

Gel Formation of Pectin from Okra Leaves, Pulp and Seeds

Dhakane Priti Subhash, Mr.V.G.Rokade, Dr.L.D.Hingane
Aditya Pharmacy College Beed-431122

Submitted: 05-12-2021

Accepted: 20-12-2021

ABSTRACT

Okra plant particularly its fruit is highly mucilage which composed of pectin and high content of carbohydrate. By products of okra plants such as leaves and matured fruits will be discarded whenever the young fruits are harvested which eventually leads to environmental pollution. These by products have potential to become plant-based alternative for bovine and pork related gelatin. This study aimed to determine the gel formation of pectin extracted from okra plant by products particularly the leaves, pulp (skin without seeds) and seeds. Pectin was extracted using sequential extraction with the applications of hot buffer (HB) and hot buffer with chelating agents (CH). CH extraction gave the highest pectin yield (>40%) compared to HB and DA. The HB fraction harbored highly purified pectin due to high anhydrous uronic acid content and degree of esterification. The highest pectin yield was extracted from seeds with an overall fraction yield of 86 %, followed by the leaves (75%) and pulp (71%). The pectin was blended with konjac glucomannan (KG) in 5.0: 1.6 ratio to form gel and stored for 16-18 hr at 4°C ± 1.0. The gel formed using HB extraction was found to have significantly lower ($p < 0.05$) gel strength than HB with CH extraction.

I. INTRODUCTION

Okra (*Abelmoschus esculentus* (L.) Moench) is known as 'lady's fingers', 'bhindi' (India), 'gumbo'

(Africa), and 'bendi or kacang bendi' (Malaysia). The okra leaves are slightly bigger than tea leaves with longer stem; they alternate at wide space intervals up to the top of the stem and they are deeply divided with toothed margins. Okra is widely known as a viscous mucilaginous plant.

Pectin is a methylated ester of polygalacturonic acid that contains 1,4-linked α -D-galacturonic acid residues and can be found in many types of plants. For example, heteropolysaccharides derived from the cell wall of higher terrestrial plants and the peel of citrus fruit, guavas and apples (Sengkhamparnet et al., 2009). It can form gel in the presence of sugar and acidic solution under suitable conditions. It is typically added to jams and jellies as a gelling agent.

Several studies have conducted sequential extraction of pectin from okra leaves, pulp, and seeds and identified its functional groups. Besides, there was a study involved optimization extraction and rheological properties of pectin from okra.

However, limited studies have been done upon comparing the gelling formation of animal-based (gelatin) and plant-based (pectin).

This study applied the concept of "waste into gold" which involved utilization of okra by-product that eventually reduces the waste from post-harvest cultivated into valuable plant-based hydrocolloid.

BOTANICAL DESCRIPTION



ScientificName	ABELMOSCHUS ESCULENTUS(HIBISCUS ESCULENTUS)
PlantFamily	MALVACEAE(MALLROWS)
HindiName	BHINDI
SanskritName	TINDISHA,PITALI,GANDHAMULA
EnglishName	OKRA,LADIES'FINGER,EDIBLEHIBISCUS,OCKRO
OtherNames	BENDI, RAMTURAI, DUK

DESCRIPTION OF OKRA PLANTS



Botanical Classification(Taxonomy)

Kingdom	PLANTAE
Sub-Kingdom	VIRIDIPLANTAE
InfraKingdom	STREPTOPHYTA (landplants)
SuperDivision	EMBRYOPHYTA
Division	TRACHEOPHYTA(TRACHEOPHYTEsOrVascularPlants)
SubDivision	SPERMATOPHYTINA(SPERMATOPHYTEsOrSeedPlants)
Class	MAGNOLIOPSIDA
SuperOrder	ROSANAE
Order	MALVALES
Family	MALVACEAE(MALLOWS)
Genus	ABELMOSCHUS(OKRA)
Species	ABELMOSCHUSESculentus(OKRA)

Plant Description

TypeofPlant	Perennialfloweringplants
NativeRange (GeographicDistribution)	India,NorthAfrica,SouthAfrica
Height (growsupto)	1to2meter
Habitat (typeofenvironment)	Warmtemperateandtropicalregions
Leaves	10–20cm long having 5to7 palmelobes and broadinshape
Flowers	4–8cm indiameter with 5lightyellowpetals
Fruits/Pods	5to10cm long Fibrousfruits(seedpods) containingmucilagoussubstance and white seeds
Seeds	Roundandwhite seeds

THERAPEUTIC INDICATIONS

Okra(Bhindi) is helpful in following health conditions.

Digestive Health

Constipation

Chronicdysentery

Skin & Hairs

Used as soothing and softening poultice

Kidney & Bladder

Dysuria(painfulurination)

Gonorrhœal cystitis

Cystitis

MEDICINAL PROPERTIES

Okra(Bhindi) has following healing properties.
Emollient(softening and soothing effect on skin) –
bland viscid mucilage
Demulcent(soothes the inflamed skin) –
bland viscid mucilage
Diuretic
Aphrodisiac(mucilages substance in unripe pods)
Nutritious

Nutritional composition of okra

Okras more a diet food than staple. Okra seeds have been used on a small scale for oil production. Lipid components greatly contribute to the nutritional and sensory value of almost all types of foods. Nature provides a large number of fat that differ in their chemical and functional properties. Four classes of lipids are habitually found in vegetable oils: triacylglycerols, diacylglycerols, polar lipids, and free fatty acids. The fatty acid composition determines the physical properties, stability, and nutritional value of lipids.

Monoenoic fatty acids and polyunsaturated fatty acids are structurally distinguished by the presence of repeatin g methylene units. These units produce an extremely flexible chain that rapidly reorients through conformational states and constitutes an influential group of molecules that promote health.

The oil mainly consists of linoleic acid (upto 47.4%). Okra seed oil is a rich source of linoleic acid, a polyunsaturated fatty acid essential for human nutrition. Proteins play a particularly important role in human nutrition. The amino acid contents, proportions, and their digestibility by humans characterize a protein's biological value. Okra has been called "a perfect village's vegetable" because of its robust nature, dietary fiber, and distinct seed protein balance of both lysine and tryptophan amino acids (unlike the proteins of cereals and pulses). Okra seed is known to be rich in high quality proteins especially with regard to its content of essential amino acids relative to other plant protein sources. Hence, it plays a vital role in the human diet. Okra also contains carbohydrates and vitamins, and plays a vital role in human diet.

Consumption of young immature okra pods is important as fresh fruits, and it can be consumed in different forms.

The main components are galactose (25%), rhamnose (22%), galacturonic acid (27%) and amino acids (11%).

The mucilage is highly soluble in water. Its solution in water has an intrinsic viscosity value of about 30%.

Dried okra sauce (pods mixed with other ingredients and regularly consumed in West Africa) does not provide an β -carotene (vitamin A) or retinol.

Okra seed is a potential edible oil and flour source

Okra seeds contain about 20 to 40% oil

Okra seed oil yield is comparable to oilseed crops except oil palm and soybean. Moreover, okra seed oil has potential hypocholesterolemic effect. The potential for wide cultivation of okra for edible oil as well as for cake is very high. For example, supplementing maize oil with okra meal increases protein, ash, oil and fiber content.

Okra seed flour has been used to supplement corn flour for a very long time in countries like Egypt to make better quality dough.

The enormous nutritional and other biological activities in the pods and seeds.

Okra seeds were determined to have appreciable protein content. The variations in polysaccharides found in the mucilage are higher in okra pods.

Health benefits of okra

In recent years, increasing attention has been paid to the oil of diet in human health. The high intake of plant products is associated with a reduced risk of a number of chronic diseases, such as atherosclerosis and cancer.

The major antioxidant of vegetables are vitamins C and E, carotenoids, and phenolic compounds, especially flavonoids.

These antioxidants scavenge radicals and inhibit the chain initiation or break the chain propagation (the second defense line). Vitamin E and carotenoids also contribute to the first defense line against oxidative stress, because it quenches single oxygen.

Flavonoids as well as vitamin C showed a protective activity to α -tocopherol in human LDL, and they can also regenerate vitamin E from the α -chromanoxyl radical.

Nutrient antioxidants may act together to reduce reactive oxygen species levels more effectively than single dietary antioxidants, because they can function as synergists.

In addition, a mixture containing both water-soluble and lipid-soluble antioxidants is capable of quenching free radical sin both aqueous and lipid phases.

. Combinations of α -tocopherol or vitamin C plus phenolic compounds also provided synergistic effects in human erythrocyte membrane ghosts and phosphatidylcholine liposomes systems.

. Okra seed is rich in protein and unsaturated fatty acid such as linoleic acid. In some countries, okra also is used in folk medicine as anti-ulcerogenic, gastroprotective, diuretic agents.

. However, little information on antioxidant capabilities of major phenolic compounds from okra seed is available. Okra is also a popular health food due to its high fiber, vitamin C, and folate content. Okra is also a good source of calcium and potassium.

Okra pods contain thick slimy polysaccharides, which are reused to thicken soups and stews, as an egg white substitute, and as a fat substitute in chocolate bar cookies and in chocolate frozen dairy dessert.

Okra is also known for being high in antioxidants activity with different parts of the plant. Atawodi has reported an in vitro antioxidant assay of methanolic extract of okra fruit seeds.

PHYTOCONSTITUENTS

Vitamins C and E, carotenoids, and phenolic compounds, especially flavonoids.

Vitamin E and carotenoids also contribute to the first defense line against oxidative stress, because they quench single oxygen.

Phenolic compounds on antioxidant capabilities of major from okra seed is available.

Flavonoids as well as vitamin C showed a protective activity to α -tocopherol in human LDL, and they can also regenerate vitamin E from the α -chromanoxyl radical.

Chemical constituents

Petals yield flavonoid glycosides; gossypetin and hibiscetin glucosides.

Fresh fruits are rich in pectin and mucilage; it contains oxalic acid, protein, fat, minerals (potassium, sodium, magnesium, sulphur, copper, manganese and iodine), carbohydrate, calcium and phosphorus.

Fresh fruits also contain vitamin A, thiamine, riboflavin, ascorbic acid and niacin d-Galactose, Lrhamnose and dd-dalacturonic acid also isolated from the mucilage of the fruit.

Flavonoid compound has been reported from fruits.

Essential oil isolated from pods and seeds contain aliphatic alcohols, cyclohexanol, p-tolualdehyde (in fruits), a-terpenyl acetate (in seeds) and citral; non-volatile neutral part contains β -sitosterol & its 3 β -galactoside (in seeds).

Leaves have got more or less same constituents. Ripe seeds contain 10-22% edible oil.

Edible Uses:

Immature fruit-cooked on their own or added to soups etc.

They can be used fresh or dried. Mucilaginous, they are commonly used as thickening for soups, stews and sauces.

Seed-cooked or ground into a meal used in making bread or made into 'tofu' or 'tempeh'. The roasted seed is a coffee substitute.

The seed contains up to 22% of an edible oil. (1)

Medicinal Uses Of Okara

Antipasmodic:-

This quality in the seeds is beneficial to the gastrointestinal tract.

Demulcent:

This quality of the roots is very active due to the mucilage which can be used to replace plasmid. It is quality that is present in the leaves, the skin and young pods.

Diuretic:

The young pods act as a diuretic and emollient. Releases accumulation of water that leads to swelling/water retention.

Emollient:

Soothes and softens the skin in infusion of the roots is used in the treatment of syphilis.

The juice of the roots is used externally in Nepal to treat cuts, wounds and boils.

The leaves furnish an emollient poultice. A decoction of the immature capsules is demulcent, diuretic and emollient.

It is used in the treatment of catarrhal infections, dysuria and gonorrhoea.

The seeds are antispasmodic, cordial and stimulant.

OkrainEthnomedicine:

FR UI TS	Infusion offruitmucilage	Indian ethnomedicine	For treating dysentery and diarrhoea in acute inflammation and irritation of the stomach, bowel, and kidney catarrhal infection, and ourinae, and gonorrhoea
	Infusion offruitmucilage	Indian ethnomedicine	Antipyretic and palcereplacement
	A decoction of the immature fruits	Indian ethnomedicine	Demulcent and emollient poultice
	Extract of fleaves and fruits	Indian ethnomedicine	Demulcent, tough less so than that of okra fruit
LE AV ES	Extract of fleaves	Indian ethnomedicine	Extract of leaves mixed with egg albumin and applied on hair which makes black and silky hair
	Leaves	Latin America	Remedies for tumour
RO OT	Extract of froots	Indian ethnomedicine	Demulcent and emollient poultice
	The juice of the roots	Nepal	To treat cuts, wounds and boil
	Infusion of the roots	Traditional medicine of Nicaragua, Atlantic coast and Turkey	Used as stomachic, to treat diabetes, ulcer, used as laxative and treatment of jaundice
SE ED	Seeds	Indian ethnomedicine	Antispasmodic, cordial and stimulant
	Infusion of the roasted seeds	Indian ethnomedicine	Hassudorific properties
	Okraseed	Indian ethnomedicine	Treatment of spermatorrhoea
	Seeds in infusion of roasted okraseed	Latin America	Remedies for tumour

METHOD OF PREPARATION OF GELPECTIN

Plant material

Plant materials used were okra fruits and okra leaves. The plant materials were cleaned and the okra pulp (fruit without seed) was separated manually from the seeds. Then, they were immediately freeze-dried for 96 hr using a freezedrier.

Commercial pectin (CP) of apple pomace and gelatin (bovine) was purchased from Sigma Aldrich Co.

The solvents and chemicals employed in this study were obtained commercially and were of analytical grade

Preparation of 6.67% concentration of gelatin and commercial pectin

The gelatin and commercial pectin of 6.67% concentration were prepared by dissolving 2.66 g of dried gelatin and commercial pectin separately in 40 mL distilled water at 60°C.

The dissolved samples were held in a refrigerator (7°C) for 16–18 hr and were used in viscosity analysis.

Fractionation of alcohol-insoluble solids

Leaves, pulp and seeds were homogenized twice with 70% (v/v) aqueous ethanol at room temperature for 1 hr. After filtration, the insoluble residues were pooled together and soaked in chloroform/methanol (1:1 v/v) with gentle stirring for 30 min to remove low molecular weight (colored) compound

Sequential extraction of okra AIS

The sequential extraction method was conducted according to Vierhuis et al., (2000) with several modifications. The okra AIS (20 g) were extracted using 600 mL of the following extractants: 0.05 M sodium acetate buffer, pH 5.2 (hot buffer, HB) at 70°C, 0.05 M ethylenediaminetetraacetic acid (EDTA) and 0.05 M sodium acetate in 0.05 M sodium oxalate, pH 5.2 at 70°C (chelating agent, CH) of insoluble solids.

After 30 min of extraction, the extract was separated from the insoluble residue by centrifugation at 18,500 × g for 25 min.

Then, the supernatant was coagulated with isopropanol and lyophilized

Sugar composition

The natural sugar composition of all fractions (HB and CH fractions of okra leaves, pulp, and seeds) were determined by high-performance liquid chromatography. Ten milligrams of each fraction were redissolved in 10 mL of 2 M HCl in methanol and hydrolyzed initially at 80°C for 16 hr and subsequently increased the temperature to 121°C for 1 hr.

The solvent was evaporated using a rotary evaporator. Monosaccharides (rhamnose, xylose, arabinose, mannose, fructose, glucose, and galactose) were determined

using a HPLC and refractive index (RI) was used as a detector.

The mobile phase used was acetonitrile/water (75:25, v/v), with a flow rate of 1 mL/min.

A monosaccharide standard consisting of rhamnose, xylose, arabinose, mannose, fructose, glucose, and galactose were used at concentrations between 1 and 5 mg/mL in order to get the equation of calibration curve.

The quantification of uronic acid in the fractions was performed using galacturonic acid

Gel formation

Gel formation was carried out by mixing the fractions with konjac glucomannan (KG) in the ratio of 5.00:1.67 (w/w).

The fractions and KG were weighed separately and mixed together in a beaker.

Then, water was added to solubilize the mixtures and stirred continuously for 15 min in a water bath at 60°C until dissolved completely.

Next, the mixture was stored in the refrigerator for 16–18 hr.

The gel mixture was analyzed to determine its melting point, gel strength and viscosity at 25°C

II. OBSERVATION

Determination of melting point

Solution contained a total of 6.67% (w/v) of fractions (5.00%) and KG (1.67%) was prepared in a thin-walled (12 mm × 75 mm) screw cap test tubes.

The test tubes were filled to leave some headspace for gas released during gel formation and closed.

Determination of gel strength

Gel strength was measured according to the method recommended by the manufacturer of the texture measuring instrument (Gómez-Guillén and Montero, 2001)

The prepared sample will also be used for viscosity determination.

The dimension of the sample in the container was 6 cm in diameter and 2 cm in height.

The solution was cooled down in a refrigerator at 7°C for 16–18 hr.

Measurements were done at 25°C using an Instron mode 14501 Universal Testing Machine (Instron Co., Canton, Mass., USA) with a 5 kN load cell, cross-head speed 1 mm/s, equipped with a 1.27 cm diameter cylindrical Teflon plunger.

Maximum force (expressed in g) was determined when the plunger had penetrated 4 mm into the pectingels. Three replicates were performed for each fraction.

Determination of viscosity

The similar prepared sample (6.67% at 60°C) from previous gel strength test was used.

The viscosity of the sample was determined using an AR

- G2 rheometer (TA Instruments, West Sussex, England) equipped with a heating circulator (Julabo, model F12, Germany).

A cone and plate geometry (60 mm diameter, 1° angle) was used for the measurements.

Approximately 1 mL of each sample was transferred to the Peltier plate and excess material was trimmed off.

A platinum resistance thermometer (PRT) sensor was positioned in the middle of the lower sample plate to ensure accurate sample measurement ($25 \pm 0.1^\circ\text{C}$).

Flow curves with increasing shear rate (0.01–100 s⁻¹) were measured at 25°C.

The downward curve was analyzed using TARheology Advantage Data Analysis Software (version V5.7.0) (Sengkhamparnet al., 2010)

Statistical analysis

All data were expressed as a mean \pm standard deviation.

Data were analyzed using one-way ANOVA using SPSS 15.0. Duncan's multiple-range test was used to assess the difference between the means.

Pearson's correlation test was used to assess the correlations between the means.

III. CONCLUSION

Lady's fingers also known as okra is a plant with high natural viscosity mucilage particularly in the fruits.

Okra was selected in this study, due to the high availability, easy to get all year round at low cost.

Vegetarians, for instance, could not accept animal-based ingredient. In this case, pectin which is plant-based is the most suitable alternative for gelling agent.

In this study, a gel could not be formed by okra pectin itself; therefore, another polysaccharide known as konjac glucomannan (KG) was used.

To conclude, based on the gel properties, HBA and CH fraction derived from okra leaves, pulp, and seeds have good potential to become plant-based gelling agent.

REFERENCES

- [1]. Aina,V.O.,Barau,M.M.,Mamman,O.A.,Zakari,A.and Haruna,H.2012.Extraction and characterization of pectin from peel of lemon (Citrus limon), grapefruit (Citrus paradisi) and sweet orange (Citrus sinensis).British Journal of Pharmacology and Toxicology3(6):259–262
- [2]. Abd Rahman,N.F.,Al Sheraji,H.,Ismail,A.and Mustafa,S.2014.Okra(Abelmoschus esculentus L.Moench) pectin: Extraction yield and chemical composition. In Jamilah,B.,Raja Mohd Hafiz,R.N., Nur Fadhilah,K.M., Nurul Natasha,R., Nur'Ain Najwa,M.N. Proceeding of Malaysia International Halal Research and Education Conference, p.229.Putrajaya: Mariott Putrajaya Hotel
- [3]. Al Sheraji,S.H., Ismail,A., Manap,M.Y., Mustafa,S., Yusof,R.M. and Hassan,F.A.2012.Purification, characterization and antioxidant activity of polysaccharides extracted from the fibrous pulp of Mangifera pajang fruits. LWT-Food Science and Technology48(2):291–296.
- [4]. Anthon,G.E. and Barret,D.M.2008. Combined enzymatic and colorimetric method for determining the uronic acid/methylester content of pectin: application to tomato products. Food Chemistry110:239–247.
- [5]. Burey,P.,Bhandari,B.R.,Howes,T. and Gidley,M.2008.Hydrocolloid gel particles: formation, characterization and application. Critical Review in Food Science and Nutrition48:361–377
- Deters,A.M.,Lengfeld,C. and Hensel,A.2005.Oligo and polysaccharides exhibit a structure dependent bioactivity on human keratinocytes in vitro. Journal of Ethnopharmacology102(3):39