

“Evaluation Of Antiulceractivity Of Methanol Aqueous Flow Extract Of Canthium Diccoccum Line In Male Wistar Albino Rats”

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ABSTRACT

Investigation of the Nephroprotective activity of ethanol extract of *Ficus dalhousiae* on Gentamicin induced nephrotoxicity in male Wistar rats. In this model of nephrotoxicity, 30 adult male Wistar rats (150-200gms) were evenly divided into 5 groups. Group-1 and Group-2 served as untreated and model controls respectively, while Group-3, 4 and 5 were the treatment groups which were simultaneously treated with standard, 200 and 400 mg/kg extract respectively, after each dose of gentamicin (80 mg/kg, i.p.) for 10 days on 1st day,

blood samples for biochemical parameters, while the rat kidneys for histology were obtained under inhaled diether anaesthesia. Gentamicin treatment caused nephrotoxicity as evidenced by marked elevation in blood urea, uric acid and creatinine. Co-administration of extract with Paracetamol decreased rise in blood urea, uric acid and creatinine. Apart from these, histopathological changes also showed the protective nature of extract against Gentamicin induced necrotic damage of renal tissues. It was observed that the ethanol extract conferred nephroprotective activity by histopathological and biochemical observation against Gentamicin induced nephrotoxicity in rats. In the near future could constitute a lead to discovery of a novel drug for treatment of drug induced nephrotoxicity.

I. INTRODUCTION

A peptic ulcer is an open crater or sore that develops in the inner lining (mucosa) of the stomach or the duodenum. A coating of mucus and other chemicals normally shields the stomach and duodenum from digesting themselves.

When these protective mechanisms are disrupted, powerful digestive acids can erode into the lining of these organs and cause peptic ulcer. Peptic ulcer is a major gastro-intestinal disorder caused by an imbalance between offensive (gastric acid, pepsinogen secretion) and defensive (mucus secretion, gastric mucosal integrity) factors. It is a round or oval sore where the lining of the stomach or duodenum has been eaten away by stomach acid and digestive juices.

PATHOPHYSIOLOGY OF PEPTIC ULCER

The term peptic ulcer refers to chronic ulcerative disorder of the upper gastrointestinal tract, which have a common participation of acid and pepsin in their pathogenesis. It includes duodenal ulcer and gastric ulcer, as well as, ulcer associated with Zollinger-Ellison syndrome.

It is a physiologic marvel that gastric juice can easily digest the swallowed pieces of meat but normally has no corrosive action on the gastric mucosa itself. Several factors seen to be involved in the protection of the gastric mucosa from auto digestion. These factors, collectively termed gastric mucosal barrier include

(a) Mucus secreted by the surface epithelial cells and mucous neck cells which forms a water insoluble visco-elastic gel with poor diffusion coefficient for H^+

(b) Bicarbonates secreted by surface epithelial cells into the boundary zone between epithelial cells and the mucus layer. The secretion of mucus and bicarbonates seems to be mediated through prostaglandins.

(c) Tight junctions between the adjacent cells of epithelium

(d) Rapid turnover of the surface epithelial cells and rich mucosal blood supply

(e) Prostaglandins

Endogenous prostaglandins stimulate secretion of gastric mucus as well as gastric and duodenal mucosal bicarbonate. They also participate in the maintenance of gastric mucosal blood flow and integrity of mucosal barrier and promote epithelial cell renewal in response to mucosal injury. A breakdown of the balance between the corrosive action of acid-pepsin and the mucosal resistance results in peptic ulcers. In duodenal ulcer or ulcers due to Zollinger-Ellison syndrome, evidence of an absolute or at least relative gastric hypersecretion can be demonstrated. In contrast, defective mucosal resistance seems to be a major contributory factor in gastric ulcers. It is less common than the DU. A direct gastric mucosal injury is the most important factor in the pathogenesis of GU. Many cases of GU have hyposecretion of gastric acid with secondary slightly increased gastrin secretion (feedback effect). Gastric emptying time is often delayed. GU always occurs in the nonacid-secreting portion of the stomach, often at the lesser curvature. Infection by *H. pylori* is also implicated in the pathogenesis of GU. History of chronic ingestion of aspirin or other non-steroidal anti-inflammatory drug (NSAIDs) as present in 15-25% cases of GU. These drugs are believed to act by depletion of prostaglandins mediated protective mechanisms in gastric mucosa.

SCREENING METHODOLOGIES FOR ANTIULCER ACTIVITY

IN VIVO METHODS

Pylorus ligation in rats (SHAY)

Male or Female Wistar strain albino rats weighing 150-170g are starved for 48 hrs, having access to drinking water and libitum. During this time they are housed single in cages with raised bottoms of wide wire mesh in order to avoid cannibalism and coprophagy. Six animals are used per dose and as controls. Under ether anesthesia a midline abdominal incision is made. The pylorus is ligated, care being exercised that neither damage to the blood supply nor traction on the pylorus occurs. Grasping the stomach with instruments is to be meticulously avoided; else ulceration will invariably develop at such points. The abdominal wall is closed by sutures. The test compounds are given either orally by gavage or injected subcutaneously.

The animals are placed for 19 hrs in plastic cylinders with an inner diameter of 45 mm being closed on both ends by wire mesh. Afterwards, the animals are sacrificed in CO₂ anesthesia. The abdomen is opened and a ligature is placed around the esophagus close to the diaphragm. The stomach is removed and the contents are drained in a centrifuge tube. Along the greater curvature the stomach is opened and pinned on a cork plate. The mucosa is examined with a stereomicroscope. In the rat, the upper two fifths of the stomach form the lumen with squamous epithelium and possess little protective mechanisms against the corrosive action of gastric juice. Below a limiting ridge, in the glandular portion of the stomach, the protective mechanisms are better in the mucosa of the medium two fifths of the stomach than in the lower part, forming the antrum. Therefore, lesions occur mainly in the lumen and in the antrum. The number of ulcers is noted and the severity recorded with the following scores: 0 = no ulcers 1 = superficial ulcers 2 = deep ulcers 3 = perforation. The volume of the gastric content is measured. After centrifugation, acidity is determined by titration with 0.1 N NaOH.

IN VITRO METHOD

The effects of certain antiulcer agents on the antimicrobial activity of antibiotics effective against *H. pylori* were determined in vitro. *Helicobacter pylori* were cultured on Skirrow's agar. Amoxicillin, Clarithromycin, Erythromycin & Tetracycline were used. The antiulcer agents studied by measuring the urease activity. Urease activity was measured by the urease-indophenol method. The minimum inhibitory concentration was determined by a plating method, with *H. pylori* streaked on plates containing various concentrations of the antibiotics plus sublethal doses of the antiulcer agents, which may be evaluated for the antiulcer activity.

Plant profile of *Canthium dicoccum* Botanical

Name: *Canthium dicoccum* (Gaertn.) Merr. **Family:** Rubiaceae;

Common Name: Bogas (P.Bis.), Luing-luing (P.Bis.), Malakafe (P.Bis.), Tandan (Mag.), Ceylon boxwood (Engl.); **Vernacular**

Name: TAMIL: Nallamandharam; **Habit and Habitat:** Malakafe is an unarmed, smooth shrub 3 to 4 meters or more in height. **Leaves** are extremely variable,

ovate, elliptic, ovate or somewhat rounded, 5 to 15 centimeters long, 1.5 to 10 centimeters wide, leathery, shining above, and usually pointed at both ends. **Flowers** are white, with very slender stalks, 5 to 10 millimeters long and borne in compressed, short-stalked cymes. Calyx is cut off at the end or obscurely toothed. Corolla is bell-shaped, with a 4- to 6-millimeter tube, and five somewhat pointed lobes.

II. MATERIALS AND METHODS

Diclofenac sodium- 400 mg/kg (Asha analytical Pvt. Ltd, Uppal, Hyd), Ranitidine- 13.5 mg/kg (Vinallabs, Yadgir, gutta), NaOH (Asha analytical Ltd, Uppal, Hyd), Topfers reagent (Nice, chemicals Pvt Ltd, Cochin), Formalin 10% (Asha analytical Pvt Ltd, Uppal, Hyd), Tween 80 2% (Asha analytical Pvt Ltd, Uppal, Hyd), Distilled water

Collection of plant material

The flowers of *Canthium dicoccum* used for the present studies were collected from Chittoor district of Andhra Pradesh. The plant was identified, confirmed and authenticated by comparing with voucher specimen available at Survey of Medicinal Plants & Collection Unit, Department of Botany, Sri Venkateswara University, Tirupathi by Field Botanist Dr. Madhavshetty. The bark was cut into small pieces and shade dried. The dried material was then pulverized separately into coarse powder by a mechanical grinder. The resulting powder was then used for extraction.

Preparation of Methanolic Extract

The powdered drug was dried and packed well in Soxhlet apparatus and extracted with 1500 ml of methanol for seven days. The extract was concentrated and dried using Rotary flash evaporator. It was kept in desiccators until used.

III. EXPERIMENTAL ANIMALS

Swiss Albino rats adult of either sex were obtained from Mahaveer Enterprises, Hyd (169/CPCSEA/1999). The rats were divided randomly into 4 groups of 5 rats each for each model. Each rat that weighed between 180-200 gm was housed separately (Four rats per cage). The animals were left for 48 hrs to acclimatize to the animal room conditions. They were maintained in standard laboratory conditions of temperature $22 \pm 2^\circ\text{C}$, humidity, 12 hours light and dark cycle fed with standard pellet diet

(Hindustan Lever, Bangalore) and adequate tap water.

METHODS

The rats are divided into different groups each comprising of minimum of five rats as detailed below

Group I – Control rats (Diclofenac + Tween 80)

Group II – Diclofenac induced ulcer rats orally treated with Ranitidine (13.5 mg/kg.b.w)

Group III – Diclofenac induced ulcer rats orally treated with formulation (Canthium dicoccum) at the dose of 200 mg/kg.b.w (SINGLE DOSE)

Group IV – Diclofenac induced ulcer rats orally treated with formulation (DOUBLE DOSE)

The animal in all the groups were kept for 24 h. fasting after that animal of all groups received diclofenac sodium (NSAIDs, 20 mg/kg).

The oral feeding of water and diclofenac sodium was continued for 3 days, the animal of II, III and IV were administered with ranitidine (13.5 mg/kg), flower extract (200 mg/kg), flower extract (400 mg/kg) respectively after 3 h. of diclofenac sodium administration. On 4th day the animals were sacrificed, stomach were removed and cut along the greater curvature to measure the ulcer index.

EVALUATION PARAMETERS

Collection of Gastric Juice

The stomach was excised carefully keeping the esophagus closed, opened along the greater curvature and the gastric contents were removed. The gastric contents were collected in plain tubes and centrifuged at 3000 rpm for 5 min; the volume of the supernatant was expressed as ml/100 gm body weight. The mucosa was flushed with saline and observed for gastric lesions using a dissecting microscope, ulcer score was determined.

Ulcer Scoring

After sacrificing the rat, stomach was removed and opened along the greater curvature, and washed its slowly under running tap water. Put it on the glass slide and observe under 10X magnification for ulcer. Score the ulcers as below.

0=normal coloured stomach

0.5=red colouration

1=spot ulcers

1.5=haemorrhagic streaks

2=Ulcers ≥ 3 but ≤ 5

3=Ulcers > 5

Mean ulcer score for each animal is expressed as Ulcer Index.

Free acidity and Total acidity Centrifuge the gastric contents at 1000 rpm for 10 min, note the volume. Pipette out 1 ml of supernatant liquid and dilute it to 10 ml with distilled water. Note the pH of the solution with the help of pH meter. Titrate the solution against 0.01N NaOH using topfers reagent as an indicator. (It is Dimethyl- amino-azo-benzene with phenolphthalein and used for detection and estimation of hydrochloric acid and total acidity in gastric fluids) Titrate to end point when the solution turns to orange colour. Note the volume of NaOH which corresponds to free acidity. Titrate further till the solution regains its pink colour. Note the total volume of NaOH which corresponds to the total acidity. Acidity (mEq/1/100g) can be expressed as:

Acidity =

$$\frac{\text{Vol. of NaOH} \times \text{Normality} \times 100}{0.1}$$

mEq/1/100g

Statistical analysis of data

Results were expressed as mean \pm S.E.M.

The statistical difference between the groups in the term of the mean rate of wound healing was calculated in terms of ANOVA mean \pm S.E.M. The difference was considered significant if $P < 0.05$.

PHARMACOLOGICAL STUDIES DICLOFENAC SODIUM INDUCED ULCERS

Effect of Gastric Volume

Administration of the extract significantly decreased the gastric volume in comparison with rats treated with Ranitidine. Comparing the gastric volume and gastric acidity, the gastric volume gets decreased, simultaneously the gastric acidity also decreased significantly.

Effect of Free Acidity and Total Acidity

The free acidity and total acidity was determined based on the titre values. The free acidity and total acidity of extract on albino rats decreased significantly in comparison with the standard group treated with Ranitidine. **Ulcer index** The ulcer index was calculated by taking the mean ulcer score of each group. Then the mean ulcer score graph was plotted with groups on x-axis and ulcer index on y-axis. The histograms of different groups were then interpolated by comparing the ulcer index of group I with group II, III and IV. It was noticed that the ulcer index of Dose group (Dose-III&IV) was significantly less when compared to the standard group (Group-II) treated with Ranitidine.

DISCUSSION

Before screening the test extract for antiulcer protective activity, the extract was subjected to the acute toxicity studies as per OECD guidelines no. 420 (fixed dose method). The extract was found to be non-toxic at 2000 mg/kg as indicated by the mortality in the treated group. Hence, the 2000 mg/kg was treated as cut off tolerable dose,

$1/10^{\text{th}}$ (200 mg/kg), and $1/5^{\text{th}}$ (400 mg/kg) of this dose were selected for the further study.

It is evident from the result of the present investigation that the formulation of *Canthium dicoccum*

possesses antiulcer activity in diclofenac induced acute ulcer model. It has shown a significant reduction in the gastric lesions in both the models. Although the etiology of gastric ulcer is not known in most cases, it is generally accepted that it results from an imbalance between aggressive factors and the maintenance of mucosal integrity through the endogenous defence mechanisms. To regain the balance, different therapeutic agents including plant extracts are used (in experimental animals) to inhibit the gastric acid secretion or to boost the mucosal defence mechanisms by increasing mucus production, stabilizing the surface epithelial cells/or enhancing prostaglandin synthesis. Ranitidine the proton pump inhibitor play an important role in the reduction of gastric volume and total acidity and thus perform a cytoprotective effect. The present results demonstrate that the formulation of *Canthium dicoccum* protect the rat gastric mucosa against hemorrhagic

lesion produced by aspirin and ethanol. These inducing methods of gastric lesions are rapid and convenient ways of screening plant extracts for antiulcer potency and cytoprotection in macroscopically and microscopically visible lesions. Diclofenac induced gastric ulcers has been widely used for the experimental evaluation of antiulcer activity. Diclofenac induced gastric lesion formation may be due to stasis in gastric blood flow, which contributes to the development of the hemorrhagic and necrotic aspect of tissue injury. It is of interest to note that administration of antioxidants inhibit aspirin induced gastric injury in the rats. Canthium dicocum possess significant antioxidant activity. In conclusion, the antiulcer effects of the above plants have been reported earlier, but there are no studies reporting the combination of these herbals and their activity in these models are quite impressive. The antiulcer activity of the formulation

Canthium dicocum can be compared to the activity of the standard drug Ranitidine.

IV. SUMMARY & CONCLUSION

From the results discussed above it can be summarized that the Formulation of Canthium dicocum possess the antiulcer activity against the Diclofenac sodium induced gastric ulceration animal model of rats. At the dose level tested it does not show any signs of toxic effects in treated mice as well as rats. Peptic ulcer is the most common disease. Many drugs are there in market to treat the ulcer, but they are having lot of adverse effects. In the present theory, using combination of herbal drug has proved that these are the effective alternatives for chemical drugs. The Anti-ulcer Herbal formulation Canthium dicocum is having significant activity in animal models used, as compared to the standard drug Ranitidine.

EFFECT OF FORMULATION ON GASTRIC VOLUME

Groups	Body wt. of rats	Drug given	Gastric volume
GROUP I	177.2±1.15	Diclofenac sodium +2% Tween 80	1±0.04
GROUP II	161.2±2.15	Ranitidine+ Diclofenac sodium	0.8±0.05
GROUP III	172.5±4.45	Herbal formulation +Diclofenac sodium	0.6±0.03*

GROUP IV 164.4±1.16
 Herbal formulation (double dose)+ Diclofenac sodium
 0.5±0.04**

Values are expressed in terms of mean \pm SEM of 5 rats (ANOVA)

P values: ** < 0.001 - Highly significant * < 0.05 - Significant NS: Non Significant

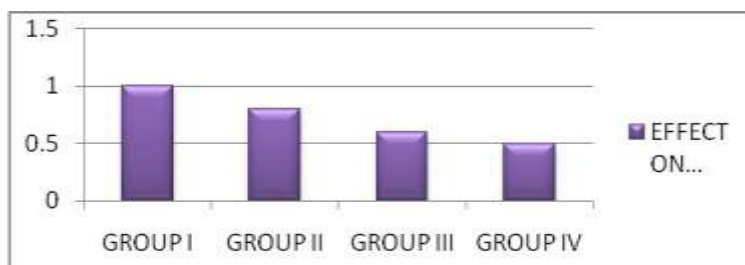
Dose Pattern In Diclofenac Sodium Induced Ulcer Rats

S.NO	Wt of rats (g)	Drugs	Dose (ml)
I) CONTROL GROUP:			
H			0.7+0.5
B	177.2 \pm 1.15		0.6+0.5
T ₁			0.8+0.5
T ₂			0.6+0.5
T ₃			0.7+0.5
II) STANDARD GROUP:			
H			0.8+0.62
B	161.2 \pm 2.15	Diclofenac +	0.9+0.67
T ₁		Ranitidine	0.8+0.63
T ₂			0.8+0.64
T ₃			0.9+0.65
III) DOSE I:			
H			0.5+0.64
B	172.5 \pm 4.45	Extract + Diclofenac	0.6+0.75
T ₁			0.5+0.66
T ₂			0.5+0.70
T ₃			0.6+0.72
IV) DOSE II:			
H			1+0.67
B	164.4 \pm 1.16	Extract (double dose) + diclofenac	1+0.65
T ₁			1+0.65
T ₂			1+0.66
T ₃			1+0.68

PHYTOCHEMICAL SCREENING

S No.	Test	Pet ether Extract	Chloroform Extract	Methanol Extract
1 Carbohydrates				
1	Mohlish's test	+	+	++
	Fehling's test	+	+	++
2 Proteins and amino Acids				

	Ninhydrin test	+		+		+
	Biuret test	+		+		+
	Alkaloids					
3	Mayer's test	++	++			++
	Wagner's test	++	++			++
	Fixed oils and fats					
4	Spot test	+	-			-
	Glycosides					
5	Borntrager's test	+	+			+
	Legal's test	++	+			+
	Iriterpenoids					
6	Tin+thionylchloride		+		-	-
	Phenolics and tannins					
7	Ferric chloride test	+		+		+
	Gelatin test	+		+		+
	Lead acetate test	+		+		+
8	Flavonoids	+		+		+



EFFECT OF FORMULATION ON FREE ACIDITY AND TOTAL ACIDITY

Groups	Body wt. of Drug given rats	Free Acidity	Total Acidity
I	Diclofenac sodium 177.2 ± 1.15 + 2% Tween 80	14.70 ± 0.29	29.6 ± 0.69
II	Ranitidine + 161.2 ± 2.15 Diclofenac sodium	8.8 ± 0.31	15.56 ± 0.69
III	Formulation + 172.5 ± 4.45 Diclofenac sodium	7.3 ± 0.32*	12.56 ± 0.68*
IV	Formulation 164.4 ± 1.16 (double dose) + Diclofenac	4.6 ± 0.42**	9.5 ± 0.59**

Values are expressed in terms of mean ± SEM of 5 rats (ANOVA)

P values: ** < 0.001 - Highly significant

* < 0.05 - Significant NS: Non Significant



EFFECT OF FORMULATION ON ULCER INDEX

Groups	Body wt of rats	Drugs given	Ulcer index
GROUP I	177.2±1.15	Diclofenac sodium +2% Tween 80	3.7±0.14
GROUP II	161.2±2.15	Ranitidine+Asprin Formulation+ Diclofenac sodium Formulation (double dose)+ Diclofenac sodium	2.2±0.10
GROUP III	172.5±4.45		2±0.18*
GROUP IV	164.4±1.16		1.5±0.9**

Values are expressed in terms of mean±SEM of 5 rats (ANOVA)

P values: * < 0.001 - Highly significant; ** < 0.05 - Significant; NS: Non Significant

REFERENCES

- [1]. Textbook of pathophysiology by R.K. Marya, year 2002, Page No. 52-54.
- [2]. Essentials of Medical pharmacology KD Tripathi 4th edition, Year 1999, Page No. 629-642.
- [3]. Goodman and Gilman's The pharmacological basis of therapeutics 11th edition, year 2006, Page No. 978-980.
- [4]. Pharmacology and pharmacotherapeutics by R.S. Satoskar, SDBhandarkar, S.S. Aina pure Revised 20th edition, 2007, Page No. 610-618.
- [5]. Clinical pharmacy and therapeutics. Roger Walker and Clive Edwards 3rd edition, year 2003, page no. 146-147.
- [6]. Drug discovery and evaluation - pharmacological assay. Gerhard vogel, wolfgang. Vog 62.
- [7]. Arthur Guyton C. Text book of medical physiology. 10th ed. Harcourt publisher international company, Singapore; 2000: 264-379. el. Year 1997, page no. 486-491.
- [8]. Herfindal, Gourley. Text book of therapeutic drug and disease management. 7th edn. Chancillivingstone, London; 2000: 425-36.
- [9]. Barrym, Brenner, Floyd, Rector. The kidney 6th ed. W.B. Saunders company, Philadelphia; 2000: 3-67.
- [10]. Paul Munson L. Principles of pharmacology, basic concepts and clinical applications. Chapman and Hall IT Pan International Thomson publishing company, New York; 685.
- [11]. Best, Taylor. Physiological basis of medical practice. 11th ed. Williams and Wilkins. London; 1984: 451-544.
- [12]. Goodman, Gilman's The pharmacological basis of therapeutics. 10th ed.
- [13]. Ch. Santhoshkumari, Prasad & Sreeramulu. Determination of in-vitro & in-vivo activities of aloe vera L against S. pylori.
- [14]. S. Subramanian, D. Sathishkumar, P. Arulselvan, G. P. Senthilkumar & U. Smahadevrao. Evaluation of anti-ulcerogenic potential of aloe-vera leaf gel extract studied in experimental rats.
- [15]. Kossimetowogo, Amegnonaagbonon. Gastro protective effect of hydroalcoholic extract of aloe burtneri.
- [16]. R. Teradaira, N. Shinzato, H. Beppu & K. Fujita. Antigastric ulcer effects in rats of



- aloearborscensmillervar. Natalensis berger extract.
- [17]. Narayaeomlamnam, suthilukpatumraj & narue monvisedo pas, effect of aloevera & sucralfateon gastric microcirculatory changes, cytokine levels, gastric ulcer healing in rats.
- [18]. Rafatullah s, tariq m, al-yahja ma, mossaj, s, ageela, m, evaluation of turmeric (curcuma longa) for gastric and duodenal antiulcer activity in rats. J. Ethnopharmacol. 1990, 29(1):25-34.
- [19]. Bhargava KP, Singh N; Evaluation of the gastric antiulcer activity of fixed oil of *Focimus sanctum* (holy basil).