

Role of Arsenic in the Development of Neurodegenerative Diseases and Memory Impairment

Sonia Bhatt^{1*}, Dr. Mojahid-Ul-Islam²

¹School of Pharmacy, Lingayas Vidyapeeth, Faridabad, Haryana, India

²Head of Department, School of Pharmacy, Lingaya's Vidyapeeth, Faridabad, Haryana.

Date of Submission: 01-10-2021

Date of Acceptance: 15-10-2021

ABSTRACT

Arsenic is a naturally occurring toxic metalloid present worldwide. Arsenic in its inorganic form is the most hazardous and is responsible for causing different diseases like cancer, skin disease, and many neurological disorders in humans which can be demonstrated by several experimental models. Contamination of the drinking water by metal toxins has been a major problem worldwide in which arsenic is one of the major. For the past few decades, there has been increased concern about the health risk due to arsenic and lots of epidemiological studies have been done suggesting an arsenic role in developing neurotoxicity. They also suggest the neurological damage caused by arsenic in children. In this current work, we are trying to explore the different mechanisms involved in arsenic neuropathy and the effects of arsenic-contaminated water on the spatial memory, frontal cortex, and hippocampal ultra-structures, and many other different regions of the brain disrupted by arsenic. Additionally, this review will guide the viewer to determine their future directions for the remission of developing arsenic neuropathy.

Keywords: Arsenic · Oxidative stress · Neurotoxicity · Frontal cortex · Hippocampus

I. INTRODUCTION

Arsenic, a metalloid occurring worldwide found in the environment from various natural and anthropogenic sources [1] at an average concentration of 1.8 ppm [2]. Arsenic is first of all the toxicants posing a significant known or direct and indirect potential threat to humans [3]. It is a co-carcinogen and in lower concentration also known to increase cognitive impairment [4]. It exists both in plants and animals, naturally, it occurs in its organic form which is less toxic [5]. Most exposure to humans occurs from the consumption of water in the form of inorganic arsenic (iAs) i.e., arsenite (As [III]) and arsenate (As[V]), which is abundantly available in water and

the most toxic form of arsenic [6]. The presence of arsenic in the groundwater is a major health problem globally, due to its various ailing effects [7, 8, 9]. Therefore, the identification of new regions with arsenic contamination in Asia has aroused great concern, as the large population is at risk of exposure [10, 11, 12]. Around 140 million people all over the world are exposed to arsenic-contaminated water [13]. Besides, its anthropogenic uses as an alloy, different semiconductor, transistors, metal adhesives and pigment factories [14, 15], mining, fossil fuels burning, natural weathering, and volcano eruptions also introduce arsenic to the environment [16] and therefore the presence of arsenic in the environment enhances the risk of exposure to humans which arises a need to explore its possible clinical effects and its various other links and other environmental sources.

Consumption of arsenic-containing water above 50g/l during pregnancy enhances the risk of fetal loss [17, 18]. It is also found to enhance immune suppression and incident to various infectious diseases in both mother and child [18, 19]. Peripheral neuropathies are quite common in arsenic-exposed individuals [20]. Neurological deficits in children and adults have been reported as the consequences of environmental and occupational exposure to arsenic [21, 22]. Arsenic exposure (>10 µg/L), the permissible water arsenic concentration considered safe by the WHO which can lead to declines in its different ailing effects.

1.1 Arsenic metabolism

The most common valence state of inorganic arsenic (AsV) is its arsenate form whereas arsenite is potentially found in groundwater in the form of sodium arsenite (Na₃AsO₃) due to the interaction with aquifer minerals and physiochemical conditions favoring its release.

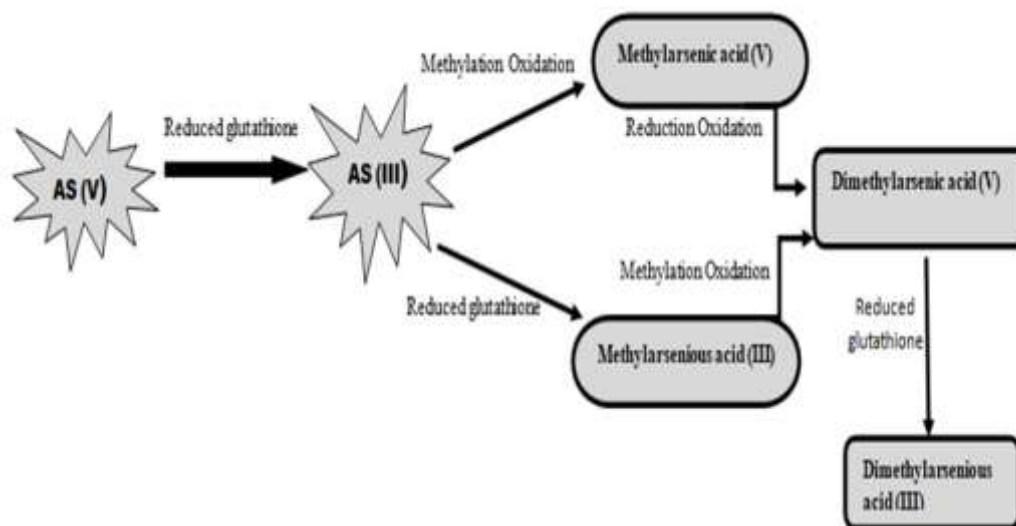


Figure 1. Proposed metabolic pathway of conversion of inorganic arsenic to organic arsenic

The proposed metabolic pathway of Arsenic is shown in Figure1 [23, 24]. Glutathione conjugation and Oxidative methylation are known to be the primary pathways of Arsenic metabolism [25]. Inorganic Arsenic (V) is reduced to Arsenic (III). Inorganic Arsenic (III) is methylated to methyl arsenic acid (V) and methylarsenious acid (III) which is reduced and methylated to Dimethylarsenic acid respectively. Dimethylarsenic acid formed by oxidation and reduction of As(III) and As(v) forms Dimethylarsenious acid (III).

Higher occurrence of skin lesions, cancer, neurological disorders, impairment of psychological and mental health, cardiovascular disorders, and infertility problems are reported in countries like Colombia, Argentina, Japan, Mexico, Bangladesh, and Taiwan where consumption of groundwater is high [26, 27, 28, 29, 30, 31]. Rising evidence of animal and human studies indicates that arsenic has toxic effects on the central and peripheral nervous systems [32, 33]. Presently, the concern over the increasing neurotoxicity of arsenic has been raised. [34, 35]. Development of physiological abnormalities including decreased growth rate, neural defects, malformation [36, 37, 38], and other behavioral changes as arsenic can directly reach the brain as it can cross the blood-brain barrier and placental barrier freely [39, 40, 41, 42, 43]. Several epidemiological studies have demonstrated that arsenic exposure leads to poor, impaired cognitive, and neuropsychological functioning suggesting its role in brain dysfunctions [44- 47]. Infertility, Neural tube defects, neonatal deaths, spontaneous abortion were reported in pregnant women consuming water

contaminated with high arsenic content [36, 48, 49, 50]. Arsenic levels are also found in the breast milk of Bangladeshi women which can adversely affect the infant's health [51]. This finding suggests arsenic may cause impaired fetal growth and can also affect infant health adversely. The underlying mechanism of arsenic-induced neurotoxicity is not clearly known, though several mechanisms have been proposed from various animal and human studies. Arsenic metabolites cause the inactivation of the enzymes involved in the cellular pathways as well as the formation and repair of DNA is the target of the metabolites formed [52], oxidative stress, thiamine deficiency, and decreased acetylcholinesterase activity are some of the mechanisms involved in the arsenic-induced neurotoxicity.

II. EXPERIMENTAL AND EPIDEMIOLOGICAL EVIDENCES OF ARSENIC INDUCED NEUROTOXICITY

2.1 Experimental evidences

Long-term arsenic exposure to humans has been associated with impaired intellectual function in children and adults. Various In vitro and In vivo studies have been conducted as epidemiological evidence of As-induced impaired cognition.

Several studies have been conducted that in mice providing evidence for the toxic effects of arsenic on the entire brain [53, 54] and its discrete regions [55, 56]. Studies also support the evidence that assures the effect of acute and chronic exposure of arsenic develops deficits in spatial

working memory, short term, and long term memory impairment, and damaging different neurological regions responsible for memory impairment. The study indicates the effects of arsenic-containing drinking water on different regions of brain hippocampal ultra-structures, spatial memory, and N-methyl-d-aspartate receptor expression in SD rats [57]. Some studies also indicate that Postnatal exposure to low-concentration of arsenic induces autism-like behavior and affects the frontal cortex in the brain of rats [58].

A. Anwar-Mohamed et. al. investigated and found that acute arsenite (As(III)) exposure can lead to a decrease in cytosolic phospholipase A2 (cPLA2) with a subsequent decrease in brain catalytic activity of mice. It also alters CYP epoxygenases and CYP -hydroxylases, increases expression of cyclooxygenase-2 (COX-2), 5-lipoxygenase (5-LOX), and 15-LOX mRNA while decreasing prostaglandins F2 (PGF2) and PGJ2. This altered state of the different enzymes affects brain development and neurochemistry [59].

2.2 Epidemiological evidences

Many Epidemiological investigations found that low level and chronic exposure to arsenic can be broadly related to serious toxic effects on intellectual functions and early exposure to arsenic can cause full memory deficit by interfering with different brain functions.

In children, adverse neurobehavioral outcomes have been associated with acute and chronic arsenic exposure. A meta-analysis in arsenic-exposed children indicated intelligence deficits; considering 4 cross-sectional studies in China on arsenic exposure and IQ effects, this analysis found that the overall mean IQ score of children who lived in arsenic-exposed areas was more than 6 points lower than the mean score in unexposed children. ³⁶ Indeed, a growing number of studies are confirming intellectual deficits associated with arsenic exposure in children as young as 5 years of age [60-68].

A cross-sectional study conducted on older age relates to chronic arsenic exposure in adolescents. The study found that adolescent exposure to the arsenic-contaminated water in early life performed poorly in 3 of 4 neurological subtests when compared with the unexposed control groups, indicating an alteration in neurobehavioral development in later life might be due to the exposure during childhood exposure [69].

A study on the geriatric population exposed with long term low-level arsenic exposure (average 6.33 mg/L), estimated by the geographic information system (GIS)-based models was significantly associated with impaired executive functioning, slower processing speed, diminished visuospatial skills, poorer global cognition, slower processing speed, reduced language and reduced short term memory [70].

A study on 6-8year old school children in Mexico suggested that arsenic toxicity can lead to different neurological alterations including memory, problem-solving ability, and attention span [63]. A longitudinal cohort study was conducted on children in Bangladesh suggested that there was a reduction in verbal IQ and full-scale IQ associated with arsenic exposure [71].

Urinary arsenic has been inversely associated with full and verbal IQ in 6-8yr old children in Mexico [60]. Finally, arsenic in drinking water has been associated with decreased full IQ scores in children 6- to 10 years of age [62]. In a meta-analysis, including 15 studies evaluating the effect of arsenic on neurodevelopment, in which 13 articles showed a significant effect on neurodevelopment in children of age between 5-15years. In separate meta-analysis assessing arsenic exposure in urine (n ¹/₄ 6) and those that studying drinking water (n ¹/₄ 4), observed that combined effect suggests that a 50% increase in arsenic level in urine would cause a reduced IQ level by 0.39 points, whereas 50% increase in water causes a significant reduction in full-scale IQ by 0.65 points [72]. The timing of exposure to arsenic seems to affect the outcome.

In a study that directly quantified water arsenic exposure, above-median early prenatal maternal arsenic exposure in drinking water was found to be associated with a decreased verbal IQ in children, and late gestational maternal arsenic exposure was associated with a decreased performance IQ in children at 5 years of age [51].

III. MECHANISMS INVOLVEMENT IN NEURODEGENERATION AND MEMORY IMPAIRMENT OF ARSENIC

3.1 Oxidative stress and intracellular pathways activation : In vivo studies have demonstrated that high iAs concentration (i.e., 4 mg/L), leads to neurological damage-inducing oxidative stress and decreased amount of superoxide dismutase, 8-nitroguanine and peroxiredoxin 2 and expression in the neurological tissue of exposed rodents [73],

glutathione after exposure to 50mg/L [74] and increased expression of superoxide anion, singlet oxygen, hydrogen peroxide, hydroxyl radical, and peroxy radicals in different cells [75, 76, 77]. The main signal transduction pathways altered by ROS are: (i) the tyrosine phosphorylation system; (ii) transcription factor families, including the activating protein-1 (AP-1) and nuclear factor-kB (NF-kb); and (iii) mitogen-activated protein kinases (MAPKs) like ERK1/2 [78].

3.2 Pro-inflammatory mediators: Fry et al., found that iAs in concentrations higher than 10mg/L are known to cause overexpression of NF-Kb and IL-1B in the umbilical cord of newborns which causes the overproduction of the inflammatory mediators in urothelial cells [79]. An increase in different inflammatory markers – such as TNF-a, IL-1a, IL-8, and IL6 are also get increased in human and rodents peripheral blood [80, 81, 82].

3.3 Neurotransmitter synthesis and regulation: Arsenic has been found to produce decreased levels of different neurotransmitters, such as norepinephrine (NE), epinephrine (EPN), Dopamine (DA), serotonin (5-HT), and acetylcholine in the different regions of rats brain exposed to sodium arsenite(20mg/L p.o) [83], whereas glutamate expression also gets reduced in the brain when rats are being exposed to sodium arsenite(70mg/L) [84]. As arsenic activates multiple pro-inflammatory signaling pathways which in turn activates indoleamine 2,3-dioxygenase or IDO leading to a reduction in serotonin availability increasing KP's (kynurenine pathway) intermediates which negatively modulates the release of different neurotransmitters including Ach, dopamine, GABA, and glutamate [85]. Therefore, KP impairment may impose a negative impact on the brain and can lead to many neurological and neurodegenerative diseases and also cognitive deficits [86].

3.4 Mitochondrial Dysfunction: There are several studies that support the damage in the mitochondrial region of the brain region by arsenic at different concentrations in rats that are 2 mg /kg BW for 10 weeks [87], 2.5 mg /Kg BW for 4weeks [88], 20 mg/Kg BW for 28 days [89], 10 mg/ kg BW for 16 weeks [90] and 100 ppm for GD 6 to PND 21, 28 and 3 months [91] and found that there is an increase in ROS, lipid peroxidation in the frontal cortex

and hippocampus region of the brain and reduced content of GSH, MnSOD and CAT, GPx and GST in rats and pups respectively in the mitochondria of the cerebral cortex, cerebellum and hippocampus regions of the brain.

3.5 Autophagy impairment: Studies show that autophagy plays an important role in regulating pathophysiology [92], as there are many neurodegenerative diseases associated with impaired autophagy such as Parkinson's, Autism, Alzheimer's, and Huntington's where defective BBB plays a major role [93]. Ram Kumar Manthari et.al. found that the leaky BBB in the cerebral cortex and hippocampus may facilitate the transfer of As and induces autophagy by inhibiting PI3K/Akt/mTOR signaling pathway that causes the rats at PND21 more vulnerable to As-induced neurotoxicity [94]. Qi et al., found that sodium arsenite (0.25mM) inhibited autophagy in human bronchial epithelial cells [95].

3.6 Ultra-structural changes in Brain and accumulation of proteins: Ultrastructural changes in neurons and endothelial cells in the hippocampus, found when the rats are being chronically exposed to sodium arsenite. The expressions of NMDAR subunits in the hippocampus were decreased and there was a reduction in NR2A mRNA levels in the hippocampus after arsenic exposure [96]. Many In vitro and In vivo studies showed that the metabolites induce tau protein hyperphosphorylation, which is acytoskel biomarker for different neurological disorders [97,98]. As arsenic is known to cause oxidative stress which can lead to lead to the activation of kinases, including GSK-3and p38, which phosphorylates tau proteins leading to disassembling of microtubules and is responsible for the formation of tau oligomers [99]. Sodium arsenite has also been reported to decrease PPARg expression while increasing TNF-a and NF-kb contributing to the formation of reactive species and Ab oligomers.

3.7 Impaired expressions of proteins: Studies have shown that arsenic in its trioxide form at 0.15 mg or 1.5 mg or 15 mg doses when administered from gestational to lactational and continued to the pups till PND42 in

drinking water causes declined mRNA expression of TJ proteins, Occludin protein, PI3K, Akt, mTOR, and p62 [94].

3.8 Endoplasmic reticulum stress:Endoplasmic reticulum stress: Accumulation of misfolded proteins triggers unfolded protein response (UPR). An impaired UPR, leading to apoptosis. In brain protein aggregation, improper synaptic function, impair signal transduction contributes to the development of several neurodegenerative diseases. AD, PD, HD, and ALS besides their protein folding and aggregation are also characterized by increased ER stress and UPR activation [100]. The ability of sodium arsenite and metabolites in rat liver cells was also demonstrated [101]. Bolt et al. established that In vitro 1.5mM sodium arsenite activates the ER stress in three-pathways: protein kinase-like endoplasmic reticulum kinase (PERK), inositol requiring enzyme (IRE-1) in human B lymphoblastoid cell line, and activating transcription factor 6 (ATF6) [102]. Chiu et al. found that programmed cell death was induced on exposure to arsenic trioxide through the stimulation of ER stress. It was also found to suppresses the ubiquitin-proteasome system and Akt/mTOR signaling pathways in human sarcoma cells [102, 103].

Studies indicate that arsenic-induced ER stress is associated with both ROS-dependent and ROS-independent pathways, and includes phosphorylation of eIF2 α (the translation initiation factor) and over-expression of chaperones [101,104,105]. Also, involve activation of JNK/Erk pathway has been found to be involved in ER-related cellular apoptosis [104].

Some of the In vitro studies account that iAs also disrupt mitochondrial membrane potential, increasing intracellular calcium level and increased cytochrome C level and impair Akt expression and activation leading to arise different mechanisms for the development of arsenic toxicity activation [106,107,104].

IV. CONCLUSION

Several experiments have been conducted in the last few decades which shows arsenic is one of the major toxins contaminating water and it seems that it is the major cause behind many neurodegenerative diseases and cognitive impairment in adults as well as in children. On analysis, the mechanism underlying the

neurotoxicity include neurotransmitter synthesis and their transmission, Protein accumulation and their impaired expressions, increased oxidative stress, production of pro-inflammatory mediators, impaired autophagy. Based on these, we can conclude that arsenic via the above-discussed mechanisms may lead to imposing neurotoxic effects on different regions of the brain that may lead to neurodegeneration and cognitive impairment.

REFERENCES

- [1]. Brinkel, J., Khan, M.M.H., Kraemer, A., 2009. A systematic review of arsenic exposure and its social and mental health effects with special reference to Bangladesh. *Int. J. Environ. Res. Public Health* 6 (5), 1609–1619.
- [2]. Dani, S.U., 2010. Arsenic for the fool: an exponential connection. *Sci. Total Environ.* 408 (8), 1842–1846.
- [3]. Hughes MF et al. Arsenic exposure and toxicology: a historical perspective. *Toxicol Sci.* 2011;123(2):305–32.
- [4]. Naujokas MF et al. The broad scope of health effects from chronic arsenic exposure: update on a worldwide public health problem. *Environ Health Perspect.* 2013;121(3):295–302.
- [5]. Agency for toxic substances and disease registry (ATSDR). *Priority List of Hazardous Substances*, 2013.
- [6]. Feng Z, Xia Y, Tian D, Wu K, Schmitt M, Kwok RK, et al. DNA damage in buccal epithelial cells from individuals chronically exposed to arsenic via drinking water in Inner Mongolia, China. *Anticancer Res* 2001;21(1A):51–7.
- [7]. Kapaj, S., Peterson, H., Liber, K., Bhattacharya, P., 2006. Human health effects from chronic arsenic poisoning – a review. *J. Environ. Sci. Health A* 41, 2399–2428.
- [8]. Hughes, M.F., Beck, B.D., Chen, Y., Lewis, A.S., Thomas, D.J., 2011. Arsenic exposure and toxicology: a historical perspective. *Toxicol. Sci.* 123 (2), 305–332.
- [9]. Chen, Y., Graziano, J.H., Parvez, F., Liu, M., Slavkovich, V., Kalra, T., Argos, M., Islam, T., Ahmed, A., Rakibuz-Zaman, M., 2011. Arsenic exposure from drinking water and mortality from cardiovascular disease in Bangladesh: A prospective cohort study. *BMJ* 342, d2431.

- [10]. Rahman, M.M., Naidu, R., Bhattacharya, P., 2009. Arsenic contamination in groundwater in the Southeast Asia region. *Environ. Geochem. Health* 31(1), 9–21.
- [11]. Saha, D., Shukla, R.R., 2013. Genesis of arsenic-rich groundwater and the search for alternative safe aquifers in the Gangetic Plain, India. *Water Environ. Res.* 85 (12), 2254–2264.
- [12]. Van Geen, A., Win, K.H., Zaw, T., Naing, W., Mey, J.L., Mailloux, B., 2014. Confirmation of elevated arsenic levels in groundwater of Myanmar. *Sci. Total Environ.* 15 (478), 21–24.
- [13]. Kris, F., 2009. Nutrient protection against arsenic toxicity: folate, cysteine support methylation in children. *Environ. Health Perspect.* 117 (5), A211.
- [14]. Fowler, B.A., Sexton, M.J., 2007. Gallium and semiconductor compounds. In: Nordberg, M. (Ed.), *Handbook on the Toxicology of Metals*. Academic Press Inc., NY, pp. 547–555.
- [15]. Wlodarczyk, B.J., Palacios, A.M., Chapa, C.J., Zhu, H., George, T.M., Finnell, R.H., 2011. Genetic basis of susceptibility to teratogen induced birth defects. *Am. J. Med. Genet. C: Semin. Med. Genet.* 157 (3), 215–226.
- [16]. Vall, O., Gómez-Culebras, M., Garcia-Algar, O., Joya, X., Velez, D., Rodríguez-Carrasco, E., Puig, C., 2012. Assessment of prenatal exposure to arsenic in Tenerife Island. *PLoS ONE* 7 (11), e50463.
- [17]. Rahman, A., Vahter, M., Ekstrom, E.C., Rahman, M., GolamMustafa, A.H., Wahed, M.A., Yunus, M., Persson, L.A., 2007. Association of arsenic exposure during pregnancy with fetal loss and infant death: a cohort study in Bangladesh. *Am. J. Epidemiol.* 165, 1389–1396.
- [18]. Ahmed, S., Mahabbat-e Khoda, S., Rekha, R.S., Gardner, R.M., Ameer, S.S., Moore, S., Ekström, E.C., Vahter, M., Raqib, R., 2011. Arsenic-associated oxidative stress, inflammation, and immune disruption in human placenta and cord blood. *Environ. Health Perspect.* 119 (2), 258–264.
- [19]. Raqib, R., Ahmed, S., Sultana, R., Wagatsuma, Y., Mondal, D., Hoque, A.M., Nermell, B., Yunus, M., Roy, S., Persson, L.A., Arifeen, S.E., Moore, S., Vahter, M., 2009. Effects of in utero arsenic exposure on child immunity and morbidity in rural Bangladesh. *Toxicol. Lett.* 28 (185(3)), 197–202.
- [20]. Barton, A., McLean, B., 2013. An unusual case of peripheral neuropathy possibly due to arsenic toxicity secondary to excessive intake of dietary supplements. *Ann. Clin. Biochem.* 50 (Pt 5), 496–500.
- [21]. Rosado, J.L., Ronquillo, D., Kordas, K., Rojas, O., Alatorre, J., Lopez, P., Garcia-Vargas, G., Del Carmen Caamaño, M., Cebrián, M.E., Stoltzfus, R.J., 2007. Arsenic exposure and cognitive performance in Mexican schoolchildren. *Environ. Health Perspect.* 115 (9), 1371–1375.
- [22]. Dong, J., Su, S.Y., 2009. The association between arsenic and children's intelligence: a meta-analysis? *Biol. Trace Elem. Res.* 129 (1–3), 88–93, Summer.
- [23]. Liu J, Waalkes MP (2018) Liver is a target of arsenic carcinogenesis. *Toxicol Sci* 105:24–32.
- [24]. Mukherjee B, Bindhani B, Saha H, Sinha D, Ray MR (2014) Platelet hyperactivity, neurobehavioral symptoms and depression among Indian women chronically exposed to low level of arsenic. *Neurotoxicology* 45:159–167.
- [25]. Zhao F, Liao Y, Tang H, Piao J, Wang G, Jin Y (2017) Effects of developmental arsenite exposure on hippocampal synapses in mouse offspring. *Metallomics* 9:1394–1412.
- [26]. Guha Mazumder, D.; Dasgupta, U.B. Chronic arsenic toxicity: Studies in West Bengal, India. *Kaohsiung J. Med. Sci.* 2011, 27, 360–370.
- [27]. Ishii, N.; Mochizuki, H.; Ebihara, Y.; Shiomi, K.; Nakazato, M. Clinical Symptoms, Neurological Signs, and Electrophysiological Findings in Surviving Residents with Probable Arsenic Exposure in Toroku, Japan. *Arch. Environ. Contam. Toxicol.* 2018, 75, 521–529.
- [28]. Mochizuki, H.; Phyu, K.P.; Aung, M.N.; Zin, P.W.; Yano, Y.; Myint, M.Z.; Thit, W.M.; Yamamoto, Y.; Hishikawa, Y.; Thant, K.Z.; et al. Peripheral neuropathy induced by drinking water contaminated with low-dose arsenic in Myanmar. *Environ. Health Prev. Med.* 2019, 24, 23.
- [29]. Onishi, H. Arsenic. In *Handbook of Geochemistry*; Wedepohl, K.H., Ed.; Springer: New York, NY, USA, 1969;

- Volume II. 8. Lunde, G. Occurrence and transformation of arsenic in the marine environment. *Environ. Health Perspect.* 1977, 19, 47–52.
- [30]. Newcombe, C.; Raab, A.; Williams, P.N.; Deacon, C.; Haris, P.I.; Meharg, A.A.; Feldmann, J. Accumulation or production of arsenobetaine in humans? *J. Environ. Monit.* 2010, 12, 832–837.
- [31]. Aposhian, H.V.; Gurzau, E.S.; Le, X.C.; Gurzau, A.; Healy, S.M.; Lu, X.; Ma, M.; Yip, L.; Zakharyan, R.A.; Maiorino, R.M.; et al. Occurrence of monomethylarsonous acid in urine of humans exposed to inorganic arsenic. *Chem. Res. Toxicol.* 2000, 13, 693–697.
- [32]. Chandravanshi LP, Yadav RS, Shukla RK, Singh A, Sultana S, Pant AB, Parmar D, Khanna VK (2014a) Reversibility of changes in brain cholinergic receptors and acetylcholinesterase activity in rats following early life arsenic exposure. *Int J Dev Neurosci* 34:60–75.
- [33]. Chandravanshi LP, Shukla RK, Sultana S, Pant AB, Khanna VK (2014b) Early life arsenic exposure and brain dopaminergic alterations in rats. *Int J Dev Neurosci* 38:91–104.
- [34]. Chandravanshi LP, Gupta R, Shukla RK (2018 Mar 3) Developmental neurotoxicity of arsenic: involvement of oxidative stress and mitochondrial functions. *Biol Trace Elem Res.*
- [35]. Chen Y, Graziano JH, Parvez F, Liu M, Slavkovich V, Kalra T, Argos M, Islam T, Ahmed A, Rakibuz-Zaman M (2011) Arsenic exposure from drinking water and mortality from cardiovascular disease in Bangladesh: prospective cohort study. *BMJ* 342:d2431.
- [36]. Mazumdar M (2017) Does arsenic increase the risk of neural tube defects among a highly exposed population? A new case–control study in Bangladesh. *Birth Defects Res* 109:92–98.
- [37]. Rahman A, Granberg C, Persson LA (2017) Early life arsenic exposure, infant and child growth, and morbidity: a systematic review. *Arch Toxicol* 91:3459–3467.
- [38]. Shih YH, Islam T, Hore SK, Sarwar G, Shahriar MH, Yunus M, Graziano JH, Harjes J, Baron JA, Parvez F, Ahsan H, Argos M (2017) Associations between prenatal arsenic exposure with adverse pregnancy outcome and child mortality. *Environ Res* 158:456–461.
- [39]. Hirner AV, Rettenmeier AW (2010) Methylated metalloid species in humans. *Met Ions Life Sci* 7:465–521.
- [40]. Jin Y, Xi S, Li X, Lu C, Li G, Xu Y, Qu C, Niu Y, Sun G (2006) Arsenic speciation transported through the placenta from mother mice to their newborn pups. *Environ Res* 101:349–355.
- [41]. Rudge CV, Röllin HB, Nogueira CM, Thomassen Y, Rudge MC, Odland JØ (2009) The placenta as a barrier for toxic and essential elements in paired maternal and cord blood samples of South African delivering women. *J Environ Monit* 11:1322–1330.
- [42]. Sanders AP, Desrosiers TA, Warren JL, Herring AH, Enright D, Olshan AF, Meyer RE, Fry RC (2014) Association between arsenic, cadmium, manganese, and lead levels in private wells and birth defects prevalence in North Carolina: a semi-ecologic study. *BMC Public Health* 14:955.
- [43]. Willhite CC, Ferm VH (1984) Prenatal and developmental toxicology of arsenicals. *Adv Exp Med Biol* 177:205–228.
- [44]. Guha Mazumder D., Dasgupta U.B. Chronic arsenic toxicity: Studies in West Bengal, India. *Kaohsiung J. Med. Sci.* 2011;27:360–370.
- [45]. Ishii N., Mochizuki H., Ebihara Y., Shiomi K., Nakazato M. Clinical Symptoms, Neurological Signs, and Electrophysiological Findings in Surviving Residents with Probable Arsenic Exposure in Toroku, Japan. *Arch. Environ. Contam. Toxicol.* 2018;75:521–529.
- [46]. Mochizuki H., Phyu K.P., Aung M.N., Zin P.W., Yano Y., Myint M.Z., Thit W.M., Yamamoto Y., Hishikawa Y., Thant K.Z., et al. Peripheral neuropathy induced by drinking water contaminated with low-dose arsenic in Myanmar. *Environ. Health Prev. Med.* 2019;24:23.
- [47]. Onishi H. Arsenic. In: Wedepohl K.H., editor. *Handbook of Geochemistry*. Volume II Springer; New York, NY, USA: 1969.
- [48]. Milton AH, Hussain S, Akter S, Rahman M, Mouly TA, Mitchell K (2017) A review of the effects of chronic arsenic exposure on adverse pregnancy outcomes. *Int J Environ Res Public Health* 6.

- [49]. Rahman A, Persson LÅ, Nermell B, El Arifeen S, Ekström EC, Smith AH, Vahter M (2010) Arsenic exposure and risk of spontaneous abortion, stillbirth, and infant mortality. *Epidemiology* 21:797–804.
- [50]. Rahman A, Kumarathanan P, Gomes J (2016) Infant and mother related outcomes from exposure to metals with endocrine disrupting properties during pregnancy. *Sci Total Environ* 569- 570:1022–1031.
- [51]. Fängström B, Moore S, Nermell B, Kuenstl L, Goessler W, Grandér M, Kabir I, Palm B, Arifeen SE, Vahter M (2008) Breast-feeding protects against arsenic exposure in Bangladeshi infants. *Environ Health Perspect* 116:963–969.
- [52]. Ratnaik R.N. Acute and chronic arsenic toxicity. *Postgrad. Med. J.* 2003;79:391–396.
- [53]. Chaudhuri, A.N., Basu, S., Chattopadhyay, S., and Das Gupta, S. 1999. Effect of high arsenic content in drinking water on rat brain. *Indian J. Biochem. Biophys.* 36(1): 51–54.
- [54]. Tripathi N., Kannan G.M., Pant B.P., Jaiswal D.K., Malhotra P.R., and Flora S.J. 1997. Arsenic-induced changes in certain neurotransmitter levels and their recoveries following chelation in rat whole brain. *Toxicol. Lett.* 92(3): 201– 208.
- [55]. Sanchez Pena, L.C., Petrosyan, P., Morales, M., Gonzalez, N.B., Gutierrez-Ospina, G., Del Razo, L.M., et al. 2010. Arsenic species, AS3MT amount, and AS3MT gene expression in different brain regions of mouse exposed to arsenite. *Environ. Res.* 110(5): 428–434.
- [56]. Yen, C.C., Ho, T.J., Wu, C.C., Chang, C.F., Su, C.C., Chen, Y.W., et al. 2011. Inorganic arsenic causes cell apoptosis in mouse cerebrum through an oxidative stress-regulated signaling pathway. *Arch. Toxicol.* 85(6): 565–575.
- [57]. Jia -hua Luo, Zhi-qun Qiu, Wei-qun Shu, Yong-yan Zhang, Liang Zhang, Ji-an Chen. ,2008. Effects of arsenic exposure from drinking water on spatial memory, ultra-structures and NMDAR gene expression of hippocampus in rats. *Toxicology Lett.* 184 (2009) 121–125.
- [58]. Hao Zhou, Weiqing Zhao, Liu Ye, Zhihe Chen, Yuxia Cui . ,2018. Postnatal low-concentration arsenic exposure induces autism-like behavior and affects frontal cortex neurogenesis in rats. *Environmental Toxicology and Pharmacology.*
- [59]. Anwar Anwar-Mohamed, Osama H. Elshenawy, Ahmed A. El-Sherbeni .,et al., 2014. Acute arsenic treatment alters arachidonic acid and its associated metabolite levels in the brain of C57Bl/6 mice. *Can. J. Physiol. Pharmacol.* 92: 693–702.
- [60]. Calderón J, Navarro ME, Jimenez-Capdeville ME, et al. Exposure to arsenic and lead and neuropsychological development in Mexican children. *Environ Res* 2001; 85:69e76.
- [61]. Ehrenstein O, Poddar S, Yuan Y, et al. Children’s intellectual function in relation to arsenic exposure. *Epidemiology* 2007;18:44e51.
- [62]. Rocha-Amador D, Navarro ME, Carrizales L, Morales R, Calderón J. Decreased intelligence in children and exposure to fluoride and arsenic in drinking water. *Cad Saude Publica* 2007;23:579e87.
- [63]. Rosado JL, Ronquillo D, Katarzyna K, et al. Arsenic exposure and cognitive performance in Mexican schoolchildren. *Environ Health Perspect* 2007;115:9.
- [64]. Siripitayakunkit U, Visudhiphan P, Pradipasen M, Vorapongsathron T. Association between chronic arsenic exposure and children’s intelligence in Thailand. In: Chappell WR, Abernathy CO, Calderon RL, eds. *Arsenic Exposure and Health Effects III*. Oxford, UK: Elsevier Science Ltd; 1991:141e9.
- [65]. Wang SX, Wang ZH, Cheng XT, et al. Arsenic and fluoride exposure in drinking water: children’s IQ and growth in Shanyin County, Shanxi Province, China. *Environ Health Perspect* 2007;115:643e7.
- [66]. Wasserman GA, Liu X, Parvez F, et al. Water arsenic exposure and children’s intellectual function in Araihasar, Bangladesh. *Environ Health Perspect* 2004;112:1329e33.
- [67]. Wasserman GA, Liu X, Parvez F, et al. Arsenic and manganese exposure and children’s intellectual function. *Neurotoxicology* 2011;32:450e7.
- [68]. Wright RO, Amarasiriwardena C, Woolf AD, Jim R, Bellinger DC. Neuropsychological correlates of hair arsenic, manganese, and cadmium levels in school-age children residing near a

- hazardous waste site. *NeuroToxicology* 2006;27:210e6).
- [69]. Tsai SY, Chou HY, The HW, Chen CM, Chen CJ. The effects of chronic arsenic exposure from drinking water on the neurobehavioral development in adolescence. *Neurotoxicology* 2003;24:747e53.
- [70]. O'Bryant SE, Edwards M, Menon CV, Gong G, Barber R. Long-term low-level arsenic exposure is associated with poorer neuropsychological functioning: a project FRONTIER study. *Int J Environ Res Public Health* 2011;8:861e74.
- [71]. Hamadani JD, Tofail F, Nermell B, et al. Critical windows of exposure for arsenic-associated impairment of cognitive function in pre-school girls and boys: a population-based cohort study. *Int J Epidemiol* 2011;40:1593e604.
- [72]. Rodríguez-Barranco M, Lacasana M, et al. Association of arsenic, cadmium and manganese exposure with neurodevelopment and behavioral disorders in children: A systematic review and metanalysis. *Sci Total Environ* 2013;454-455:562e77.
- [73]. Piao, F.Y., Li, S., Li, Q.J., Ye, J.X., Liu, S.A., 2011. Abnormal expression of 8-nitroguanine in the brain of mice exposed to arsenic sub chronically. *Ind. Health* 49 (2), 151–157.
- [74]. Dwivedi, N., Flora, S.J., 2011. Concomitant exposure to arsenic and organophosphates on tissue oxidative stress in rats. *Food Chem. Toxicol.* 49 (5), 1152–1159.
- [75]. Gharibzadeh, S., Hoseini, S.S., 2008. Arsenic exposure may be a risk factor for Alzheimer's disease. *J. Neuropsychiatry Clin. Neurosci.* 20 (4), 501.
- [76]. Kumagai, Y., Sumi, D., 2007. Arsenic: signal transduction, transcription factor, and biotransformation involved in cellular response and toxicity. *Annu. Rev. Pharmacol. Toxicol.* 47, 243–262.
- [77]. Wnek, S.M., Medeiros, M.K., Eblin, K.E., Gandolfi, A.J., 2009. Persistence of DNA damage following exposure of human bladder cells to chronic monomethylarsonous acid. *Toxicol. Appl. Pharmacol.* 241 (2), 202-209.
- [78]. Shi, H., Shi, X., Liu, K.J., 2004. Oxidative mechanism of arsenic toxicity and carcinogenesis. *Mol. Cell Biochem.* 255 (1–2), 67–78.
- [79]. Fry, R.C., Navasumrit, P., Valiathan, C., Svensson, J.P., Hogan, B.J., Luo, M., et al., 2007. Activation of inflammation/NF-kappaB signaling in infants born to arsenic-exposed mothers. *PLoS Genet.* 3 (11), e207.
- [80]. Valavanidis, A., Vlachogianni, T., Fiotakis, K., 2009. Tobacco smoke: involvement of reactive oxygen species and stable free radicals in mechanisms of oxidative damage, carcinogenesis and synergistic effects with other respirable particles. *Int. J. Environ. Res. Public Health* 6 (2), 445–462.
- [81]. Wu, J., Liu, J., Waalkes, M.P., Cheng, M.L., Li, L., Li, C.X., et al., 2008. High dietary fat exacerbates arsenic-induced liver fibrosis in mice. *Exp. Biol. Med.* (Maywood) 233 (3), 377–384.
- [82]. Yang, C., Wu, J., Zhang, R., Zhang, P., Eckard, J., Yusuf, R., et al., 2005. Caffeic acid phenethyl ester (CAPE) prevents transformation of human cells by arsenite (As) and suppresses growth of As-transformed cells. *Toxicology* 213 (1–2), 81–96.
- [83]. Yadav, R.S., Shukla, R.K., Sankhwar, M.L., Patel, D.K., Ansari, R.W., Pant, A.B., et al., 2010. Neuroprotective effect of curcumin in arsenic-induced neurotoxicity in rats. *Neurotoxicology* 31 (5), 533–539.
- [84]. Jiang, S., Su, J., Yao, S., Zhang, Y., Cao, F., Wang, F., et al., 2014. Fluoride and arsenic exposure impairs learning and memory and decreases mGluR5 expression in the hippocampus and cortex in rats. *PLoS One* 9 (4), e96041.
- [85]. Pershing, M.L., Bortz, D.M., Pocivavsek, A., Fredericks, P.J., Jorgensen, C.V., Vunck, S.A., et al., 2015. Elevated levels of kynurenic acid during gestation produce neurochemical, morphological, and cognitive deficits in adulthood: implications for schizophrenia. *Neuropharmacology* 90, 33–41.

- [86]. Schwarcz, R., Pellicciari, R., 2002. Manipulation of brain kynurenines: glial targets, neuronal effects, and clinical opportunities. *J. Pharm. Exp. Ther.* 303 (1), 1–10.
- [87]. Ram Kumar M, Flora SJS, Reddy GR. 2013. Monoisooamyl 2,3- dimercaptosuccinic acid attenuates arsenic induced toxicity: behavioral and neurochemical approach. *Environ. Toxicol. Pharmacol.* 36: 231–242.
- [88]. Dwivedi N, Mehta A, Yadav A, Binukumar BK, Gill KD, Flora SJS. 2011. MiADMSA reverses impaired mitochondrial energy metabolism and neuronal apoptotic cell death after arsenic exposure in rats. *Toxicol. Appl. Pharmacol.* 256: 241–248.
- [89]. Srivastava P, Yadav RS, Chandravanshi LP, Shukla RK, Dhuriya YK, Chauhan LK, Dwivedi HN, Pant AB, Khanna VK. 2014. Unraveling the mechanism of neuroprotection of curcumin in arsenic induced cholinergic dysfunctions in rats. *Toxicol. Appl. Pharmacol.* 279: 428–440.
- [90]. Ghosh S, Dungdung SR, Chowdhury ST, Mandal AK, Sarkar S, Ghosh D, Das N. 2011. Encapsulation of the flavonoid quercetin with an arsenic chelator into nanocapsules enables the simultaneous delivery of hydrophobic and hydrophilic drugs with a synergistic effect against chronic arsenic accumulation and oxidative stress. *Free Radic. Biol. Med.* 51: 1893–1902.
- [91]. Kadeyala PK, Sannadi S, Gottipolu RR. 2013. Alterations in apoptotic caspases and antioxidant enzymes in arsenic exposed rat brain regions: reversal effect of essential metals and a chelating agent. *Environ. Toxicol. Pharmacol.* 36: 1150–1166.
- [92]. Meijer AJ, Codogno P (2006) Signalling and autophagy regulation in health, aging and disease. *Mol Aspects Med* 27:411–425.
- [93]. Shintani T, Klionsky DJ (2004) Autophagy in health and disease: a double-edged sword. *Science* 306:990–995.
- [94]. Ram Kumar Manthari, Chiranjeevi Tikka, Mohammad Mehdi Ommati, et al., 2018. Arsenic induces autophagy in developmental mouse cerebral cortex and hippocampus by inhibiting PI3K/Akt/mTOR signaling pathway: involvement of blood–brain barrier’s tight junction proteins. *Archives of Toxicology*.
- [95]. Qi, Y., Zhang, M., Li, H., Frank, J.A., Dai, L., Liu, H., et al., 2014. Autophagy inhibition by sustained overproduction of IL6 contributes to arsenic carcinogenesis. *Cancer Res.* 74 (14), 3740–3752.
- [96]. Luo, J.H., Qiu, Z.Q., Shu, W.Q., Zhang, Y.Y., Zhang, L., Chen, J.A., 2009. Effects of arsenic exposure from drinking water on spatial memory, ultra-structures and NMDAR gene expression of hippocampus in rats. *Toxicol. Lett.* 184 (2), 121–125.
- [97]. Giasson, B.I., Sampathu, D.M., Wilson, C.A., Vogelsberg-Ragaglia, V., Mushynski, W.E., Lee, V.M.Y., 2002. The environmental toxin arsenite induces tau hyperphosphorylation. *Biochemistry* 41 (51), 15376–15387.
- [98]. Vahidnia, A., van der Voet, G.B., de Wolff, F.A., 2007b. Arsenic neurotoxicity— a review. *Hum. Exp. Toxicol.* 26 (10), 823–832.
- [99]. Ballatore, C., Lee, V.M., Trojanowski, J.Q., 2007. Tau-mediated neurodegeneration in Alzheimer’s disease and related disorders. *Nat. Rev. Neurosci.* 8 (9), 663–672.
- [100]. Dunys, J., Duplan, E., Checler, F., 2014. The transcription factor X-box binding protein-1 in neurodegenerative diseases. *Mol. Neurodegener.* 9, 35.
- [101]. Naranmandura, H., Xu, S., Koike, S., Pan, L.Q., Chen, B., Wang, Y.W., et al., 2012. The endoplasmic reticulum is a target organelle for trivalent dimethylarsinic acid (DMAIII)-induced cytotoxicity. *Toxicol. Appl. Pharmacol.* 260 (3), 241–249.
- [102]. Bolt, A.M., Zhao, F., Pacheco, S., Klimecki, W.T., 2012. Arsenite-induced autophagy is associated with proteotoxicity in human lymphoblastoid cells. *Toxicol. Appl. Pharmacol.* 264 (2), 255–261.
- [103]. Chiu, H.W., Tseng, Y.C., Hsu, Y.H., Lin, Y.F., Foo, N.P., Guo, H.R., et al., 2015. Arsenic trioxide induces programmed cell death through stimulation of ER stress and inhibition of the ubiquitin-proteasome system in human sarcoma cells. *Cancer Lett.* 356 (2 Pt B), 762–772.
- [104]. Lu, T.H., Tseng, T.J., Su, C.C., Tang, F.C., Yen, C.C., Liu, Y.Y., et al., 2014. Arsenic induces reactive oxygen species-caused

- neuronal cell apoptosis through JNK/ERK-mediated mitochondria-dependent and GRP78/CHOP-regulated pathways. *Toxicol. Lett.* 224 (1), 130–140.
- [105]. Weng, C.Y., Chiou, S.Y., Wang, L., Kou, M.C., Wang, Y.J., Wu, M.J., 2014. Arsenic trioxide induces unfolded protein response in vascular endothelial cells. *Arch. Toxicol.* 88 (2), 213–226.
- [106]. King, Y.A., Chiu, Y.J., Chen, H.P., Kuo, D.H., Lu, C.C., Yang, J.S., 2014. Endoplasmic reticulum stress contributes to arsenic trioxide-induced intrinsic apoptosis in human umbilical and bone marrow mesenchymal stem cells. *Environ. Toxicol.*
- [107]. Yen, Y.P., Tsai, K.S., Chen, Y.W., Huang, C.F., Yang, R.S., Liu, S.H., 2012. Arsenic induces apoptosis in myoblasts through a reactive oxygen species-induced endoplasmic reticulum stress and mitochondrial dysfunction pathway. *Arch. Toxicol.* 86 (6), 923–933.