

A Review of Use of Amphotericin B in the Treatment of Mucormycosis

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ABSTRACT

As the spores of the fungus are inhaled, it affects the oral and nasal cavity, which is the most typical type of mucormycosis. Due to the rarity of the illness, the variety of offending Mucorales, the variability of the hosts and locations of infection, and the lack of reliable data for the treatment of mucormycosis, hence no prospective clinical trial exists. Except for a few isolates of Cunninghamella and Apophysomyces, Amphotericin B is the medication with the highest activity. Amphotericin B in a lipid formulation is recommended as the first-line treatment for mucormycosis in both the ESCMID/ECMM guidelines and the 2016 recommendations from the European Conference on Infections in Leukaemia. The efficacy of Amphotericin B has been shown in both laboratory and clinical studies. In order to avoid the severe side effects such as pyrexia and chills, treatment with amphotericin B is usually started in small doses which are gradually increased during a period of 3-4 days.

KEY WORDS: Mucormycosis, Amphotericin B, Antifungal

I. INTRODUCTION

The "Mucoromycetes" genus of fungus is responsible for the dangerous but uncommon fungal illness known as mucormycosis.⁷ The infection, previously termed zygomycosis, is caused by mucoralean fungi, which have collectively also been called Mucoromycetes. However, we prefer to use the name of the order, i.e. Mucorales.² Fungi belonging to the order Mucorales are distributed into six families, all of which can cause cutaneous and deep infections. These organisms are ubiquitous in nature as they can be found in decomposing or rotting organic substrates like vegetable matters and wood, soil and dust.

Mucorales are multiplying rapidly and they are releasing immensely large number of airborne spores. Humans are exposed to these spores on a daily basis, but the intact immune system does not allow development of infection in the human body. Thus, with the exception of victims of major diseases, the disease affects mainly immunocompromised patients with severe underlying diseases, such as hematologic malignancies (HM), solid organ (SOT) or hematopoietic stem cell transplantation (HSCT), uncontrolled diabetes mellitus, severe trauma, or burns.¹

In case of rise in Covid 19, Mucormycosis was seen as an after-effect post Covid 19 treatment. Suspected reason of this occurrence was due to suppressed immunity. Mucormycosis is also known as Black fungus. Mucormycosis can be categorised into one of six types based on anatomic localization: (1) rhino-orbital-cerebral mucormycosis (ROCM), (2) pulmonary, (3) cutaneous, (4) gastrointestinal (GI), (5) diffuse, and (6) mucormycosis of uncommon sites.

The most frequent clinical manifestation of ROCM in diabetics is lung involvement, although it is less prevalent. In contrast, among organ transplant recipients or patients with haematological malignancies (HemeM), pulmonary and disseminated diseases are most common. Mucormycosis can progress rapidly, and delay in initiation of treatment by even a few days markedly worsens outcomes.² Although mucormycosis represents a rare disease, its consequences are devastating, since it is associated with unacceptably high mortality rates, ranging from 20–50% if localised, up to 70–90% in cases of disseminated disease.¹

More often than any other family, mucormycosis patients are isolated from species that belong to the family Mucoraceae. *Rhizopus oryzae* (*Rhizopus arrhizus*) is by far the most prevalent infection-causing agent in the Mucoraceae family. *Rhizopus microsporus* var. and

other Mucoraceae species that are less commonly isolated cause an identical range of illnesses. Mucor species, Absidia corymbifera, Rhizopodiformis, Apophysomyces elegans, and Rhizomucor pusillus. As a result of Cunninghamella spp. infection, mucormycosis instances have also been observed to be rising (in the Cunninghamellaceae family). To date, rare case reports have demonstrated the ability of species belonging to the remaining four families to cause mucormycosis.³The mortality of mucormycosis remains high. Surgical intervention is combined with antifungal medications for treatment. Isavuconazole is the sole novel medicine having efficacy against Mucorales, although it doesn't seem to have any discernible benefits over Posaconazole or Amphotericin B-based medications, which have traditionally been used as first-line therapies. Since early diagnosis of mucormycosis improves survival, several researchers are working to develop novel techniques for making the diagnosis of the disease earlier. This review will outline the various fields of research targeting diagnosis, as well as the modalities used either as primary or as adjunctive treatment of this frequently lethal disease.⁵

EPIDEMIOLOGY

The incidence of mucormycosis is increasing worldwide, but it is rising particularly quickly among people with uncontrolled diabetes mellitus in China and India. A recent review of 851 cases from January 2000 to January 2017 shows a different picture, showing that the disease burden is

higher in Europe than in Asia. They reported 34% in Europe, followed by Asia (31%) and North or South America (28%), Africa (3%), Australia, and New Zealand (3%), and then Asia (31%) and Africa (3%). The contrary findings may be the result of underreporting from Asian nations during this time. In reality, a rising number of cases are reported from India.⁷ Without population-based estimates, it is difficult to determine the exact incidence and prevalence of mucormycosis in the Indian population. A 10-year study from Southern India (Tamil Nadu) showed an annual incidence of 18.4 cases per year during 2005–2015 [17]. Another study from Tamil Nadu reported 9.5 cases per year during 2015–2019 [18].⁷

A new immediate threat has developed as a problem for India as it works to maintain stability in the current environment, and it is known as mucormycosis related with coronavirus illness. The incidence of mucormycosis, sometimes known as the "black fungus," increased more quickly in India during the second COVID-19 wave compared to the first wave, 14872 cases as of May, 2021.⁸

CLINICAL PRESENTATION

Mucormycosis almost typically affects patients with compromised host defence and/or elevated blood iron levels, while a few instances have also been observed in individuals who appeared to be healthy. If treatment with a combination of surgical debridement and antifungal medication is not started right once, the infection usually progresses relentlessly and kills its victim.³

Relationship between the location of infection and the predisposing condition

Predisposing condition	Predominant infection location
Diabetic ketoacidosis	Rhinocerebral
Neutropenia	Pulmonary and disseminated
Corticosteroids	Pulmonary, disseminated, or rhinocerebral
Deferoxamine	Disseminated
Malnutrition	Gastrointestinal
Trauma, catheter/injection site, skin maceration	Cutaneous/ Subcutaneous

PATHOPHYSIOLOGY

In addition to being prevalent in the environment, fungus may also be found in a healthy person's nasal and oral mucosa. As the spores of the fungus are inhaled, it affects the oral and nasal cavity, which is the most typical type of mucormycosis.¹³This particular kind of mucormycosis begins as an infection in the nasal turbinates and then rapidly spread to affect the sinuses, palate, orbit, and brain.¹⁴

In most cases, the infection is dangerous and results in death unless treatment with a combination of surgical debridement and antifungal therapy is initiated promptly.

Fungal-Endothelial Interactions

Mucormycosis infections are characterized by extensive angioinvasion which causes vascular thrombosis and ensuing tissue necrosis.¹⁵

Leukocyte and antifungal agent distribution to infection foci can be hindered by ischemic necrosis of infected tissues. This angioinvasion probably contributes to the capacity of the organism to hematogenously spread to additional target organs. As a result, a crucial stage in *R. oryzae*'s pathogenetic approach is probably damage to and penetration through endothelial cells or the extracellular matrix proteins lining blood arteries.¹⁶

In vitro, laminin and type IV collagen are two subendothelial matrix proteins that *R. oryzae* spores can cling to, but not germlings (i.e., pregerminated spores).¹⁷

Host Defences

In a healthy individual, phagocytic leukocytes assist the immune system in eliminating the spores. In contrast, the spores change into hyphae in immune-compromised individuals; but since the white blood cells have lower efficacy on the hyphae, the fungi proliferates with greater ease.

Mucorales are killed by normal hosts' mononuclear and polymorphonuclear phagocytes through the production of oxidative metabolites and the cationic peptides defensins.¹⁸

Patients who are neutropenic and have dysfunctional phagocytes are at increased risk of developing mucormycosis.¹⁹

Hyperglycemia and acidosis are known to impair the capacity of phagocytes to approach and eradicate the organisms through both oxidative and nonoxidative pathways.²⁰

Therefore, understanding the mechanisms by which these events occur may result in novel approaches to prevent and/or treat mucormycosis.

II. MANAGEMENT AND TREATMENT

Rapid identification, reversal of the underlying predisposing conditions, adequate surgical debridement of affected tissue, and suitable antifungal medication are essential for eliminating mucormycosis. Early detection is crucial because tiny, isolated lesions may frequently be surgically removed before they spread or get involved with crucial tissues.³ Early detection and therapeutic action may lessen the need for significant surgery and subsequent deformity, avoid progressive tissue invasion and its consequences, and increase survival.¹

Patients with uncontrolled diabetes who are suspected of having mucormycosis must immediately address any metabolic abnormalities.

In this regard, experimental data shows that the administration of sodium bicarbonate (in combination with insulin) to correct ketoacidosis, regardless of how severe the acidosis is, may be associated with a better prognosis for the illness since Mucorales' capacity to infiltrate host tissues is reversed.⁵ Due to the extensive tissue necrosis that results from mucormycosis and may not be stopped by eliminating the organism, surgery is required. Infected and necrotic tissue should be surgically debrided as soon as possible.³ Due to the rarity of the illness, the variety of offending Mucorales, the variability of the hosts and locations of infection, and the lack of reliable data for the treatment of mucormycosis, hence no prospective clinical trial exists.

The course of treatment for mucormycosis are Antifungal agents either individually or in combination. In vitro tests show that mucoraceous fungus are resistant to many antifungals, including voriconazole. Except for a few isolates of *Cunninghamella* and *Apophysomyces*, Amphotericin B is the medication with the highest activity.⁵ Amphotericin B in a lipid formulation is recommended as the first-line treatment for mucormycosis in both the ESCMID/ECMM guidelines and the 2016 recommendations from the European Conference on Infections in Leukemia (ECIL-6).⁵ Along with Amphotericin B (AMB) and its lipid formulations, Isavuconazole is also used as well as more recently, have been investigated. Posaconazole, on the other hand, has mostly been researched as a salvage treatment. Preclinical in vitro/in vivo studies indicating action against Mucorales and limited clinical data are the foundations for these drugs' effectiveness. However, it should be noted that none of these medicines have verified minimum inhibitory concentration (MIC) breakpoints.¹ These medications are given by vein (Amphotericin B, Isavuconazole and Posaconazole) and orally (Isavuconazole and Posaconazole). AMB 5 mg/kg/d or isavuconazole is recommended for individuals for whom amphotericin B is unsuitable (loading dose 372 mg q 8 h for 6 doses IV/oral, then 372 mg QD oral or IV) 12- 24 h following previous dose).¹ Posaconazole has mostly been researched as a salvage medication.¹

AMPHOTERICIN B

Amphotericin B, often known as AMB, is thought to be the best medicine for treating Mucormycosis on a long-term basis. The efficacy of Amphotericin B has been shown in both

laboratory (in vitro and in vivo) and clinical studies.⁹The dose of amphotericin B deoxycholate that has been advised has been 1 to 1.5 mg/kg/day, which has a very high toxicity rate.³The usage is constrained by a dose-dependent nephrotoxicity that manifests as tubular dysfunction and a decrease in glomerular filtration rate. Amphotericin B nephrotoxicity is not a benign consequence, and its avoidance is crucial. An raised creatinine associated with amphotericin B is not only a marker for renal failure but also connected to a significant risk for the need of haemodialysis and a higher death rate.¹¹

Newer formulations like, Lipid versions of AMB (LFABs) can be given at larger doses for a longer period of time than AMB without harming the kidneys as they have a much lower nephrotoxic effect. At regimens as high as 10 to 15 mg/kg/day, LAMB is associated with significantly less renal toxicity compared to amphotericin B deoxycholate.²³ Amphotericin B lipid complex (ABLC), used as salvage treatment for mucormycosis, had a 71% success rate in one research.¹²Amphotericin B, a broad-spectrum polyene antifungal drug, is available in a lipid-associated formulation called liposomal amphotericin B. It is approved for the treatment of invasive fungal infections in many nations across the world and is effective against clinically relevant yeasts and moulds including *Candida* spp., *Aspergillus* spp., and filamentous moulds like *Zygomycetes*.¹⁰ The development of less toxic, lipid-based polyene formulations in the periods between late 1980s and early 1990s may be considered a breakthrough in antifungal chemotherapy, particularly for patients with invasive aspergillosis and mucormycosis.²¹

It was developed to improve the tolerability profile of amphotericin B deoxycholate, which, although being linked to nephrotoxicity and infusion-related complications, was for a long time regarded as the gold standard of antifungal therapy.¹⁰

PHARMACOKINETICS

The absorption amphotericin B of minimal from the digestive tracts, For this reason, amphotericin B has to be injected intravenously. During IV therapy, 90 % of Amphotericin B disappears from the blood although only consistently low levels are found in body fluids other than serum. Since the dose must be gradually raised, amphotericin B takes some time to reach fungistatic levels. The half-life of amphotericin B is

long (days); amphotericin B, probably owing to the high protein binding, is not haemodialysable.²⁰The highest tolerated dose (1 0-1 5 mg/kg body weight or 50-100 mg) was administered daily, but it gradually became apparent that a double dose on every other day was more effective and better tolerated.

Daily administration of the maximum tolerable dose (50-100 mg or 1 0-1 5 mg/kg body weight) progressively gave way to the conclusion that every other day administration of a double dose was both more efficient and better tolerated.²⁰

In order to avoid the severe side effects such as pyrexia and chills, treatment with amphotericin B is usually started in small doses which are gradually increased during a period of 3-4 days. The disadvantage of this is that the production of therapeutically active serum levels is correspondingly delayed.²⁰

The liposomal formulation of Amphotericin B was developed to improve the tolerability of intravenous AmB, while optimizing its clinical efficacy.²¹

Both formulations have triphasic plasma profiles with extended terminal half-lives (Liposomal AMB, 152 ± 116 h; AMB, 127 ± 30 h), but plasma concentrations were higher ($P < 0.01$) after administration of liposomal AMB (maximum concentration of drug in serum [C_{max}], 22.9 ± 10 g/ml) than those of AMB (C_{max}, 1.4 ± 0.2 g/ml).¹⁹

Liposomal AMB had a volume of the central compartment (50 ± 19 ml/kg) that was comparable to plasma and a volume of distribution at steady state (V_{ss}) (774 ± 550 ml/kg) V_{ss} that was less than that of AMB ($1,807 \pm 239$ ml/kg) ($P < 0.01$).¹⁹

However, renal and faecal clearances of Liposomal AMB were 10-fold lower than those of AMB ($P < 0.01$), despite total clearances being similar (about 10 ml hr⁻¹ kg⁻¹).Two-thirds of the AMB-DOC was excreted unchanged in the urine (20.6%) and feces (42.5%) with >90% accounted for in mass balance, suggesting that metabolism plays at most a minor role in AMB elimination.¹⁹

PHARMACODYNAMICS

Starting 12 hours after infection, L-AMB or ABLC are given intravenously every day at dosages of 1, 5, or 10 mg/kg of body weight for 5 days. Both L-AMB and ABLC were effective at reducing the lung fungal load at a dosage of 10 mg/kg/day and produced lung tissue concentrations that were higher by 72 hours than the isolate mean fungicidal concentration (MFC) of 8 g/ml.²⁴

Compared to amphotericin B deoxycholate, LAmB has lower potency per mg. The most glaring example of differences in exposure-response relationships comes from an in vitro model of the human alveolus, where the effective dosage (ED50) for LAmB and DAmB was 1.03 and 0.12 mg/L, respectively.²³

At 24 and 72 hours after receiving a dose of 5 mg/kg/day of ABLC, the AMB lung concentrations in the ABLC-treated animals were higher than those in the L-AMB-treated animals by a significant margin (6.64 and 1.44 g/g, respectively; $P=0.013$) and 72 h (7.49 and 1.03 g/g, respectively; $P=0.005$). These higher concentrations were also associated with improved fungal clearance as determined by quantitative real-time PCR (mean conidial equivalent of fungal DNA per lung 4.44 ± 0.44 and 6.57 ± 0.74 log₁₀, respectively; $P < 0.001$). The inhibition of growth in the lung required tissue concentrations that were close to the MFC for the infecting isolate (50% effective concentration, 8.19 g/g [95% confidence interval, 2.81 to 18.1 g/g]), according to analysis of the AMB tissue concentration-response correlations. The survival rates in the mice given 10 mg/kg/day treatments of L-AMB and ABLC were comparable.²⁴

ADVERSE DRUG REACTION

The most common ADR of Amphotericin B is Renal failure and Metabolic like Hypokalaemia with reporting more than 10% of the total reported cases (10 or >).

Nephrotoxicity

The incidence of Amphotericin B nephrotoxicity is very high and there is a reason to be cautious. Elevated creatinine associated with Amphotericin B is not only a marker for Renal dysfunction, but it is also linked to an increase in hospital costs and a substantial because to the use of hemodiafiltration and a greater mortality rate. Therefore, Amphotericin B nephrotoxicity is not a benign complication and its prevention is essential.²² Of all the currently available lipid formulations of amphotericin B, LAmB consistently has the lowest nephrotoxicity. When compared to amphotericin B deoxycholate, regimens as high as 10 to 15 mg/kg/day of LAmB are associated with much reduced renal damage in infected animals.²³

III. CONCLUSIONS

Amphotericin B is still used as the novel drug for treatment of several fungal infections, one

of them being Mucormycosis. Even though newer agents like Isavuconazole and Posaconazole have been introduced Amphotericin B is still the drug of choice. Amphotericin B has newer formulations like Lipid based Amphotericin which are broadly used. The most common ADR with Amphotericin B is nephrotoxicity. Amphotericin B has shown renal damage in patients who are treated with them. Nephrotoxicity not only needs to be monitored but also made economical. It is also shown to cause hypokalaemia. More information needs to be collected regarding the pharmacology, pharmacokinetics and pharmacodynamics of the drug.

REFERENCE

- [1]. Sipsas NV, Gamaletsou MN, Anastasopoulou A, Kontoyiannis DP. Therapy of mucormycosis. *Journal of Fungi*. 2018 Jul 31;4(3):90.
- [2]. Cornely O, Arian-Akdagli SE, Dannaoui E, Groll AH, Lagrou K, Chakrabarti A, Lanternier F, Pagano LI, Skiada A, Akova M, Arendrup MC. ESCMID and ECMM joint clinical guidelines for the diagnosis and management of mucormycosis 2013. *Clinical Microbiology and Infection*. 2014 Apr;20:5-26.
- [3]. Spellberg B, Edwards Jr J, Ibrahim A. Novel perspectives on mucormycosis: pathophysiology, presentation, and management. *Clinical microbiology reviews*. 2005 Jul;18(3):556-69.
- [4]. Pilmis B, Alanio A, Lortholary O, Lanternier F. Recent advances in the understanding and management of mucormycosis. *F1000Research*. 2018;7.
- [5]. Skiada A, Lass-Floerl C, Klimko N, Ibrahim A, Roilides E, Petrikkos G. Challenges in the diagnosis and treatment of mucormycosis. *Medical mycology*. 2018 Apr 1;56(suppl_1):S93-101.
- [6]. Reid G, Lynch III JP, Fishbein MC, Clark NM. Mucormycosis. In *Seminars in respiratory and critical care medicine 2020 Feb* (Vol. 41, No. 01, pp. 099-114). Thieme Medical Publishers.
- [7]. Prakash H, Chakrabarti A. Global epidemiology of mucormycosis. *Journal of Fungi*. 2019 Mar 21;5(1):26.
- [8]. Prakash H, Chakrabarti A. Epidemiology of mucormycosis in India. *Microorganisms*. 2021 Mar 4;9(3):523.

- [9]. Raut A, Huy NT. Rising incidence of mucormycosis in patients with COVID-19: another challenge for India amidst the second wave?. *The Lancet Respiratory Medicine*. 2021 Aug 1;9(8):e77.
- [10]. Sipsas NV, Gamaletsou MN, Anastasopoulou A, Kontoyiannis DP. Therapy of mucormycosis. *Journal of Fungi*. 2018 Jul 31;4(3):90.
- [11]. Moen MD, Lyseng-Williamson KA, Scott LJ. Liposomal amphotericin B: a review of its use as empirical therapy in febrile neutropenia and in the treatment of invasive fungal infections. *Drugs*. 2009 Feb;69:361-92.
- [12]. Deray G, Mercadal L, Bagnis C. Nephrotoxicity of amphotericin B. *Nephrologie*. 2002 Jan 1;23(3):119-22.
- [13]. Goldstein EJ, Spellberg B, Walsh TJ, Kontoyiannis DP, Edwards Jr J, Ibrahim AS. Recent advances in the management of mucormycosis: from bench to bedside. *Clinical Infectious Diseases*. 2009 Jun 15;48(12):1743-51.
- [14]. Veisi A, Bagheri A, Eshaghi M, Rikhtehgar MH, Rezaei Kanavi M, Farjad R. Rhino-orbital mucormycosis during steroid therapy in COVID-19 patients: A case report. *European journal of ophthalmology*. 2022 Jul;32(4):NP11-6.
- [15]. Kumar A, Sidhu J, Goyal A, Tsao JW, Svercauski J. *StatPearls* [Internet].
- [16]. Ibrahim AS, Spellberg B, Walsh TJ, Kontoyiannis DP. Pathogenesis of mucormycosis. *Clinical infectious diseases*. 2012 Feb 1;54(suppl_1):S16-22.
- [17]. Diamond RD, Haudenschild CC, Erickson 3rd NF. Monocyte-mediated damage to *Rhizopus oryzae* hyphae in vitro. *Infection and immunity*. 1982 Oct;38(1):292-7.
- [18]. Chinn RY, Diamond RD. Generation of chemotactic factors by *Rhizopus oryzae* in the presence and absence of serum: relationship to hyphal damage mediated by human neutrophils and effects of hyperglycemia and ketoacidosis. *Infection and immunity*. 1982 Dec;38(3):1123-9.
- [19]. Bekersky I, Fielding RM, Dressler DE, Lee JW, Buell DN, Walsh TJ. Pharmacokinetics, excretion, and mass balance of liposomal amphotericin B (AmBisome) and amphotericin B deoxycholate in humans. *Antimicrobial agents and chemotherapy*. 2002 Mar;46(3):828-33.
- [20]. Polak AN. Pharmacokinetics of amphotericin B and flucytosine. *Postgraduate Medical Journal*. 1979 Sep 1;55(647):667-70.
- [21]. Groll AH, Rijnders BJ, Walsh TJ, Adler-Moore J, Lewis RE, Brüggemann RJ. Clinical pharmacokinetics, pharmacodynamics, safety and efficacy of liposomal amphotericin B. *Clinical Infectious Diseases*. 2019 May 2;68(Supplement_4):S260-74.
- [22]. Deray G. Amphotericin B nephrotoxicity. *Journal of antimicrobial chemotherapy*. 2002 Jan 1;49(suppl_1):37-41.
- [23]. Stone NR, Bicanic T, Salim R, Hope W. Liposomal amphotericin B (AmBisome®): a review of the pharmacokinetics, pharmacodynamics, clinical experience and future directions. *Drugs*. 2016 Mar;76:485-500.
- [24]. Lewis RE, Albert ND, Liao G, Hou J, Prince RA, Kontoyiannis DP. Comparative pharmacodynamics of amphotericin B lipid complex and liposomal amphotericin B in a murine model of pulmonary mucormycosis. *Antimicrobial agents and chemotherapy*. 2010 Mar;54(3):1298-304.