

Development and Validation of Stability Indicating Hptlc Method for Estimation of Stiripentol

Mrinalini C.Damle^{a*}, Akash R. Bhusari^b, and Raj Y. Dhodi^c

^aDepartment of Quality Assurance, AISSMS College of Pharmacy, Kennedy Road, near RTO Pune, 411 001, Maharashtra, India E-mail: damle_mc@aissmscop.com Contact No: 9860230912

^{b,c}Master of Pharmacy, Department of Pharmaceutical Quality Assurance, AISSMS College of Pharmacy, Kennedy Road, Near RTO Pune, 411 001, Maharashtra, India

*Corresponding Author: Mrinalini C. Damle, E-mail: damle_mc@aissmscop.com, Department of Quality Assurance, AISSMS College of Pharmacy, Kennedy Road, near RTO Pune, 411 001, Maharashtra, India

Submitted: 25-03-2024

Accepted: 05-04-2024

ABSTRACT: Stiripentol is an antiepileptic drug that is used for the treatment of Dravet syndrome. A stability-indicating HPTLC method has been established & validated for the determination of pestilential. The stationary phase used was Silica Gel plate 60F254 & mobile phase was selected as toluene: ethyl acetate (9.5:0.5 V/V). The R_f was found to be (0.46 ± 0.02). The developed stability-indicating method was validated for linearity, accuracy, precision, limit of detection, limit of quantitation, and robustness parameters after establishing stability by forced degradation study. Stiripentol was found to be sensitive to acidic as well as alkaline hydrolytic stress, and oxidative and thermal degradation conditions but no peak for degradation product was detected in spite of multiple wavelength scanning.

KEYWORDS: Stiripentol, High-Performance Thin Layer Chromatography, forced degradation, validation, Stability-indicating method.

I. INTRODUCTION

Chemically, pestilential is (1E)-1-(1,3-benzodioxol-5-yl)-4,4-dimethyl-pent-1-en-3-ol^[14]. It is soluble in methanol and practically insoluble in water at room temperature. Stiripentol, (Diacomit; Biocodex Inc.) has been approved by the European and Canadian marketing authorization for the treatment of Dravet syndrome as adjunctive therapy with clobazam and valproic acid not only in children but also during adolescence and adulthood when seizures are not adequately controlled with the association of these two medications^[1,2,4,18]. Dravet syndrome is also called severe myoclonic epilepsy of infancy which is progressive epileptic encephalopathy^[10,13,19]. Because of the inhibitory effect of pestilential on hepatic cytochrome-P450, its clinical development was delayed^[6]. Its

principal mechanism of action in the brain is GABA inhibitory neurotransmission and by blocking the GABA-transaminase activity it prevents the GABA metabolism^[16]. The literature search shows that there are reports of bioanalytical High-Performance Liquid Chromatography (HPLC) method^[12,15,17], "stability indicating method" by HPLC^[3] and HPTLC^[11]. After looking at the literature survey, it was concluded that there was work done on stability indicating method by HPTLC & HPLC method, but it is observed that there is a scope to optimize the HPTLC method in terms of the level of stress degradation. So, in this work by enhancing the stress condition parameters like strength of stress reagent and time of exposure, the method for stability study was well optimized. Hence the main aim of our study was to optimize the various stress conditions for pestilential and evaluate the optimized HPTLC method.

II. MATERIALS AND METHODS

Materials

The drug pestilential was received as a gift sample. All the chemicals used were of AR grade viz. methanol, toluene, ethyl acetate, dichloromethane, hydrochloric acid, sodium hydroxide, hydrogen peroxide.

Methods Instruments:

UV-spectral analysis was performed on UV-Visible spectrophotometer (Make-JASCO, Model-V730). CAMAG HPTLC system equipped with a Linomat 5 sample applicator operated under a gentle stream of nitrogen, coupled with a Hamilton microliter syringe (100 µL) was used for application purpose. CAMAG TLC SCANNER 3

was used for detection and densitometry scanning. Data acquisition was done by WINCATS software (version 1.4.3). Photostability study was performed in a photostability chamber (Make- NEWTRONIC, Model- NEC103RSPI).

Detection of wavelength

A solution of pestiential ($10 \mu\text{g mL}^{-1}$) was prepared using methanol and UV spectrum was recorded. It showed maximum absorbance at 262 nm. Spectrum is shown in (Fig.1)

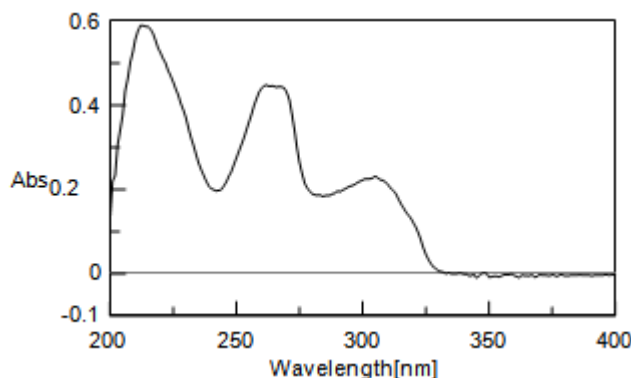


Fig.1: UV spectrum of pestiential in methanol ($10 \mu\text{g mL}^{-1}$)

III. PREPARATION OF STANDARD AND SAMPLE SOLUTION

For the preparation of standard solution, 10 mg of the drug was accurately weighed and transferred to the volumetric flask of 10 mL capacity. Then added some methanol and made up the volume after the drug was dissolved to obtain the solution of $1000 \mu\text{g mL}^{-1}$. A solution of strength $50 \mu\text{g mL}^{-1}$ was prepared by appropriate dilution & was used as a working solution. Stiripentol capsules and sachets are available as Diacomit in 250 & 500 mg strength. But the brand Diacomit was not available in local market. Hence, we prepared an excipient blend to which API was spiked. For the preparation of spiked blend, 125 mg starch & 125 mg lactose were mixed in the mortar and pestle. Then 250 mg of pestiential was mixed with the above excipients. From this spiked blend, 20 mg of blend (equivalent to 10 mg of drug) was accurately weighed and diluted to 10

mL with methanol to obtain a solution ($1000 \mu\text{g mL}^{-1}$). The solution was sonicated & then filtered using Whatman filter paper. It was diluted to get $50 \mu\text{g mL}^{-1}$ working solution.

Chromatographic Conditions

Chromatographic separation of pestiential drug was performed on Aluminium plates pre-coated with a band of 6 mm width using a $100 \mu\text{L}$ syringe with a Linomat applicator. The mobile phase was composed of toluene: ethyl acetate (9.5:0.5 V/V). A twin trough glass chamber ($10 \text{ cm} \times 10 \text{ cm}$) was used for linear ascending development of the TLC plate with 15 min saturation time, migration distance was 70 mm. Densitometric scanning was performed at 262 nm, operated by software, slit dimensions were $4 \times 0.45 \text{ mm}$. The standard densitogram of pestiential (250 ng band^{-1}) is shown in (Fig. 2).

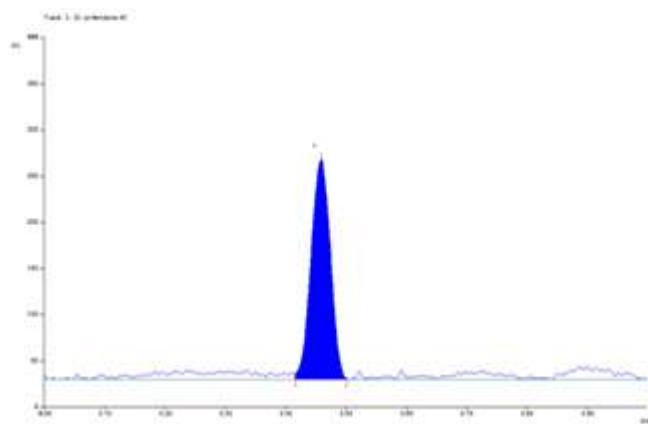


Fig.2:Representative densitogram of pestilential (250 ng band⁻¹, Rf = 0.46 ± 0.02)

Calibration curve preparation

The range selected was 250-1250 ng band⁻¹ to silicagel 60F₂₅₄. Samples were applied on the plates as determine the linear relationship between peak area and amount spotted.

IV. FORCED DEGRADATION STUDY

Various stress conditions have been applied to check the degradation^[7]. The conditions were hydrolysis under various pH viz. acid, alkali conditions; oxidative, thermal stress, photostability^[9] under UV and fluorescent light. The conditions were optimized to achieve degradation in the range of 10- 30 %.

Acid Hydrolysis:

For sample preparation, 1 mL of stock solution (500 µg mL⁻¹) was mixed with 1 mL 0.05N HCL and the volume was made up to 10 mL with methanol. The solution was kept for 20 min at room temperature. The resultant solution of 50 µg mL⁻¹ was applied to the TLC plate and the densitogram was shown in (Fig 3).

Alkali Hydrolysis

For sample preparation, 1 mL of stock solution (500 µg mL⁻¹) was mixed with 1 mL 1N NaOH, and the volume was made up to 10 mL with methanol. The solution was kept for 1 h at room temperature. The resultant solution of 50 µg mL⁻¹ was applied to the TLC plate.

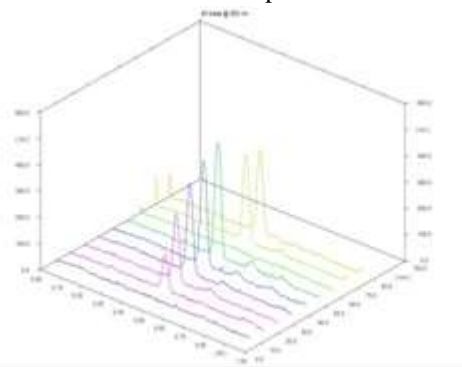


Fig.3:3D densitogram of acid hydrolysis (Track 1 methanol blank; track 2-6 standard linearity; track 7 acid blank; track 8,9 acid degradation)

Oxidative Degradation

For sample preparation, 1 mL of stock solution (500 µg mL⁻¹) was transferred to a volumetric flask then 1 mL of 3% V/V H₂O₂ was added and volume made up to 10 mL with methanol and was kept for

30 min at room temperature. The resultant solution of 50 µg mL⁻¹ was applied to the TLC plate.

Thermal Degradation

Bulk drug powder was exposed to 50°C temperature in a hot air oven for 1 h. The sample was

cooled to room temperature and then 10 mg of powder was accurately weighed and dissolved in methanol to 10 mL. A solution of strength $50 \mu\text{g mL}^{-1}$ was prepared by appropriate dilution and applied to the TLC plate.

Photolytic Degradation

The sample was exposed to UV light for not less than 200 WH m^{-2} and white fluorescent light of illumination for not less than 1.2 million lux. After exposure, 10 mg of powder was accurately weighed and dissolved in methanol to 10 mL. A solution of strength $50 \mu\text{g mL}^{-1}$ was prepared by appropriate dilution and applied to the TLC plate.

Method validation^[8]

The method was validated for the parameters like linearity, precision, accuracy, assay, robustness, LOD, LOQ.

Linearity

From the working stock solution of strength $50 \mu\text{g mL}^{-1}$, appropriate volumes like 5, 10, 15, 20, 25 μL were spotted on the TLC plate to obtain the range of 250- 1250 ng band^{-1} .

Precision

Repeatability and intermediate precision were studied by spotting six replicates of the lowest concentration from the linearity range i.e. 250 ng band^{-1} on the same day & three consecutive days.

Assay and Accuracy

For assay, the sample required i.e. $50 \mu\text{g mL}^{-1}$ was

prepared from the $1000 \mu\text{g mL}^{-1}$ sample solution. For the accuracy study, the standard addition method was used at 80%, 100%, and 120%.

Limit of Detection and Limit of Quantitation (LOD and LOQ)

The detection and quantitation limit were calculated from calibration curve. Both were calculated using the formulae, $\text{LOD} = 3.3 \text{ SD/S}$ and $\text{LOQ} = 10 \text{ SD/S}$, where SD is the standard deviation of the lowest response; S is the slope of the calibration curve.

Robustness

Five parameters were changed deliberately and slightly i.e. time from scanning to development, time of development from application, wavelength, mobile phase ratio, chamber saturation time.

V. RESULTS AND DISCUSSION

Forced Degradation Studies

In order to evaluate the stability indicating property of the developed method, forced degradation studies were carried out and optimized to 10-30 % degradation in accordance with International Conference on Harmonization (ICH) guidelines Q1A (R2). See (Table I). Stiripentol was found to be sensitive to acidic as well as alkaline hydrolytic stress, oxidative and thermal degradation conditions but no peak for degradation product was detected, even after multiple wavelength scanning which is shown in (Fig. 4)

Sr. no.	Stress condition	Concentration and time	% recovery
1	Acid hydrolysis	0.05N HCl for 20 min	76.23
2	Alkaline hydrolysis	1N NaOH for 1h	75.87
3	Oxidative degradation	3% H ₂ O ₂ for 30 min.	74.56
4	Thermal Degradation	50°C for 1h	87.36
5	UV degradation	200 WH m^{-2}	96.71
6	Fluorescence degradation	1.2 million lux	93.84

Table I: Summary of forced degradation study

Method validation

Linearity and Range

The range selected was 250-1250 ng band⁻¹ and the correlation coefficient were found 0.99 with equation of $y = 7.319x + 1765.7$. The densitogram of linearity and residual plot are shown in the (Fig. 5) and

(Fig. 6) respectively.

The linear relationship between amount spotted and peak area is confirmed by residual plot.

This residual plot without any tendency proves the linearity of calibration^[5].

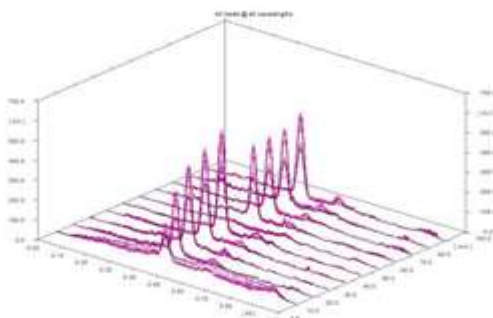


Fig. 4: 3D densitogram of multiple wavelength scanning

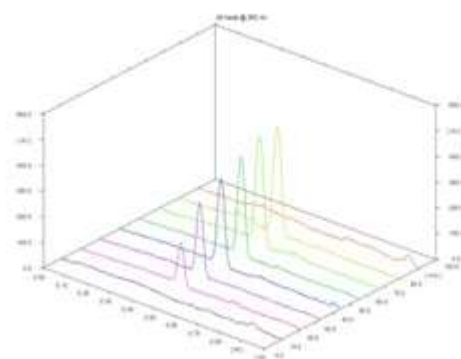


Fig. 5: 3D densitogram of pestilent linearity (250-1250 ng band⁻¹)

Assay and Accuracy

Assay was found to be 100.67% and accuracy result was shown in (Table II) and (Fig. 7).

Limit of detection (LOD) and limit of quantitation (LOQ)

LOD and LOQ were calculated by intercept method. LOD and LOQ was found to be in range i.e., 10.89 and 33.01 ng band⁻¹ respectively.

Precision

Repeatability and intermediate precision were performed and densitogram was shown in (Fig. 8). %RSD for was found to be 0.28% & 1.25% respectively.

Sr.No	Amount from marketed formulation (ng band ⁻¹)	Amount of standard added (ng band ⁻¹)	Total amount of the drug (ng band ⁻¹)	% recovery
1	500	400	900	101.34
2	500	500	1000	101.72
3	500	600	1100	100.54

Table II: % recovery (accuracy) studies

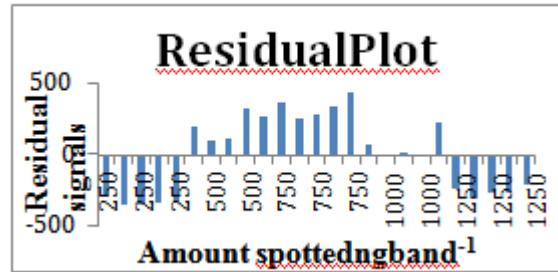


Fig.6:Residualplot

Limit of detection (LOD) and limit of quantitation (LOQ)

LOD and LOQ were calculated by intercept method. LOD and LOQ was found to be in range i.e., 10.89 and 33.01 ng band⁻¹ respectively.

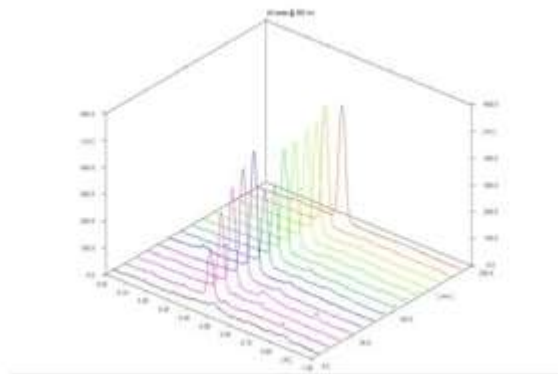


Fig.7:3D densitogram of accuracy (% recovery) (Track 1 methanol blank; track 2-6-linearity, track 7,8-assay, track 9,10-standard addition 80 %, track 11,12-standard addition 100%, track 13,14-standard addition 120 %)

Robustness

It was observed that % relative standard deviation (% RSD) NMT 1.5 %, which confirmed that the developed method was robust. For results of robustness see (Table III).

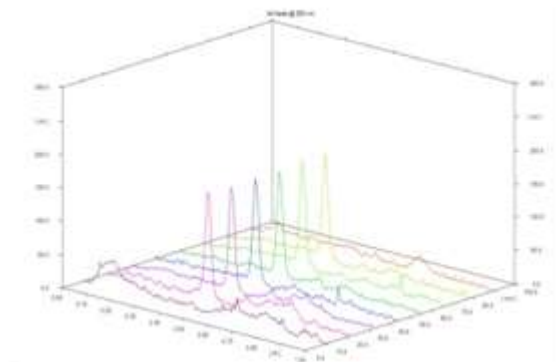


Fig.8:3D Densitogram of precision (250 ng band⁻¹)

VI. DISCUSSION

The stability-indicating HPTLC method for pestilential was developed & validated. In comparison to the results reported by Kashid S. et. al, our proposed method has an advantage of being a binary mobile phase instead of ternary. The stress conditions for degradation were optimized to obtain degradation

in the range of 10-30 % as per ICH guidelines. To the contrary, in Kashid S. et. al. study, the reported % degradation values are < 10 %. Our work is simple, well optimized & reproducible. The application of forced degradation by the proposed method could be used for monitoring extent of degradation.

Parameter	Condition	%RSD
Mobile phase composition Toulene: ethyl acetate(9.5:0.5 V/V)	9.4:0.6V/V	0.96
	9.6:0.4V/V	1.15
Saturation time(15 ±5min)	10min	0.89
	20min	1.68
Time from application to development	Immediate	1.16
	After 2h	1.16
Time from development to scanning	Immediate	1.27
	After 2h	1.27
Change in wavelength(262±2 nm)	260nm	1.26
	264nm	1.00

Table III: Robustness studies

VII. CONCLUSION

The developed HPTLC method for estimation of pestilential was found to be accurate, precise, specific. The method was validated as per guidelines of ICH Q2R1. Stiripentol was found to be sensitive to acidic as well as alkaline hydrolytic and oxidative and thermal degradation conditions but no degradation product peak was detected. The developed chromatographic method may be used for routine analysis of pestilential.

REFERENCE

[1]. Chiron C., Helias M., Kaminska A.,

Laroche C., de Toffol B., Dulac O., Nabbout R. and An I.: D o children with dravets syndrome continue to benefit from pestilential for long through adulthood?, *Epilepsia*. 2018, 59(9), 1705-1717.
 [2]. Chiron C.: Stiripentol for the treatment of seizures associated with dravets syndrome, *Expert Rev Neurother*, 2019, 19(4), 301-310
 [3]. Darwish HW., Abdelhameed AS., Attia MI., Bakheit AH., Khalil NY. and Al-Majed AA.: A stability-indicating HPLC-DAD method for determination of

- pestilential: development, validation, kinetics, structure elucidation and application to commercial dosage form, *J. Anal. Methods Chem*, 2014, 2014.
- [4]. European Medicines Agency, Stiripentol: Scientific Discussion, http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_Scientific_Discussion/human/000664/WC500036521.pdf.
- [5]. Ferenczi-Fodor K., Renger B., and Vegh Z.: The frustrated reviewer – recurrent failures in manuscripts describing validation of quantitative TLC/HPTLC procedures for analysis of pharmaceuticals, *J. Planar Chromatogr.*, 2010, 23(3), 173–179.
- [6]. Fisher J.: The effects of pestilential on GABAA receptors, *Epilepsia*, 2011, 52(2), 76–78.
- [7]. ICH. Q1A (R2) Stability Testing of New Drug Substances and Products. Geneva: ICH: 2003.
- [8]. ICH. Q2 (R1) Validation of analytical procedures: Text and Methodology. Geneva: ICH: 2005.
- [9]. ICH. Q1B Stability testing: Photostability Testing of New Drug Substances and Products. Geneva: ICH: 2003.
- [10]. Jacob S. and Nair A.: An updated overview on therapeutic drug monitoring of recent antiepileptic drugs, *Drugs RD*, 2016, 16(4), 303- 316.
- [11]. Kashid SK., Tapkir A. and Choudhari P.: Analytical method development and validation for stability indicating HPTLC method for assay of pestilential in bulk and dosage form, *J. Pharm. Sci. Res.*, 2020, 3(4), 26-30.
- [12]. May T., Boor R., Mayer T., Jürgens U., Rambeck B., Holert N., Korn-Merker E, and Brandt C.: Concentrations of pestilential in children and adults with epilepsy: the influence of dose, age, and comedication, *Ther Drug Monit*, 2012, 34(4), 390-397.
- [13]. Nickels K C. and Wirrell E C.: Stiripentol in the management of epilepsy, *CNS Drugs*, 2017, 31(5), 405-416.
- [14]. O’Neil M., *The Merck Index: An Encyclopedia of Chemicals, Drugs, and Biologics*, 15th(Ed.), The Royal Society of Chemistry, New Jersey 2013, pp.1631.
- [15]. Peigne S., Chhun S., Tod M., Rey E., Rodrigues C., Chiron C., Pons G. and Jullien V.: Population pharmacokinetics of pestilential in paediatric patients with dravet syndrome treated with pestilential, valproate, and clobazam combination therapy, *Clin Pharmacokinet*, 2018, 57(6), 739-748.
- [16]. Poisson M., Hugué F., Savattier A., Bakri-Logeais F. and Narcisse G.: A new type of anticonvulsant, pestilential. pharmacological profile and neurochemical study, *Arzneimittelforschung*, 1984, 34(2), 199-204.
- [17]. Takahashi R., Imai K., Yamamoto Y., Takahashi Y., Hamano SI. And Yoshida H.: Determination of pestilential in plasma by high-performance liquid chromatography with fluorescence detection, *Iryo Yakugaku*, 2015, 41(9), 643-50.
- [18]. Wirrell E., Laux L., Franz D., Sullivan J., Saneto R., Morse R., Devinsky O., Chugani H., Hernandez A., Hamiwka L., Mikati M., Valencia I., LeGuern M., Chancharme L. and DeMenezes M.: Stiripentol in dravet syndrome: Results of a retrospective U.S. study, *Epilepsia*, 2013, 54(9), 1595-1604.
- [19]. Yıldız E., Ozkan M., Uzunhan T., Bektaş G., Tatlı B., Aydın N., Çalışkan M. and Özmen M.: Efficacy of pestilential and the clinical outcome in dravet syndrome, *J Child Neurol*, 2019, 34(1), 33-37.