action at all dosages tested. Finally, the aqueous extract of *D. linearis* displays antinociceptive, anti-inflammatory, and antipyretic action, corroborating prior assertions of its traditional use by the Malays to cure a number of diseases, including sickness. [37, 38].

**8) Beedi Leaf Tree (*Bauhinia racemosa, Caesalpinaceae*) :**

*Bauhinia racemosa Lam* is a tiny, crooked, thick tree with hanging limbs that can flourish in the harshest climates. This variety may be found all throughout India. The mature leaves of *B. racemosa* are used to make Beedi (Indian cigarettes), whereas the young leaves are utilized by Tamilians as greens (side dish) (Tamil Nadu, India). *B. racemosa* is a sweetish, astringent plant that is used to cure headaches, fevers, skin ailments, blood illnesses, dysentery, and diarrhea. A decoction of the bark is suggested as a good ulcer wash [39, 40].

The anti-inflammatory, analgesic, and antipyretic effects of 50, 100, and 200 mg/kg body weight of methanol extract extracted from *Bauhinia racemosa* stem bark, also known as MEBR, were examined in this study. In carrageenan, dextran, and mediators (histamine and serotonin)-induced paw edema and cotton pellet-induced granuloma, the effects of MEBR on the acute and chronic stages of inflammation were investigated. In acetic acid-induced writhing and hotplate tests, MEBR's analgesic efficacy was assessed. Yeast-induced hyperpyrexia in rats was used to test MEBR's antipyretic efficacy. MEBR's anti-edema impact was compared to 10 mg/kg indomethacin taken orally. After 3 h of treatment with MEBR in carrageenan, dextran, histamine, and serotonin-induced paw edema, a maximal inhibition of 44.9, 43.2, 44.8, and 45.9% ( $P < 0.001$ ) was seen in the acute phase of inflammation in carrageenan, dextran, histamine, and serotonin-induced paw edema, accordingly [41, 42].

#### 9) Gokharu (*T. terrestris* L, *Zygophyllaceae*) :

*T. terrestris* L. (*Zygophyllaceae*) is an annual creeping herb widely growing in Iran. It is also distributed in Japan, Korea, and the western part of Asia, southern Europe, and Africa [43]. *T. terrestris* is extremely rich in substances having potential biological significance, including: saponins, flavonoids, alkaloids, and other nutrients. The quantities and presence of these important metabolites depend on the various parts of the plant used. The fruit and root of *Tribulus terrestris* (Caltrop fruit) contains pharmacologically important metabolites, such as phytosteroids, flavonoids, alkaloids, and glycosides [44, 45, 46].

*Tribulus terrestris* has been used in traditional medicine for relieving rheumatic pain and as an analgesic plant for a long time. In this investigation the analgesic effect of methanolic extract of this plant on male albino mice was evaluated by formalin and tail flick test. Extraction of the fruits of the plant was done by two different methods (suxheletion and percolation) with methanol 80%. The percolated extract was injected intraperitoneally in mice at 50, 100, 200, 400, and 800 mg/kg. The results showed that a dose of 100 mg/kg of percolated extract had the highest significant analgesic effect compared to the control group ( $P < 0.01$ ) in formalin and tail flick test. There is no significant difference in the analgesic effect of suxheleted and percolated extract [47, 48].

The analgesic effects of the extract were lower than morphine, 2.5 mg/kg in both tests, and higher than ASA 300 mg/kg in chronic phase of pain in formalin test ( $P < 0.05$ ). Pretreatment of animal with naloxone did not change the analgesia induced by the plant extract in both tests, therefore the involvement of opioid receptor in the analgesic effect of this plant was excluded. The results of ulcerogenic studies indicate that the gastric ulcerogenicity of plant extract is lower than the indomethacin in the rat's stomach. It can therefore be concluded that *T. terrestris* extract has a suitable analgesic effect and further studies are required to produce a more effective product of this plant to substitute for conventional analgesic drugs [47, 48, 49, 50].

#### 10) Bharangi (*Clerodendron serratum*, *Verbenaceae*) :

In the Ayurveda system of medicine, *C. serratum* (Linn.) Moon (*Verbenaceae*) is known as Bharangi. *Clerodendron* is a significant *Verbenaceae* genus with a large number of species known to be found in India. Flavonoids, diterpenoid, and sterols have already been discovered from the *Clerodendron* genus [51, 52].

The anti-nociceptive, anti-inflammatory, and antipyretic effects of the ethanol roots extract of *C. serratum* at doses of (50, 100, 200 mg/kg.) roots were tested in animal models [53, 54].

#### 11) Ashwagandha (*Withania somnifera*, *Solanaceae*) :

It belongs to the *Solanaceae* family. *Withania somnifera* is an esteemed herb in Ayurvedic medication, and as such was utilized and developed for quite a long time in India (Uttar Pradesh, Madhya Pradesh, Gujarat, Rajasthan, and Punjab). *W. somnifera* have 1.5-m high shrub with ovate leaves and yellowish-green blossoms. The Roots of *W. somnifera* are 20-30 cm long and 6-12 mm. It has a trademark scent, taste severe and horse-like smell [1, 55].

*W. somnifera* has analgesic, anti-inflammatory, and antipyretic [56, 57]. The powder of ashwagandha roots was discovered to have a significant inhibition effect on, as well as other inflammatory markers including interleukin (IL - 6), and tumor necrosis factor (TNF- $\alpha$ ). The extract of roots contains biologically active chemical constituents are Alkaloids (*Withanine*, *withaninine*,

withasomnine, somnine, somniferine, nicotine, isopelletierine, tropeltigloate, anaferine, and somniferinine), Flavonoids (Quercetin, kaempferol), Steroidal Lactones (Withaferin-A, withanone, withanolides), and Steroids (stigmasterol,  $\beta$  - sitosterol, cholesterol, diosgenin, stigmastadien, sitoindosides), and Nitrogen containing Somnitol somnisol, and withanol [55, 59].

It contains are alkaloids, flavonoids, saponins, sugars, and proteins. The seeds powder of *A. aspera* gives calmness from solidness and headache. The leaves and seeds of *Achyranthes aspera* show pain-relieving activity [58, 59].

When active components of *W. somnifera* Withaferin A were compared to the conventional medicine Indomethacin in mice using the Acetic acid-induced writhing method, they revealed analgesic potency at a dose of 30 mg/kg body mass [60].

## 12) Latjira (*Achyranthes aspera*, *Amaranthaceae*) :

It belongs to the *Amaranthaceae* family. It is a yearly, stiff herb, about 0.3 to 0.9m high, and is a typical plant found in badlands, all through roadside and rural fields. It is known by various names in India like Latjira and Chirchira in Hindi, Apamargah, Chirchitaa, and Shikhari in Sanskrit, Chirchitaa in Unani, Pricklychaff flower plant in English, Nayuruvi in Tamil, Uttaraene in Telugu, Kutri in Punjabi, and Kadalad in Malayalam [61, 62].

It contains are alkaloids, flavonoids, saponins, sugars, and proteins. The seeds powder of *A. aspera* gives calmness from solidness and headache. The leaves and seeds of *Achyranthes aspera* show pain-relieving activity. Latjira has been recommended to be a possible lead for another kind of anti-inflammatory agent having a double inhibitory action on phospholipase a (corticosteroid) and cyclooxygenase (COX-1, COX-2) which is responsible for pain and inflammation [63].

Ethanol extract of leaf of *A. aspera* (400 mg/kg) has produced an analgesic effect at 30, 60, 90 and 120min using Tail flick response and Hot plate methods [64].

## II. CONCLUSION:

Herbology is the study of finding healthier and better ways to relieve pain. Several persons seem to recognize the value of pain management for these analgesic plants, which include some of the most well-known and well-liked medicines. Analgesic herbs are used to treat headaches, toothaches, tight muscles, back pain, and nephropathy, among other things. This review paves the path for further investigation into the bioactive components found in these plants, as well as their identification and extraction.

## Conflict of Interest:

We declare that we have no conflict of interest.

## REFERENCE

- [1]. Balkrishna, Acharya, and Laxminarain Misra. "Chemo-botanical and neurological accounts of some ayurvedic plants useful in mental health." *The Natural Products Journal* 8, no. 1 (2018): 14-31.
- [2]. Balkrishna, A., and L. N. Misra. "Ayurvedic plants in brain disorders: the herbal hope." *J. Tradit. Med. Clin. Nat* 6 (2017): 1-9.
- [3]. Prasad, Mrinalini, Jenendra Nath Srivastava, Prem Kumar Dantu, and Rajiv Ranjan. "Medicinal plants of DEI herbal garden, Dayalbagh: A survey." *Journal of Pharmacognosy and Phytochemistry* 8, no. 4 (2019): 06-22.
- [4]. Oketch-Rabah, Hellen A., Robin J. Marles, Scott A. Jordan, and Tieraona Low Dog. "United States Pharmacopeia safety review of willow bark." *Planta medica* 85, no. 16 (2019): 1192-1202
- [5]. Awtry, Eric H., and Joseph Loscalzo. "Aspirin." *Circulation* 101, no. 10 (2000): 1206-1218.
- [6]. Ong JJ, De Felice M. Migraine treatment: current acute medications and their potential mechanisms of action. *Neurotherapeutics*. 2018 Apr 1;15(2):274-90.
- [7]. Nappi, Giuseppe, and Michael A. Moskowitz. "Secondary headaches: introduction." In *Handbook of clinical neurology*, vol. 97, pp. 497-500. Elsevier, 2010.
- [8]. Vane, John. "The evolution of non-steroidal anti-inflammatory drugs and their mechanisms of action." *Drugs* 33, no. 1 (1987): 18-27.
- [9]. Al-Harbi, Naif O., Riyadh M. Al-Ashban, and Arif H. Shah. "Toxicity studies on

- Harpagophytum procumbens (Devils claw) capsules in mice." Journal of Medicinal Plants Research 7, no. 42 (2013): 3089-3097.
- [10]. Stewart, Kristine M., and David Cole. "The commercial harvest of devil's claw (Harpagophytum spp.) in southern Africa: The devil's in the details." Journal of ethnopharmacology 100, no. 3 (2005): 225-236.
- [11]. Eichler, O., Koch, C., 1970. Über die antiphlogistische, analgetische und spasmolytische Wirksamkeit von Harpagosid, einem Glykosid aus der Wurzel von Harpagophytum procumbens DC. Arzneimittel-Forschung 20, 107-109.
- [12]. Aberham, Anita, Stefan Schwaiger, Hermann Stuppner, and Markus Ganzera. "Quantitative analysis of iridoids, secoiridoids, xanthenes and xanthone glycosides in Gentiana lutea L. roots by RP-HPLC and LC-MS." Journal of Pharmaceutical and Biomedical Analysis 45, no. 3 (2007): 437-442.
- [13]. Abdelouahab, Nassima, and Charles Heard. "Effect of the major glycosides of Harpagophytum procumbens (Devil's Claw) on epidermal cyclooxygenase-2 (COX-2) in vitro." Journal of Natural Products 71, no. 5 (2008): 746-749.
- [14]. Mahomed, Ismail M., and John AO Ojewole. "Analgesic, anti-inflammatory and antidiabetic properties of Harpagophytum procumbens DC (Pedaliaceae) secondary root aqueous extract." Phytotherapy research 18, no. 12 (2004): 982-989.
- [15]. Ahmed, Mohamad Ibrahim, Mohamad Ismael Afifi, and Ibrahim Hamdy Younos. "Harpagophytum procumbens (Devil's Claw): A possible natural anti-inflammatory agent (An experimental study)." Iranian Journal of Pharmacology and Therapeutics 4, no. 1 (2005): 54-0.
- [16]. Uchida, Shinya, Keita Hirai, Junya Hatanaka, Junko Hanato, Keizo Umegaki, and Shizuo Yamada. "Antinociceptive effects of St. John's wort, Harpagophytum procumbens extract and Grape seed proanthocyanidins extract in mice." Biological and Pharmaceutical Bulletin 31, no. 2 (2008): 240-245.
- [17]. Zaini, Nurul Aqilah Mohd, Farooq Anwar, Azizah Abdul Hamid, and Nazamid Saari. "Kundur [Benincasa hispida (Thunb.) Cogn.]: A potential source for valuable nutrients and functional foods." Food Research International 44, no. 7 (2011): 2368-2376.
- [18]. WU, CHUNG- MAY, SHU- ER LIOU, YUNG- HO CHANG, and Wenchang Chiang. "Volatile compounds of the wax gourd (Benincasa hispida, Cogn) and a wax gourd beverage." Journal of Food Science 52, no. 1 (1987): 132-134.
- [19]. Yoshizumi, S., T. Murakami, M. Kadoya, H. Matsuda, J. Yamahara, and M. Yoshikawa. "Medicinal foodstuffs. XI. Histamine release inhibitors from wax gourd, the fruits of Benincasa hispida Cogn." Yakugaku zasshi: Journal of the Pharmaceutical Society of Japan 118, no. 5 (1998): 188-192.
- [20]. Rachchh, M. A., P. N. Yadav, R. H. Gokani, and S. M. Jain. "Anti-inflammatory activity of Benincasa hispida fruit." Int J Phar Biosci 2, no. 3 (2011): 98-106.
- [21]. Qadrie ZL, Tayebhawisan N, Alikhan MW, Samuel M and Anandan R. Antinociceptive and anti-pyretic activity of Benincasa hispida (Thunb) Cogn. in Wistar albino rats. Pak J Pharm Sci 2009;22(3):287-290
- [22]. Paranjpe, Prakash. Indian medicinal plants: forgotten healers: a guide to ayurvedic herbal medicine with identity, habitat, botany, photochemistry, ayurvedic properties, formulations & clinical usage. Vol. 26. Chaukhamba Sanskrit Pratishthan, 2001.
- [23]. Shukla, Shruti, Archana Mehta, Pradeep Mehta, Suresh Prasad Vyas, Savita Shukla, and Vivek K. Bajpai. "Studies on anti-inflammatory, antipyretic and analgesic properties of Caesalpinia bonducella F. seed oil in experimental animal models." Food and Chemical Toxicology 48, no. 1 (2010): 61-64.
- [24]. Kannur, Dayanand M., Mukta P. Paranjpe, Lalit V. Sonavane, Prerana P. Dongre, and Kishanchand R. Khandelwal. "Evaluation of Caesalpinia bonduc seed coat extract for anti-inflammatory and analgesic activity." Journal of advanced pharmaceutical technology & research 3, no. 3 (2012): 171.
- [25]. Le Crouéour, Gaëlle, Philippe Thépenier, Bernard Richard, Christian Petermann, Kamel Ghédira, and Monique Zèches-Hanrot. "Lotusine G: a new cyclopeptide alkaloid from Zizyphus lotus." Fitoterapia 73, no. 1 (2002): 63-68.
- [26]. Ghedira, Kamel, Rachid Chemli, Catherine Caron, Jean-Marc Nuzillard, Monique Zèches, and Louisette Le Men-Olivier. "Four

- cyclopeptide alkaloids from *Zizyphus lotus*." *Phytochemistry* 38, no. 3 (1995): 767-772.
- [27]. Renault, Jean-Hugues, Kamel Ghedira, Philippe Thepenier, Catherine Lavaud, Monique Zeches-Hanrot, and Louissette Le Men-Olivier. "Dammarane saponins from *Zizyphus lotus*." *Phytochemistry* 44, no. 7 (1997): 1321-1327.
- [28]. Borgi, W., K. Ghedira, and N. Chouchane. "Antiinflammatory and analgesic activities of *Zizyphus lotus* root barks." *Fitoterapia* 78, no. 1 (2007): 16-19.
- [29]. Liu, Pu, Rui-Xue Deng, Hong-Quan Duan, Wei-Ping Yin, and Tian-Zeng Zhao. "Phenylethanoid glycosides from the roots of *Phlomis umbrosa*." *Journal of Asian natural products research* 11, no. 1 (2009): 69-74.
- [30]. Shang, Xiaofei, Jinhui Wang, Maoxing Li, Xiaolou Miao, Hu Pan, Yaoguang Yang, and Yu Wang. "Antinociceptive and anti-inflammatory activities of *Phlomis umbrosa* Turcz extract." *Fitoterapia* 82, no. 4 (2011): 716-721.
- [31]. Ahmed, Faiyaz, and Asna Urooj. "Traditional uses, medicinal properties, and phytopharmacology of *Ficus racemosa*: A review." *Pharmaceutical biology* 48, no. 6 (2010): 672-681.
- [32]. Zulfiker, A. H. M., M. Mahbubur Rahman, M. Kamal Hossain, K. Hamid, M. E. H. Mazumder, and M. Sohel Rana. "In vivo analgesic activity of ethanolic extracts of two medicinal plants-*Scoparia dulcis* L. and *Ficus racemosa* Linn." *Biol Med* 2, no. 2 (2010): 42-8.
- [33]. Ferdous, Muhshina, Razina Rouf, Jamil Ahmad Shilpi, and Shaikh Jamal Uddin. "Antinociceptive activity of the ethanolic extract of *Ficus racemosa* Lin.(Moraceae)." *Oriental Pharmacy and Experimental Medicine* 8, no. 1 (2008): 93-96.
- [34]. Li, Rachel W., David N. Leach, Stephen P. Myers, Guiliu D. Lin, Gregory J. Leach, and Peter G. Waterman. "A new anti-inflammatory glucoside from *Ficus racemosa* L." *Planta medica* 70, no. 05 (2004): 421-426.
- [35]. Harer Sunil, L., and S. Harer Priyanka. "Evaluation of analgesic and anti-inflammatory activity of *Ficus racemosa* Linn. stem bark extract in rats and mice." *Pharmacognosy Journal* 2, no. 6 (2010).
- [36]. Mandal, Subhash C., Tapan K. Maity, J. Das, B. P. Saba, and M. Pal. "Anti-inflammatory evaluation of *Ficus racemosa* Linn. leaf extract." *Journal of ethnopharmacology* 72, no. 1-2 (2000): 87-92.
- [37]. Zakaria, Zainul Amiruddin, Zuleen Delina Fasya Abdul Ghani, Raden Nur Suraya Raden Mohd Nor, Hanan Kumar Gopalan, Mohd Roslan Sulaiman, Abdul Manan Mat Jais, Muhammad Nazrul Somchit, Arifah Abdul Kader, and Johari Ripin. "Antinociceptive, anti-inflammatory, and antipyretic properties of an aqueous extract of *Dicranopteris linearis* leaves in experimental animal models." *Journal of natural medicines* 62, no. 2 (2008): 179-187.
- [38]. Zakaria, Zainul Amiruddin, Zuleen Delina Fasya Abdul Ghani, Raden Nur Suraya Raden Mohd Nor, Hanan Kumar Gopalan, Mohd Roslan Sulaiman, and Fatimah Corazon Abdullah. "Antinociceptive and anti-inflammatory activities of *Dicranopteris linearis* leaves chloroform extract in experimental animals." *Yakugaku Zasshi* 126, no. 11 (2006): 1197-1203.
- [39]. Panda, Pritipadma, D. Das, Priyanka Dash, and Goutam Ghosh. "Therapeutic potential of *Bauhinia racemosa*—a mini review." *Int J Pharm Sci Rev Res* 32, no. 2 (2015): 169-179.
- [40]. Prabhu, S., S. Vijayakumar, Raju Ramasubbu, P. K. Praseetha, K. Karthikeyan, G. Thiyagarajan, J. Sureshkumar, and N. Prakash. "Traditional uses, phytochemistry and pharmacology of *Bauhinia racemosa* Lam.: a comprehensive review." *Future Journal of Pharmaceutical Sciences* 7, no. 1 (2021): 1-18.
- [41]. Gupta, M., U. K. Mazumder, R. Sambath Kumar, P. Gomathi, Y. Rajeshwar, B. B. Kakoti, and V. Tamil Selven. "Anti-inflammatory, analgesic and antipyretic effects of methanol extract from *Bauhinia racemosa* stem bark in animal models." *Journal of ethnopharmacology* 98, no. 3 (2005): 267-273.
- [42]. Borikar, V. I., C. R. Jangde, D. S. Rekhe, and Preety Philip. "Study of Analgesic activity of *Bauhinia racemosa* lam in Rats." *Veterinary World* 2, no. 4 (2009): 135.
- [43]. Sharifi, Ali M., Radbod Darabi, and Nasrin Akbarloo. "Study of antihypertensive mechanism of *Tribulus terrestris* in 2K1C hypertensive rats: role of tissue ACE



- activity." Life sciences 73, no. 23 (2003): 2963-2971.
- [44]. Iranian Herbal Pharmacopoeia (IHP). 2002. Vol 2. 620–624. Ministry of Health Publication. Tehran.
- [45]. Heidari, Mahmoud Reza, Elham Moein Azad, and Mitra Mehrabani. "Evaluation of the analgesic effect of *Echium amoenum* Fisch & CA Mey. extract in mice: possible mechanism involved." Journal of ethnopharmacology 103, no. 3 (2006): 345-349.
- [46]. MANDGARY, ALI, and MOHSEN ENAYATI. "Antinociceptive effects and toxicity of *Fumaria parviflora* lam. in mice and rats." DARU Journal of Pharmaceutical Sciences 12, no. 4 (2004): 136-140.
- [47]. Zargari, A. "Medicinal Plants, Vol. 1. 450–451." Tehran University Publication. Tehran (1996).
- [48]. Wu, Gong, Shanhao Jiang, Fuxiang Jiang, Dayuan Zhu, Houming Wu, and Shaokai Jiang. "Steroidal glycosides from *Tribulus terrestris*." Phytochemistry 42, no. 6 (1996): 1677-1681.
- [49]. Wang, Yan, Kazuhiro Ohtani, Ryoji Kasai, and Kazuo Yamasaki. "Steroidal saponins from fruits of *Tribulus terrestris*." Phytochemistry 45, no. 4 (1997): 811-817.
- [50]. Heidari, M. R., M. Mehrabani, A. Pardakhty, P. Khazaeli, M. J. Zahedi, M. Yakhchali, and MJAotNYAoS Vahedian. "The analgesic effect of *Tribulus terrestris* extract and comparison of gastric ulcerogenicity of the extract with indomethacine in animal experiments." Annals of the New York Academy of Sciences 1095, no. 1 (2007): 418-427.
- [51]. Ravikumar, Raju, Akoni Joghee Lakshmanan, and Subban Ravi. "Chemical constituents from *Clerodendron serratum*." Journal of Asian natural products research 10, no. 7 (2008): 652-655.
- [52]. Vendantham, T. N. C., S. Sankara Subramanian, and J. B. Harborne. "4-methylscutellarein and pectolinarigenin from *Clerodendron inerme*." Phytochemistry (1977).
- [53]. Patel, Jagruti J., Sanjeev R. Acharya, and Niyati S. Acharya. "Clerodendron serratum (L.) Moon.—A review on traditional uses, phytochemistry and pharmacological activities." Journal of ethnopharmacology 154, no. 2 (2014): 268-285.
- [54]. Narayanan, N., P. Thirugnanasambantham, S. Viswanathan, V. Vijayasekaran, and E. Sukumar. "Antinociceptive, anti-inflammatory and antipyretic effects of ethanol extract of *Clerodendron serratum* roots in experimental animals." Journal of Ethnopharmacology 65, no. 3 (1999): 237-241.
- [55]. Baveesh, Pudhuvai, and D. P. Vani. "World's worst perennial weed as medicinal ailment." Trends in Biosciences 10, no. 15 (2017): 2607-2611. [49]
- [56]. Rehman, Rafia, Dalel Melki, Aamir Shehzad, Farwa Nadeem, and Talha Khalid. "Commercial Importance, Medicinal Value and Therapeutic Potentials of Chaff Flower (*Achyranthes aspera*)—A Review." International Journal of Chemical and Biochemical Sciences 14 (2018): 62-70.
- [57]. Kasahara, Yoshimasa, Hiroshi Hikino, Susumu Tsurufuji, Masako Watanabe, and Kazuo Ohuchi. "Antiinflammatory actions of ephedrine in acute inflammations." Planta Medica 51, no. 04 (1985): 325-331.
- [58]. Minhas, Ujla, Ranjana Minz, and Archana Bhatnagar. "Prophylactic effect of *Withania somnifera* on inflammation in a non-autoimmune prone murine model of lupus." Drug discoveries & therapeutics 5, no. 4 (2011): 195-201.
- [59]. Minhas, Ujla, Ranjana Minz, Prabir Das, and Archana Bhatnagar. "Therapeutic effect of *Withania somnifera* on pristane-induced model of SLE." Inflammopharmacology 20, no. 4 (2012): 195-205.
- [60]. Sabina, Evanprince, Sonal Chandel, and Mahaboob Khan Rasool. "Evaluation of analgesic, antipyretic and ulcerogenic effect of Withaferin A." Int J Integr Biol 6, no. 2 (2009): 52-56.
- [61]. Anu Iswarya Jaisankar, Jayalakshmi Somasundaram, D. Ezhilarasan,. 2020. "Can *Achyranthes Aspera* Be Used In Dentistry?". Indian Journal of Forensic Medicine & Toxicology 14 (4):4955-61.
- [62]. Lakshmi, Vijai, Abbas Ali Mahdi, Vaibhav Mishra, and Santosh Kumar Agarwal. "Biopharmaceutics and Therapeutic Challenges."
- [63]. Vetrichelvan, T., and M. Jegadeesan. "Effect of alcohol extract of *Achyranthes aspera* Linn. on acute and subacute



- inflammation." *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives* 17, no. 1 (2003): 77-79.
- [64]. Uma, Bhosale, Radha Yegnanarayan, Prachi Pophale, Mandar Zambare, and R. S. Somani. "Antinociceptive evaluation of an ethanol extract of *Achyranthes aspera* (agadha) in animal models of Nociception." *International Journal of Phytomedicine* 2, no. 4 (2010).