

“Physicochemical Characterization, Antioxidant and Antimicrobial Assessment of Some Newly Synthesized Aza Macrocylic Complexes of Cr (III) Metal Ions”

Om Prakash Gurjar, Sushama Kumari, pooja Saini, Krishna Atal, Swati Bugalia
Department of Chemistry, University of Rajasthan, Jaipur, 302004, Rajasthan, India

Submitted: 10-07-2023

Accepted: 20-07-2023

ABSTRACT: A fresh set of macrocyclic complexes of chromium typed $[\text{Cr}(\text{C}_{30}\text{H}_{20}\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ and $[\text{Cr}(\text{C}_{30}\text{H}_{18}\text{Cl}_2\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ were synthesized in methanolic medium via template synthetic route. These complexes were prepared by [2+2] cyclocondensation reaction of diamines i.e. 4-aminobenzoic hydrazide and 2-aminobenzhydrazide with α -diketones i.e. isatin and 5-chloroisatin in presence of chromium acetylacetonate template ion. Physicochemical properties were studied by decomposition temperature, conductance measurement, elemental analyses and magnetic moment measurements. These complexes were found soluble in DMSO. Various techniques were utilized for spectral analysis of synthesized complexes such as SEM, IR, NMR, mass spectrometry and UV/Vis. All the produced complexes were assessed for their antibacterial potency contrary to the bacterial strains (*E. coli* and *S. aureus*) and fungal strains (*Alternaria alternata* and *Fusarium solani*) by using Agar well diffusion method. Antioxidant activities of all these complexes were also tested by the help of DPPH method and compared with standard ascorbic acid and showed satisfactory results.

KEY WORDS: -Macrocyclic complexes, chromium(III), antibacterial, antifungal, antioxidant.

I. INTRODUCTION

Recently, researchers developed their interest on aza macrocycles in the area of coordination chemistry due to their remarkable features [1–6]. Study of this metal macrocyclic chemistry is growing very quickly [7-14]. Today creation of well-ordered macrocycles containing metal is the most intriguing area of chemistry and research [15-18]. Due to the macrocyclic effect, macrocyclic ligands are favoured over acyclic analogues [19] as they exhibit greater value of

thermodynamic stability, conformational restrictions, tiny hole for the metal ion's and preferred a well-organized ligand molecule that can be bent with little entropy and a powerful field simultaneously [20]. The structure and reactivity of a large number of marvellous naturally occurring compounds that are well known to undergo selective cation complexation are comparable to those of macrocyclic compounds [21-22]. These macrocyclic complexes have remarkable role in a number of biological activities namely antitumor, antidiabetic, antibacterial, anticancer, antifungal etc. [23-30]. The production of macrocyclic complexes can take place via template condensation process at the core of macrocyclic chemistry [31-32]. Thus, the synthesis of macrocyclic compounds has frequently employed the template process [33], wherein the transition metal ions are typically utilised as templates [34-35]. There are a great number and variety of transition metal complexes containing novel aza groups have been produced by varying cavity sizes, donor types, ring substituents several other factors [36-40]. Due to their biocidal effects, such as antiviral, anticarcinogenic, antifertility, antibacterial, and antifungal etc., transition metal macrocyclic complexes have tremendous interest in research [41–45]. On the basis of reported research work, approximately daily dietary intake of chromium as 25 μg for women and 35 μg for males plays a crucial role for proper functioning of the brain and heart, to regulate blood sugar, and to breakdown of sugars and lipids. Metal ions as a combining component with biological systems play crucial role in various biological processes and used to treat various diseases. Numerous naturally occurring complexes (Hb, chlorophylls, vitamin B12, etc.) having metals as a central atom have been discovered to perform important roles in a variety of biological processes [46]. In this study, we provide the findings from the synthesis, physicochemical analysis, and numerous biological

activities of the Cr(III) complexes derived from 4-aminobenzoic hydrazide and 2-aminobenzhydrazide with α -diketones i.e. isatin and 5-chloroisatin.

II. EXPERIMENTAL

2.1 Materials and methods

In the synthesis process, all of the used chemicals and solvents were taken in pure form. Chromium (III) acetylacetonate ($C_{15}H_{21}CrO_6$) metal salt was of Merck and ligand precursors namely 5-chloroisatin, 2-aminobenzhydrazide, 4-aminobenzoic hydrazide and isatin were of Sigma Aldrich. Reaction was carried out in methanolic medium and methanol solvent was purified before use.

2.2 Synthesis of the complexes

All the complexes were produced via [2+2] cyclocondensation reaction of 4-aminobenzoic hydrazide or 2-aminobenzhydrazide with isatin or 5-chloroisatin in methanol solvent in the existence of chromium acetylacetonate metal salt as template. For the synthesis of complex (1), in a round bottom flask 0.2 g. of 2-aminobenzhydrazide in 15-20 mL methanolic solution was taken and started to reflux on a magnetic stirrer. This solution was added drop by drop to the previous solution with continuous stirring. On the other hand (0.206 g) of isatin was also dissolved in the methanol (15-20 mL). To this mixture, methanolic solution of chromium acetylacetonate (0.0433 g. in 15-20 mL methanol) was added drop by drop with continuous stirring

and then after whole mixture was refluxed for 6-7 hours [47]. After refluxing, the mixture was kept to cool for overnight then it settled down and coloured precipitation was produced. Same process was opted for the synthesis of complexes (2), (3) and (4) with the change of their respective molar masses. Both ligands and salt of the metal were taken in [2:2:1] molar ratios respectively as shown in scheme (1)&(2).

2.3 Isolation of the complexes

The coloured precipitate of the complexes was filtered by the help of suction pump, washed many times with methanol and dried in vacuo. The complexes had a yield in between 70 and 80%.

2.4 Characterization and physicochemical properties of the complexes

Solubility of the synthesized complexes was found in DMF and DMSO. The produced complexes were yellow, orange and light orange coloured solid. Decomposition temperature of these complexes was found in the range 240 to 259 °C. Magnetic moments of these complexes were obtained in the range of 3.87 to 4.30 B.M. at room temperature, which indicated the presence of three unpaired electrons in chromium metal ion. In the complex, presence of metal was determined by the help of previously reported articles [48]. The SEM images of the synthesized complexes (1)&(3) respectively, as shown in (Fig. 1), demonstrated needle shaped structure of complexes with unordered distribution.

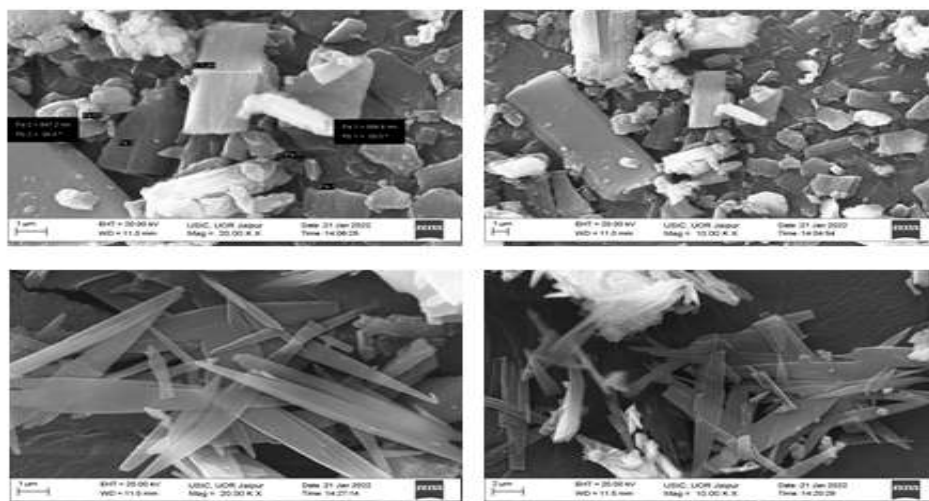


Fig. 1 SEM images of the complexes (1)&(3)

Scheme 1 Synthesis of complexes **1&2** derived by the reaction of 2-aminobenzhydrazide with isatin or 5-chloroisatin respectively

Scheme 2 Synthesis of complexes **3&4** derived by the reaction of 4-aminobenzoic hydrazide with isatin or 5-chloroisatin respectively

III. RESULT AND DISCUSSION

Fresh series of complexes of chromium having type $[\text{Cr}(\text{C}_{30}\text{H}_{20}\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ and $[\text{Cr}(\text{C}_{30}\text{H}_{18}\text{Cl}_2\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ was synthesized as shown in **scheme 1** & **2** for complexes (**1 to 4**). Each compound was found soluble and stable in DMSO and DMF and all were air stable. Based on the results of SEM, NMR, UV/Vis, IR, and mass spectrometry, it was assumed that the complexes formed.

NMR spectra

The ^1H NMR and ^{13}C NMR spectra of complexes, which were taken at room temperature in DMSO- d_6 solution, showed no signals of the primary amine protons. The ^{13}C NMR signals gave characteristic of carbon of the macrocyclic skeleton and ^1H NMR gave types of the protons. Appropriate positions of the carbon and hydrogen corresponding to the proposed structure were selected after analysing the spectral peaks. The ^1H NMR and ^{13}C NMR spectrum of the $[\text{Cr}(\text{C}_{30}\text{H}_{20}\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ complex shown in **Fig. 3** and **Fig. 4** respectively are justifying skeleton

of the compound as proposed in **Complex-3**. Two Singlet signal appeared at 5.94 ppm due to (aromatic ring connected -NH-) protons. Multiple signals at the range from 7.30-7.95 ppm are characteristic of (aromatic =CH) protons. The signals in the ^{13}C NMR range from 113.35 to 152.15 ppm are classified as aromatic =C-, whereas the aliphatic C=N and C=O signals showed at 150.21 and 163.82 ppm, respectively. The ^1H NMR and ^{13}C NMR spectra shown in **Fig. 5** and **Fig. 6** respectively were obtained for complex for **Complex- 1**. Two singlet signals appeared at 6.20 ppm due to (aromatic ring connected -NH-) protons. Multiple signals at the range from 7.19-7.95 ppm are characteristics of (aromatic =CH) protons. Multiple singlet signals appeared in between 113.35-151.86 ppm range are assigned to =C- aromatic carbon. Two doublet signals appeared at 125.38 and 127.79 ppm also assigned to aromatic =C- carbon. Two singlet signals at 143.59 ppm and 143.23 ppm appeared for -C=N carbon and C=O carbon at 167.97 ppm. Signal appeared at 151.80 ppm due to -N= connected aromatic carbon [49].

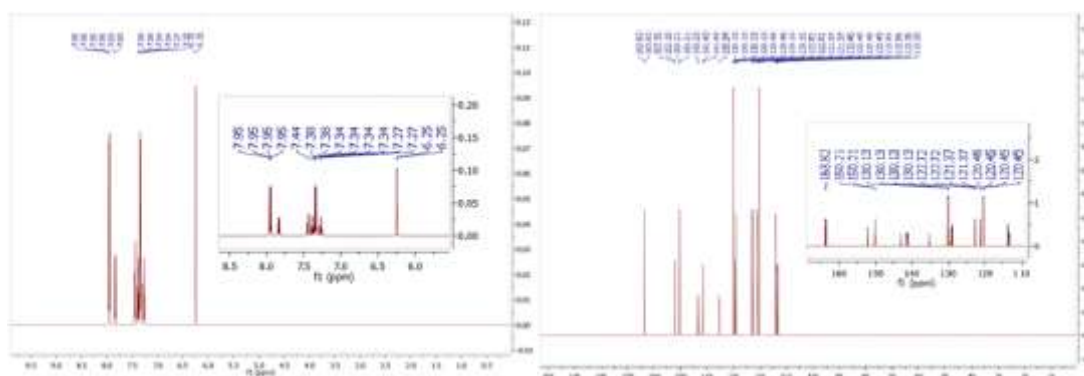


Fig. 3 Fig. 3

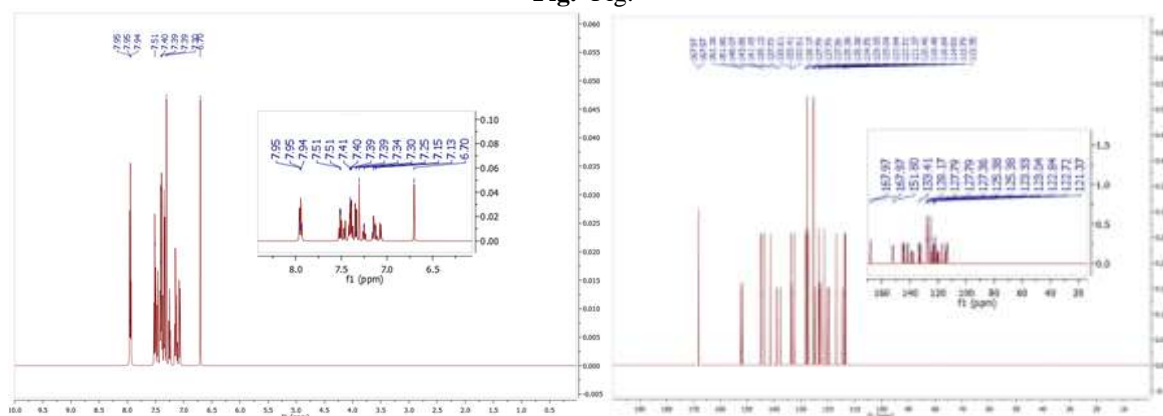


Fig. 4 Fig. 4

IR spectra

Two bands that were present in the spectrum of 4-aminobenzoic hydrazide (at 3350 cm^{-1} and 3390 cm^{-1}) and in the spectrum of 2-aminobenzhydrazide (at 3350 cm^{-1} and 3385 cm^{-1}) were discovered to be missing from all of the complexes spectra, corresponded to the $\nu(\text{NH}_2)$ group. At 1650 cm^{-1} , a prominent absorption band was seen associated with the $\nu(\text{C}=\text{O})$ downfield signal due to neighbouring electron donor group [50]. In all complexes, absence of individual ($>\text{C}=\text{O}$) and (NH_2) bands at the reported range and presence of new band at ≈ 1590 to 1690 cm^{-1} attributed to ($>\text{C}=\text{N}$) group, representing the condensation reaction of diketone with diamine. The benzene ring's $\nu(\text{C}=\text{C})$ aromatic

stretching vibrations were responsible for the many forms of absorption bands in between the range $1400\text{--}1588\text{ cm}^{-1}$ [51-52]. The bands which were in the range of $740\text{--}785\text{ cm}^{-1}$ may correspond to the $\nu(\text{C-H})$ out of the plane bending to the aromatic ring [53]. The IR spectra of all the complexes showed absorption bands at $1408\text{--}1440\text{ cm}^{-1}$, $1290\text{--}1320\text{ cm}^{-1}$, and $1010\text{--}1030\text{ cm}^{-1}$ that indicated that N atoms are unidentifiably coupled to the core metal ion [54]. It can be shown that the coordination of the acetylacetonate group with the central metal ion was unidentate because $390\text{--}370\text{ cm}^{-1}$ is the difference between ($\nu_{\text{as}} - \nu_{\text{s}}$) that is greater than 144 cm^{-1} [55]. IR spectra of the synthesized compounds (**Complex-3** and **Complex-4**) are represented in Fig. 1.

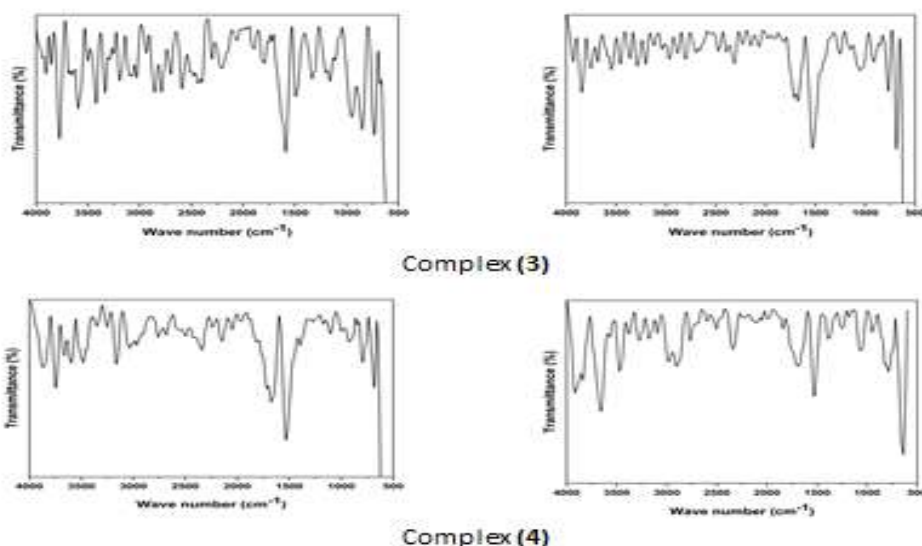


Fig. V IR spectra of the $[\text{Cr}(\text{C}_{30}\text{H}_{20}\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ and $[\text{Cr}(\text{C}_{30}\text{H}_{18}\text{Cl}_2\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ complexes

Mass spectrometry

The mass spectra of $[\text{Cr}(\text{C}_{30}\text{H}_{20}\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ and $[\text{Cr}(\text{C}_{30}\text{H}_{18}\text{Cl}_2\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ complexes have been recorded and shown in Fig. V and Fig. A respectively. The compound $[\text{Cr}(\text{C}_{30}\text{H}_{20}\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ exhibited the molecular ion peak (M^+) at $m/z = 524.17$. The development of the macrocyclic structure $[\text{Cr}(\text{C}_{30}\text{H}_{20}\text{N}_8\text{O}_2)]$ was confirmed by this peak. Additional peaks at $m/z = 521, 522.90, 523, 525.17$

etc. may be corresponded to several fragments. The complex $[\text{Cr}(\text{C}_{30}\text{H}_{18}\text{Cl}_2\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ displayed the molecular ion peak (M^+), at m/z was 592.09. The macrocyclic structure with the chemical formula $[\text{Cr}(\text{C}_{30}\text{H}_{18}\text{Cl}_2\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$ was confirmed by this peak. Additional peaks at $m/z = 425.31, 445, 500, 593.10, 595.09$ etc. may be corresponded to several fragments. Intensity of the peaks explains the stability of various fragments [56].

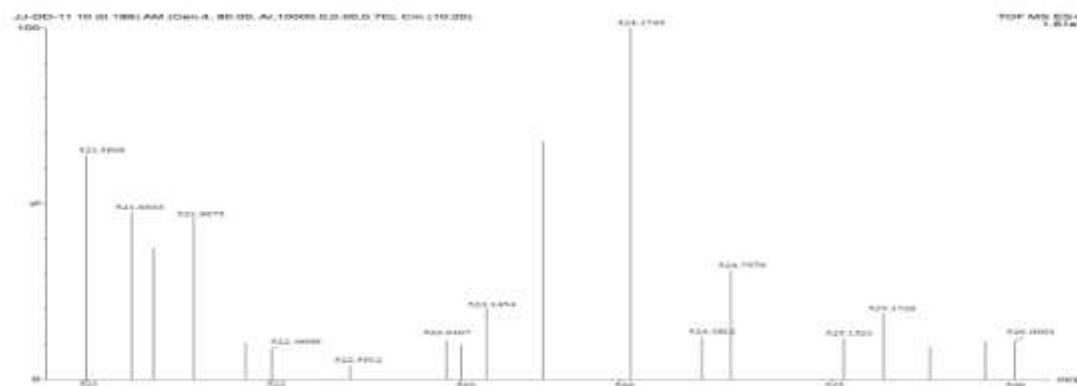


Fig. V Mass spectrum of the complex $[\text{Cr}(\text{C}_{30}\text{H}_{20}\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$

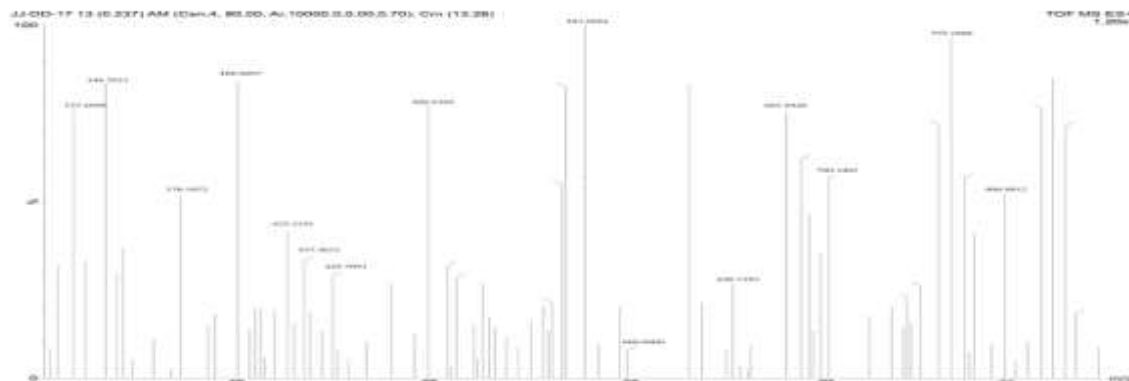


Fig. 8 Mass spectrum of the complex $[\text{Cr}(\text{C}_{30}\text{H}_{18}\text{Cl}_2\text{N}_8\text{O}_2)](\text{C}_5\text{H}_7\text{OO}^-)_3$

UV Spectra

UV-Vis spectra of respective complexes indicated strong and weak peaks around 255-257 nm and 350-400 nm respectively revealed the π to π^* and n to π^* transitions. π to π^* transitions are associated longer wavelength (255-257 nm) than normal range of transitions (180-200 nm) because of extended conjugation and polar solvent. When

conjugation increases than energy gap in between HOMO and LUMO decreases so less energy is required for transition, results bathochromic shift. Effect of polar solvents on orbitals follow the order of $n > \pi^* > \pi$, so in the presence of polar solvent wavelength of π to π^* transitions increases while n to π^* decreases, so earlier transitions show bathochromic while later shows hypsochromic shift Fig. 9.

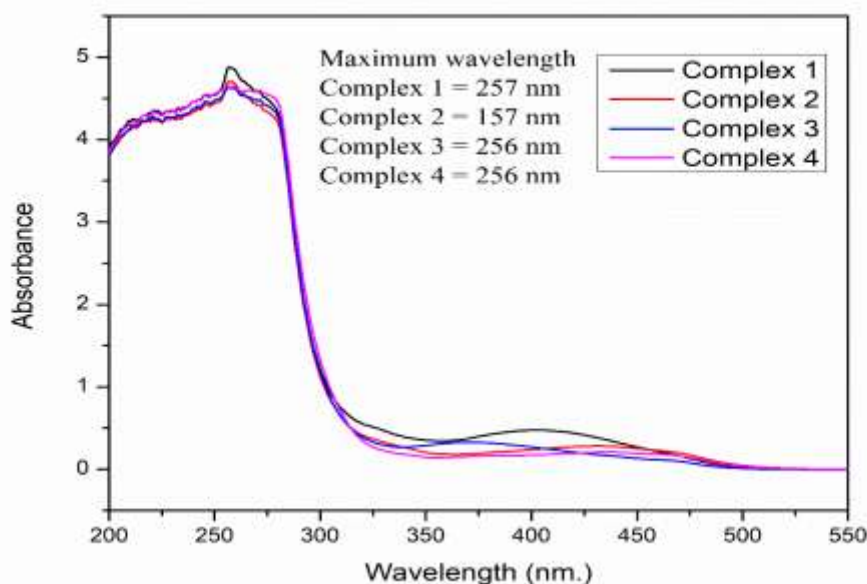


Fig. 9 UV-Vis spectra of the synthesized macrocyclic complexes

IV. BIOLOGICAL ACTIVITIES

Antibacterial and Antifungal assay by well diffusion method: -

In-vitro antibacterial assays were conducted by agar well diffusion method [57]. 10% Dimethyl sulfoxide (DMSO) was used to dilute

the various samples, and three different concentrations of each drug (50 mg/mL, 100 mg/mL, and 200 mg/mL) were taken. To inoculate the test microorganisms, nutrient agar medium (for antibacterial) and potato dextrose agar (PDA) medium (for antifungal) was placed in clean petri

dishes. The spreader was used to evenly distribute the inoculum, and the dish was let to stand for 30 minutes. The seeded agar plates were constructed with 6 mm-diameter wells. The control well was additionally set up at the same distance. The prearranged wells of the seeded plates were filled with various concentrations of each sample and the standard medication (30 µl). These plates were incubated at 37°C for 24 hours. Inhibition zone (IZ) measurements surrounding each prepared well

were used to determine the antifungal and antibacterialspectrum of the test material. Comparisons were made between the test sample's diameter of the inhibitory zone and commercially available control antibiotic Cipro (1 mg/mL) for antibacterial whereas Ketoconazole (1 mg/mL) for antifungal. Results of antibacterial (against *S. aureus* and *E. coli*) and antifungal (against *Alternaria alternative* and *Fusarium solani*) assay are given in **Table 1**, and **Fig. 1**.

Table 1 Antibacterial and Antifungal activity of complexes

Sample's Name	Anti-Bacterial Action						Anti-Fungal Action					
	E. coli			S. aureus			Alternaria alternative			Fusarium solani		
	Concentration						Concentration					
	50	100	200	50	100	200	50	100	200	50	100	200
	IZ	IZ	IZ	IZ	IZ	IZ	IZ	IZ	IZ	IZ	IZ	IZ
Standard	25	27	32	27	31	36	32	36	41	28	30	34
Complex 1	NA	1	3	3	4	6	8	13	17	5	8	10
Complex 2	5	8	11	2	4	5	7	10	14	5	8	10
Complex 3	NA	NA	3	3	6	9	NA	3	5	2	4	7
Complex 4	2	3	4	4	5	8	6	9	13	5	9	13

Note: NA= Non active, IZ= Inhibition zone

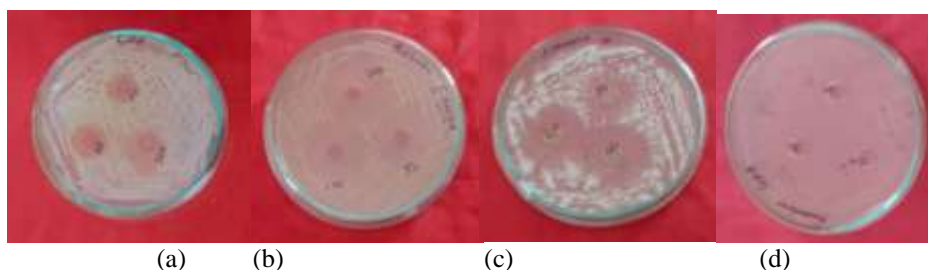


Fig. 1 (a, b, c and d) **Figure a & b (complex-2)** show antibacterial activity against *E. coli*, *S. aureus* respectively, and **figure c & d (complex-1)** show antifungal activity against *Alternaria alternate*, *Fusarium solani* respectively

Antioxidant activity: -

The DPPH free radical scavenging activity to assess the extract's antioxidant properties was done by using 1, 1-diphenyl 2-picryl-hydrazil (DPPH) as proposed by Alothman et al. [58]. The

mixing of 100 µL aliquot from compounds in different concentrations (20-100 mg/L) was done in 3.9 mL taken from 0.1 mM DPPH (methanolic) solution. Then blend was subjected to vortex and left for incubation in the dark for half an hour. Its

OD was calculated at 515 nm while methanol was used as blank.

The % Radical scavenging activity = $[(A_0 - A_c) / A_0] \times 100$

Where A_0 = absorbance of the control & A_c = sample's absorbance at C concentration

Linear plot of concentration versus % inhibition was plotted and by this IC_{50} values were

determined. The antioxidant potential of each extract was showed in form of IC_{50} (stated as the quantity of concentration necessary to prevent DPPH radical development by 50%), find out with the help of inhibition curve. Results towards antioxidant action of complexes are shown in **Table 2** and **Fig. 11**.

Table 2 Antioxidant activity

Name of Compound	Concentration(mg/L)					Regression Equation	IC ₅₀
	20	40	60	80	100		
Complex 1	26.02	28.94	30.99	34.5	36.69	Y=0.1345x+23.358	198.08
Complex 2	29.54	31.66	32.47	34.61	36.67	Y=0.0861x+27.827	257.52
Complex 3	17.54	21.05	22.95	24.56	26.6	Y=0.1082x+16.051	313.76
Complex 4	22.22	22.95	26.6	29.38	32.45	Y=0.1345x+18.653	233.06
Ascorbic acid	48.09	61.11	70.46	72.8	74.85	Y=0.3261x+45.899	12.57

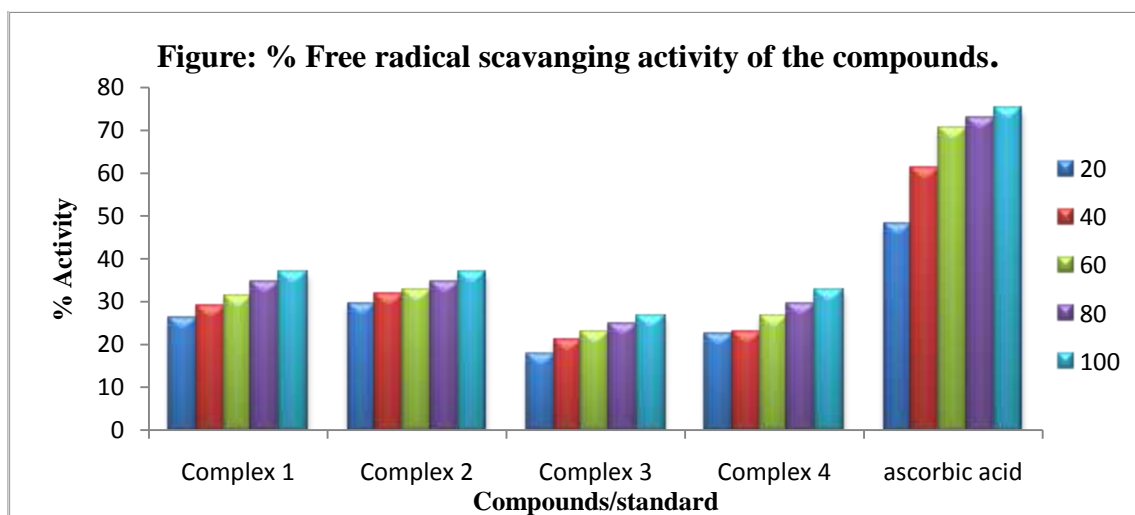


Fig. 11 Comparison of the percentage free radical scavenging activity of the complexes with standard ascorbic acid

V. CONCLUSION

Several techniques, including SEM, NMR, mass spectrometry, IR, and UV/Vis, were utilised to construct and characterise Cr(III) complexes with hexadentate macrocyclic ligands. The polarity of the metal ion was reduced by the coordination/chelation with N-donor ligand within the whole

chelate ring, because metal ion shared its partial positive charge with donor group. Against E. coli, **complex-2** exhibited strong antibacterial activity. All complexes showed activity against S. aureus but **complex-3 & 4** showed satisfactory activity. **Complex-1&2** showed good antifungal activity against Alternaria alternate were

as **complex-2&4** showed good antifungal activity against *Fusarium solani*. **Complex-1&Complex-2** showed good antioxidant activity but less than ascorbic acid. Solubility, magnetic moments, decomposition temperature etc. properties also have been discussed.

ACKNOWLEDGEMENTS

The Department of Chemistry, University of Rajasthan, Jaipur, India, is acknowledged for its contributions to the work in all forms. MNIT (Jaipur) for recording NMR and Mass spectrometry, CU (Haryana) for UV/Vis, Biomitra Life Sciences pvt. Ltd. (Jaipur) for biological activities.

REFERENCES

- [1]. Chandra, S; Tyagi, M; and Agarwal, S., 2010, "Synthesis and characterization of a tetraaza macrocyclic ligand and its cobalt (II), nickel (II) and copper (II) complexes," *J. Serbian Chem. Soc.*, **75(7)**, 935-941. <http://dx.doi.org/10.2298/JSC090804069C>
- [2]. Tyagi, M; Chandra, S; and Choudhary, S-K., 2011, "Tetraaza macrocyclic complexes: Synthesis, spectral and antifungal studies," *J. Chem. Pharm. Res.*, **3(1)**, 56-63.
- [3]. Khatun, M; Ghorai, P; Mandal, J; Chowdhury, S-G; Karmakar, P; Blasco, S; Espana, E-G; and Amrita, S., 2023, *ACS Omega*, **8(8)**, 7479-7491. DOI: 10.1021/acsomega.2c06549
- [4]. Chandra, S; Qanungo, K; and Sharma, S-K., 2012, "New hexadentate macrocyclic ligand and their copper (II) and nickel (II) complexes: spectral, magnetic, electrochemical, thermal, molecular modeling and antimicrobial studies," *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, **94**, 312-317. <https://doi.org/10.1016/j.saa.2011.12.028>
- [5]. Nirmala, G; Rahiman, A-K; Sreedaran, S; Jegadeesh, R; Raaman, N; and Narayanan, V., 2011, "New 14-membered trans-di-substituted 'tet-a' macrocycles and their copper (II) and nickel (II) complexes: Spectral, magnetic, electrochemical, crystal structure, catalytic and antimicrobial studies," *J. Mol. Struct.*, **989(1-3)**, 91-100. DOI: 10.1016/j.molstruc.2011.01.010
- [6]. Kanaoujiya, R; Singh, D; Minocha, T; Yadav, S-K; and Srivastava, S., 2022, "Synthesis, characterization of ruthenium (III) macrocyclic complexes of 1, 4, 8, 11-tetraazacyclotetradecane (cyclam) and in vitro assessment of anti-cancer activity," *Materials Today: Proceeding*, **65**, 3143-3149. DOI: 10.1016/j.matpr.2022.05.354
- [7]. Ilhan, S; and Temel, H., 2007, "Synthesis and characterization of a new macrocyclic Schiff base derived from 2, 6-diaminopyridine and 1, 10-bis (2-formylphenyl)-1, 4, 7, 10-tetraoxadecane and its Cu (II), Ni (II), Pb (II), Co (III) and La (III) complexes," *Transit. Met. Chem.*, **32**, 1039-1046. <https://doi.org/10.1007/s11243-007-0276-5>
- [8]. Aly, A-A; Abdallah, E-M; Ahmed, S-A; Rabee, M-M; and Brase, S., 2023, "Transition Metal Complexes of Thiosemicarbazides, Thiocarbohydrazides, and Their corresponding Carbazones with Cu(I), Cu(II), Co(II), Ni(II), Pd(II), and Ag(I)—A Review," *Molecules*, **28**, 1808. <https://doi.org/10.3390/molecules28041808>
- [9]. Ali, V; Singh, P; Jain, V; and Tripathi, J., 2019, *Saudi Chem. Soc.*, **23**, 52-60. <https://doi.org/10.1016/j.jscs.2018.04.005>
- [10]. Khalid, S; Sumrra, S-H; and Chohan, Z-H., 2020, *Sains Malaysiana*, **49(8)**, 1891-1904. <http://dx.doi.org/10.17576/jsm-2020-4908-11>
- [11]. Li, J; Guo, L; and Huo, H., 2023, "Preparation of nickel catalysts bearing Schiff base macrocycles and their performance in ethylene oligomerization," *Transit Met Chem.*, <https://doi.org/10.1007/s11243-023-00527-w>
- [12]. Fierro, C-M; Smith, P-D; and Light, M-E., 2023, *Polyhedron*, **230**. <https://doi.org/10.1016/j.poly.2022.116222>
- [13]. Schuman, A-J; Raghavan, A; Banziger, S-D; Song, Y; Hu, Z-B; Mash, B-L; Williams, A-L; and Ren, T., 2021,

- “Macrocyclic Chromium (III) Catecholate Complexes,” *Inorg. Chem.*, **60(7)**, 4447-4455.
- [14]. Kostova, I., 2023, *Inorganics*, **11(2)**, 56. <https://doi.org/10.3390/inorganics11020056>
- [15]. Chandra, S; Gupta, L-K;and Agrawal, S.,2007, “Modern spectroscopic and biological approach in the characterization of a novel 14-membered [N 4] macrocyclic ligand and its transition metal complexes,” *Transit. Met. Chem.*, **32**, 240-245.<https://doi.org/10.1007/s11243-006-0155-5>
- [16]. Singh, D-P; Kumar, K;Dhiman, S-S;and Sharma, J., 2010,“Antibacterial and antifungal studies of macrocyclic complexes of trivalent transition metal ions with their spectroscopic approach,” *J. Enzyme Inhib. Med. Chem.*, **25(1)**, 21-28.<https://doi.org/10.3109/14756360902932750>
- [17]. Rathi, P; Singh, D-P; and Surain, P., 2015,“Synthesis, characterization, powder XRD and antimicrobial-antioxidant activity evaluation of trivalent transition metal macrocyclic complexes,” *C R Chim.*, **18(4)**, 430-437.<https://doi.org/10.1016/j.crci.2014.08.002>
- [18]. Lash, T-D.,2023, *Molecules*, **28(3)**, 1496. <https://doi.org/10.3390/molecules28031496>
- [19]. Cabbiness, D-K;and Margerum, D-W.,1969, “Macrocyclic effect on the stability of copper (II) tetramine complexes,” *J. Am. Chem. Soc.*, **91(23)**, 6540-6541. DOI: 10.1021/Ja01051A091
- [20]. Fabbrizzi, L;Paoletti, P; and Clay, R-M., 1978, “Microcalorimetric determination of the enthalpy of a slow reaction: destruction with cyanide of the macrocyclic (1, 4, 8, 11-tetraazacyclotetradecane) nickel (II) ion,” *Inorg. Chem.*,**17(4)**, 1042-1046.<https://doi.org/10.1021/ic50182a048>
- [21]. Lindoy, L-F., 1990,“Heavy Metal Chemistry of Mixed Donor Macrocyclic Ligands: Strategies for Obtaining Metal Ion Recognition,” *J Incl Phenom Macrocycl Chem.*, 171-183.
- [22]. Chandra, S;Qanungo, K;and Sharma, S-K.,2012, “New hexadentate macrocyclic ligand and their copper (II) and nickel (II) complexes: spectral, magnetic, electrochemical, thermal, molecular modeling and antimicrobial studies,” *Spectrochim. Acta A Mol. Biomol. Spectrosc.*,**94**, 312-317.Doi: 10.1016/j.saa.2011.12.028
- [23]. Nath, R;Pathania, S;Grover, G; and Akhtar, MJ.,2020, *J. Mol. Struct.*,<https://doi.org/10.1016/j.molstruc.2020.128900>
- [24]. Mishra, P;Sethi, P;and Kumari, A., 2022, “Emerging applications and host- guest chemistry of synthetic macrocycles,” *Res. J. Chem. Environ.*, **26(7)**, 153-157.DOI:10.25303/2607rjce153167
- [25]. Lahari, K;and Sundararajan, R., 2020, *J. Chem. Sci.*, **132**, 94. <https://doi.org/10.1007/s12039-017-1398-8>
- [26]. Yang, J;Dai, D;Cai, Z;Liu, Y-Q;Qin, J-C;Wang, Y;and Yang, Y-W.,2021., *Acta Biomater.*,**134**, 664–73. <https://doi.org/10.1016/j.actbio.2021.07.050>.
- [27]. Liu, H;Yang, J;Yan, X;Li, C;Elsabhy, M;Yang, Y-W;and Gao,H-A., 2021, *J. Mater. Chem. B.*, 2021, **9**, 9594–9605.DOI: 10.1039/d1tb02134f
- [28]. Dai, D;Yang, J;and Yang, W., 2022,*Chem. Eur. J.* DOI: 10.1002/chem.202103185 (Invited Contribution).
- [29]. Dileepan, A-G-B;Prakash, T-D;Kumar, A-G;Rajam, P-S;Dhayabaran, V-V;and Rajaram, R-J., 2018,*Photochem. Photobiol.* BBiol. Doi:10.1016/j.jphotobiol.2018.04.029
- [30]. Bugalia, S;Dhayal, Y;Sachdeva, H;Kumari, S; Atal, K;Phageria, U; Saini, P;and Gurjar, O-P., 2023,“Review on Isatin- A Remarkable Scaffold for Designing Potential Therapeutic Complexes and Its Macrocyclic Complexes with Transition Metals.” *JIOPM.* <https://doi.org/10.1007/s10904-023-02666-0>
- [31]. Curtis, N-F.,“Macrocyclic coordination compounds formed by condensation of metal-amine complexes with aliphatic carbonyl compounds,” 1968, *Coord. Chem. Rev.*,**3(1)**, 3-

47. [https://doi.org/10.1016/S0010-8545\(00\)80104-6](https://doi.org/10.1016/S0010-8545(00)80104-6)
- [32]. Shakir, M;Bano, N;Rauf, M-A; andOwais, M., 2017,*J. Chem. Sci.*,**129(12)**, 1905–1920.<https://doi.org/10.1007/s12039-017-1398-8>
- [33]. Niasari, M-S;and Davar, F., 2006,“In situ one-pot template synthesis (IOPTS) and characterization of copper (II) complexes of 14-membered hexaaza macrocyclic ligand “3, 10-dialkyl-dibenzo-1, 3, 5, 8, 10, 12-hexaazacyclotetradecane”, *Inorg. Chem. Commun.*, **9(2)**, 175-179.DOI:10.1016/j.inoche.2005.10.028
- [34]. Prasad, R-N;Mathur, M;and Upadhyay, A., 2007, “Synthesis and spectroscopic studies of Cr (III), Fe (III) and Co (II) complexes of hexaazamacrocycles,” *J. Indian Chem. Soc.*, **84(12)**, 1202-1204.
- [35]. Kamboj, M; Singh, D-P; Singh, A-K;and Chaturvedi, D., 2020,“Molecular modeling, in-silico docking and antibacterial studies of novel template wangled macrocyclic complexes involving isatin moiety,” *J. Mol. Struct.*,**1207**, 127602.DOI:10.1016/j.molstruc.2019.127602
- [36]. Chandra, S;Qanungo, K;and Sharma, S-K., 2012, “Newhexadentate macrocyclic ligand and their copper (II) and nickel (II) complexes: spectral, magnetic, electrochemical, thermal, molecular modeling and antimicrobial studies,” *Spectrochim. Acta Mol. Biomol. Spectrosc.*,**94**, 312-317.Doi: 10.1016/j.saa.2011.12.028
- [37]. Martin, J-G; Wei, R-M;and Cummings, S-C.,1972, “Copper (II) complexes with 13-membered macrocyclic ligands derived from triethylenetetramine and acetylacetone,” *Inorg. Chem.*, **11(3)**, 475-479.
- [38]. Holtman, M-S;and Cummings, S-C.,1996, “Macrocyclic nickel (II) complexes with new, dimethyl-substituted 13-and 14-membered tetraaza ligands,” *Inorg. Chem.*, **15(3)**, 660-665.
- [39]. Roberts, G-W; Cummings, S-C;and Cunningham, J-A.,1976, “Synthesis and characterization of low-spin cobalt (II) complexes with macrocyclic tetraaza ligands. The crystal structure of [Co ([14
- dieneN4). cntdot. H₂O(PF₆)₂,” *Inorg. Chem.*, **15(10)**, 2503-2510.
- [40]. Coltrain, B-K;and Jackels, S-C.,1981, “Coordination chemistry of a copper (II) tetraimine macrocycle: four-, five-, and six-coordinate derivatives and reduction transmetalation to the zinc (II) complex,” *Inorg. Chem.*, **20(7)**, 2032-2039. <https://doi.org/10.1021/ic50221a021>
- [41]. Chandra, S;and Pundir, M.,2008, “Spectroscopic characterization of chromium (III), manganese (II) and nickel (II) complexes with a nitrogen donor tetradentate, 12-membered azamacrocyclic ligand,” *Spectrochim. Acta A Mol. Biomol. Spectrosc.*,**69(1)**, 1-7.
- [42]. Prasad, R-N;and Upadhyay, A.,2006, “Chromium (III), iron (III) and cobalt (II) complexes of 14-and 16-membered tetraazamacrocycles,” *J. Indian Chem. Soc.*,**83(9)**, 857-860.
- [43]. Chandra, S; Gupta, R; Gupta, N;and Bawa, S-S., 2006, “Biologically relevant macrocyclic complexes of copper spectral, magnetic, thermal and antibacterial approach,” *Transit. Met. Chem.*, **31(2)**, 147-151.
- [44]. Chandra, S; Gupta, L-K;and Agrawal, S., 2007,“Synthesis spectroscopic and biological approach in the characterization of novel [N 4] macrocyclic ligand and its transition metal complexes, *Transit. Met. Chem.*, **32**, 558-563.
- [45]. Gammal, O-A-E;Brashy, S-A-E;and El-Reash,G-M-A-E., 2020,“Macrocyclic Cr³⁺, Mn²⁺ and Fe³⁺ complexes of a mimic SOD moiety: Design, structural aspects, DFT, XRD, optical properties and biological activity,” *Appl. Organomet. Chem.*, **34(4)**, 5456. DOI:10.1002/aoc.5456
- [46]. Gurjar, O-P;Kumari, S;Saini, P;and Bugalia, S.,2022, “A short review on biological applications of macrocyclic complexes of chromium,”*J. Chil. Chem. Soc.*, **67**, N^o4.
- [47]. Zafar, H;Kareem, A;Sherwani, A;Mohammad, O;and Khan, T-A.,2015, “Synthesis, characterization and biological studies of homo and hetero-binuclear 13-membered pentaaza bis (macrocyclic) complexes,”*J. Mol. Struct.*,**1079**, 337–346.

- <http://dx.doi.org/10.1016/j.molstruc.2014.08.036>
- [48]. Singh, D-P;Kumar, K;Dhiman, S-S;and Sharma, J., 2010,“Antibacterial and antifungal studies of macrocyclic complexes of trivalent transition metal ions with their spectroscopic approach,”*J Enzyme Inhib Med Chem.*,**25(1)**, 21–28.DOI: 10.3109/14756360902932750
- [49]. Zafar, H;Kareem, A;Sherwani, A;Mohammad, O;and Khan, T-A., 2015,“Synthesis, characterization and biological studies of homo and hetero-binuclear 13-membered pentaazabis (macrocyclic) complexes,”*J. Mol. Struct.*,**1079**, 337–346.<http://dx.doi.org/10.1016/j.molstruc.2014.08.036>
- [50]. Singh, D-P; Kumar, R;and Singh, J., 2009,“Antibacterial activity and spectral studies of trivalent chromium, manganese, iron macrocyclic complexes derived from oxalyldihydrazide and glyoxal, *J Enzyme Inhib Med Chem.*,**24(3)**, 883-889.
- [51]. Prasad, R-N;Mathur, M;and Upadhyay, A.,2007, “Synthesis and spectroscopic studies of Cr (III), Fe (III) and Co (II) complexes of hexaazamacrocycles,” *J. Indian Chem. Soc.*, **84(12)**, 1202-1204.
- [52]. Costamagna,J;Ferraudi, G;Villagran, M;and Wolcan, E.,2000, “Ligand luminescence and photoinduced charge separation in bis (naphthalene) substituted fourteen-membered tetraazamacrocyclic complexes of Cu II and Ni II,” *J. Chem. Soc., Dalton Trans.*,**(15)**, 2631-2637.<https://doi.org/10.1039/B002829K>
- [53]. Singh, D-P;Shishodia, N;Yadav, B-P;and Rana, V-B., 2004“Trivalent transition metal ion directed macrocycles,” *J. Indian Chem. Soc.*, **81(4)**, 287-290.
- [54]. Chandra, S;and Gupta, L-K; Electronic, E-P-R., 2005, “ magnetic and mass spectral studies of mono and homo-binuclear Co (II) and Cu (II) complexes with a novel macrocyclic ligand,” *Spectrochim. Acta Mol. Biomol. Spectrosc.*,**62(4-5)**, 1102-1106.Doi: 10.1016/j.saa.2005.04.007
- [55]. Nakamoto,K., 2009,“Infrared and Raman spectra of inorganic and coordination compounds, part B: applications in coordination, organometallic, and bioinorganic chemistry,”John Wiley & Sons..
- [56]. Singh, D-P;Kumar, K;Dhiman, S-S; andSharma, J., 2009,“Biologically active macrocyclic complexes derived from diamionaphthalene and glyoxal: Template synthesis and spectroscopic approach,”*J. Enzyme Inhib. Med.*, **24(3)**, 795–803. DOI: 10.1080/14756360802397179
- [57]. Pérez, C;and Anesini, C.,1994, “Antibacterial activity of alimentary plants against *Staphylococcus aureus* growth, *Am. J. Chinese Med.*,**22(02)**, 169-174.
- [58]. Alothman,;Bhat, R;Karim, A-A., 2009,“Antioxidant capacity and phenolic content of selected tropical fruits from Malaysia, extracted with different solvents,” *Food Chem.*, **115(3)**, 785-788.